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(54) Title: **LIVE CELL PROCEDURES TO IDENTIFY COMPOUNDS MODULATING INTRACELLULAR DISTRIBUTION
OF PHOSPHODIESTERASE (PDE) ENZYMES**

(57) Abstract: An alternative therapeutic approach for PDE4 inhibition is disclosed. PDE4 dislocators, will remove the PDE4 away from the native location in the cell and thereby increase the concentration of cAMP in this location. By dislocating the PDE4, and thereby not acting directly on the catalytic, among phosphodiesterase inhibitors, well conserved site, the compound will act e.g. at the binding domain of the PDE4, thereby providing isoform-specific 'inhibitors' of PDE4. The dislocation of PDE4s are visualised with fusions to GFP. The native location is induced by treatment with Rolipram.

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Live cell procedures to identify compounds modulating intracellular distribution of phosphodiesterases (PDE) enzymes

Background

- 5 Cyclic AMP is a ubiquitous second messenger. It is generated through the action of adenylyl cyclase and serves to transduce the action of many hormones, neurotransmitters and other cellular effectors. cAMP exerts its effects on cells through its ability to bind to specific intracellular regulatory proteins. These are protein kinase A (PKA), cyclic nucleotide gated ion channels (CNG channels) and cyclic AMP stimulated GTPase
- 10 exchange factors (cAMP-GEFs, EPACs). Such effectors allow cAMP to regulate cellular processes in a cell-type specific fashion. Thus elevated cAMP levels can for example affect CNS function (e.g. depression), cardiovascular function, inflammatory cells/immune system, cell adhesion and metabolic processes. These actions, however, depend upon cAMP being elevated not only in particular cell types but also in particular intracellular
- 15 locations (Houslay and Milligan 1997).

- The only way to degrade cAMP is through the action of cyclic nucleotide phosphodiesterases (PDEs) (Conti and Jin 1999). These hydrolyse 3',5' cyclic adenosine monophosphate (cAMP) to 5'-adenosine monophosphate (AMP). It is now well-
- 20 recognised that a large multi-gene family encode PDEs. However, only certain of these enzymes are capable of hydrolysing cAMP. These are members of the PDE1, PDE2, PDE3, PDE4, PDE7, PDE8 and PDE11 families. Selective inhibitors have been generated against certain of these families, e.g. PDE3 and PDE4 enzymes. These inhibitors are targeted to the enzyme catalytic unit, being identified through screens which looked for
- 25 inhibition of cAMP hydrolysis. Such inhibitors thus display competitive kinetics of inhibition. PDE3 and PDE4 selective inhibitors have, for example, been shown to generate clearly distinct pharmacological responses. For example, the PDE3 inhibitor, milrinone serves as positive inotropic agent and increase the force of contraction of the heart, whereas the PDE4 inhibitor, Rolipram does not (Manganiello et al. 1995). In
- 30 contrast to this, PDE4 inhibitors (Rolipram, Ariflo®) can inhibit the action of many cells of haematopoietic origin that are associated with inflammatory responses and may also exert antidepressant effects (Rolipram), whereas PDE3 inhibitors (milrinone, cilostamide) do not. Whilst PDE isoenzymes show cell-type specific patterns of expression, PDE3 and

PDE4 enzymes are often found in many types of cells, including cardiac myocytes and inflammatory cells. Thus the ability of PDE isoenzyme selective inhibitors to exert very different effects on a particular cell type is not necessarily due to lack of expression of one or other PDE isoenzyme in that cell type. Of course, in certain instances apparent
5 selectivity can arise due to very different levels of PDE3 and PDE4 enzyme activity in cells where different actions of PDE isoenzyme-selective inhibitors have been noted. However, in many instances this is simply not the case. To explain this apparent dichotomy the concept of compartmentalised cAMP signalling has arisen. This envisages that cAMP is not uniformly distributed through the cell interior. Indeed, there is direct
10 evidence which demonstrates this (Hempel et al. 1996).

As cAMP is only degraded by cAMP-PDEs in cells, then the attenuation of their activity can be expected to lead to an increase in cAMP levels and the triggering of a cellular response. As PDE3 and PDE4 enzymes are localised to discrete intracellular sites then
15 they can be expected to control 'localised pools' of cAMP that, in turn, may control the activity of restricted PKA-RII/EPAC/CNG channels. Traditional approaches have focussed entirely on developing active site-directed selective PDE inhibitors to provide novel therapeutic agents (Souness and Rao 1997). However, the realisation (Houslay et al. 1998) that specific PDE4 isoforms show precise intracellular targeting offers a radically
20 new means of altering PDE functioning in intact cells and generating a novel class of therapeutic agents affecting PDE functioning. This exploits an ability to disrupt the intracellular targeting of specific PDE4 isoenzymes and thus to remove the target isoenzyme from its functionally relevant intracellular compartment. Such a relocation would be expected to elevate intracellular cAMP levels in a particular subcellular location
25 ('compartment') and lead to activation of PKA/EPAC/CNG channels in the vicinity. This offers the potential of generating isoform-specific 'inhibitors' that rather than acting on the enzyme catalytic unit serve to displace the target PDE4 isoforms from its functional relevant (anchor) site within a cell. This may involve the release of the enzyme into the cytosol where it will be grossly diluted or re-targeted.

30

The PDE4 enzyme family is a family where active-site directed inhibitors have anti-depressant and anti-inflammatory action. PDE4 isoforms show distinct and cell-type specific patterns of expression (Houslay et al. 1998). In addition, within cells PDE4 isoforms also exhibit highly specific intracellular distribution patterns. Thus, for example,
35 the PDE4A1 isoform appears to be expressed only in within certain brain regions

(Houslay et al. 1998). Indeed, when PDE4A1 is expressed in various cell types it shows a distinctive pattern of intracellular distribution, implying targeted association (Pooley et al. 1997).

- 5 PDE4 enzymes encoded by four distinct genes (Houslay et al. 1998), specifically hydrolyse cAMP. The large family of PDE4 isoforms arise through the use of alternative promoters and alternative mRNA splicing.

The recruitment or re-localisation of proteins plays a major role in many key signalling systems. This is evident in (i) the activation of protein kinase C (PKC), where recruitment to the plasma membrane is an inherent part of the activation process of this enzyme; (ii) the activation of p42/44 MAP Kinase depends on multiple proteins whose transfer from cytosolic to membrane compartments has a central role and (iii) the cAMP-driven relocalisation of rap1 is crucial to its activation. Thus there is a need for the ability to
10 detect the localisation and any re-localisation of PDE4 enzymes in intact, living cells, as such detection is expected to provide a novel and innovative means for identifying new therapeutic agents.

Present screening assays for compounds interfering with the activity of PDE enzymes are
20 based upon variations of methods which assess PDE catalytic activity; namely the ability of the enzyme to hydrolyse 3'5' cyclic adenosine monophosphate (cAMP) to 5'-AMP. This is usually performed in multiwell format using detection of cAMP hydrolysis using proximity-based radionuclide assays. Such screens detect compounds that alter catalytic activity. To date these have identified compounds that bind to the catalytic site as
25 competitive inhibitors. Thus all compounds reported on to date bind to the catalytic site and thus compete with the substrate cAMP for binding. The enzymes used in these screens often are cell extracts of endogenous PDE4 enzymes that have been partially purified to remove non-PDE4 enzymes. These suffer from the fact that they may be contaminated by as yet unknown PDE species and that they will contain mixtures of PDE4
30 isoforms. An alternative approach has been to use recombinant enzymes in screens where expression has been done in various of cell lines / systems such as sf9 cells, *S. cerevisiae*, *E. coli* and transfected mammalian cell lines. This allows for isoform specific analyses to be done. However, as the catalytic unit of PDE4 enzymes is identical for isoforms within each PDE4 subfamily then it is near impossible to conceive that an
35 isoform selective inhibitor could be identified through such analyses. In addition, the

catalytic subunit is highly conserved within the enzymes of the 4 gene PDE4 family itself. This means that it is likely to be extremely difficult, although not inconceivable, to obtain inhibitors that are highly selective between each of the four families. To date, the best selectivity reported is that for Ariflo® which shows some 8 to 10-fold selectivity for the PDE4D family over enzymes from the other three PDE4 families (Barnette et al. 1998). There is thus a need to develop strategies that will allow for the identification of compounds that serve as isoform-specific 'inhibitors'. There is therefore a need for procedures that can (i) allow for the rapid screening of agents that disrupt the targeting of PDE4 isoforms in living cells and (ii) identify conformationally distinct forms of PDE4 living cells. Procedures that allow for these aims to be achieved can be expected to lead to the development of novel therapeutics. In addition they will lead to the provision of diagnostic aids to identify compounds exerting conformationally distinct effects on PDE4 enzymes and thus as being of use in compound development to either screen for beneficial e.g. (anti-inflammatory, anti-depressant) or against adverse effects (e.g. nausea, vomiting, arteritis).

Summary of the invention

The examples in the application disclose, for the first time, that the selective PDE4 inhibitor Rolipram affects the physical properties and behaviour of PDE4A4 such that the general cytoplasmic distribution of PDE4A4 in most cells gradually changes to one
5 consisting of concentrations of PDE4A4 located at several distinct spots within the cytoplasm (example 3). Pre-treatment of the cells with cycloheximide, a protein synthesis inhibitor, prevents formation of spots induced by Rolipram, indicating that protein synthesis is a necessary part of the observed spot formation. Once spots have formed, removal of Rolipram results in their rapid dissolution. However, replacement of Rolipram
10 causes the spots to rapidly reform. This is the first evidence that binding of Rolipram induce changes in distribution.

Additionally, example 15 discloses, also for the first time, that Rolipram affects the physical properties and behaviour of PDE4A1 but in a way that is very different to the
15 effects this compound has on the behaviour of PDE4A4. PDE4A1 accumulates as small perinuclear spots in otherwise untreated cells, and treatment with Rolipram causes these spots to disperse into the cytoplasm. Subsequent removal of Rolipram results in the rapid re-appearance of perinuclear spots.

20 Rolipram causes a change in the distribution of probes based on both PDE4A4 and PDE4A1. The non PDE4-specific inhibitors of cyclic nucleotide phosphodiesterases such as trequinsin, etazolate, milrinone, zaprinast, caffeine, theophylline and cilostamide cause no redistribution of the PDE4A probes, even at physiologically very high concentrations, whereas the PDE4-specific inhibitors Denbufylline (BRL30892), RS25344, and Ro 20-
25 1724 produce changes in the distribution of these probes which are indistinguishable from those induced by Rolipram treatment (examples 5, 6, 15 and 16). Piclamilast (RP73401), also a highly potent and specific PDE4 inhibitor, induces no redistribution of the PDE4A probes. However, RP73401 will prevent the redistribution that normally is caused by the presence of Rolipram (examples 9 and 17). Thus, only certain classes of PDE inhibitors
30 cause intracellular redistribution of the PDE4A probes, and these are all specific inhibitors of PDE4 enzymes (such as Rolipram); certain other PDE inhibitors are unable to cause the intracellular redistribution of PDE4A, but are able to compete with, reverse or prevent the action of compounds that cause redistribution, and these are also PDE4-specific inhibitors (the non PDE4-specific inhibitors are not able to reverse or inhibit the

intracellular redistribution of PDE4A caused by, for example, Rolipram). The induction of redistribution of the PDE4A probes by a certain class of PDE4-specific inhibitors is a new and inventive finding of a concept of reverse interaction between different regions of the PDE4A protein; the impulse for redistribution originates at the catalytic cleft of the PDE4A
5 where Rolipram binds, and effects a critical switch-like change at some other domain that anchors the enzymes in position within the cell. This finding immediately provides a screen for compounds that induce the intracellular redistribution of PDE4As (agonists) from those that prevent the induction of redistribution (antagonists) and a screen for those compounds that can antagonise this induced redistribution.

Detail d disclosur of the inv ntion

In this deposition, we propose a new method to screen for compounds that can disrupt the intracellular targeting of specific PDE4 isoenzymes/isoforms and that either detect or generate specific conformational states of PDE4 isoforms which either lead to alterations
5 in or define intracellular distribution.

In summary, Rolipram and certain other PDE4 inhibitors affect the distribution of at least two isoforms of PDE4. Compounds with the same effect as Rolipram on the distribution of these PDE4 isoforms also share certain other properties:

- 10 1) They are all PDE4 inhibitors, although with very wide ranging affinities, from Ro 20-1724 with an IC_{50} against PDE4 of 2.4 μM (Souness and Rao, 1997) to RS25344 with IC_{50} of 0.28 nM (Saldou *et al.*, 1998). These compounds are termed specific to PDE4, since they inhibit other PDEs either weakly or not at all. The inverse ratio between IC_{50} of a compound to PDE4s versus its
15 inhibition of other cAMP-degrading PDEs, such as PDE3s is often used as a measure of specificity. For example, $(IC_{50} \text{ PDE3})/(IC_{50} \text{ PDE4})$ values are $>2,200$ for Rolipram, 1,170,000 for RS25344 but 0.00041 for Trequinsin, clearly marking Rolipram and RS25344 highly specific for PDE4s, whereas Trequinsin is a more specific inhibitor of PDE3 enzymes.
- 20 2) They are all known to be able to displace tritiated Rolipram from what has commonly been referred to as the high affinity Rolipram binding site (HARBS), usually assayed using microsomal vesicles obtained from brain – see Souness and Rao (1997). It is widely accepted that compounds having affinity for this site are associated with certain pharmacologies and physiological effects in
25 animals, of which some are beneficial, but others are troublesome and may be characterised as undesirable side-effects, such as headaches, nausea and emesis.
- 30 3) The compounds that change the distribution of the PDE4A probes all have a relatively lower affinity for the "cAMP binding site" in PDE4s than they do for the HARBS, so that the ratio of $(IC_{50} \text{ PDE4})/(K_i \text{ for HARBS})$ gives a high score for Rolipram-like compounds, and low scores for those compounds that do not cause changes in PDE4 distribution (see Table 1).

Tabl 1

Compound	PDE4 inhibition (IC ₅₀ : μ M)	Displacement of [³ H] Rolipram (K _i : μ M)	PDE4/HARBS
(\pm) Rolipram	0.31	0.0017	238
Ro 20-1724	2.4	0.017	141
Denbufylline	0.20	0.0041	49
IBMX	14	0.84	17
RP73401	0.0012	0.0004	3
Trequinsin	0.4	1.7	0.23

(Data from Souness and Rao, 1997)

As illustrated in Table 2, the assay of the present invention will *inter alia* identify sharing
 5 the Rolipram antidepressant and/or anti-inflammatory properties, without inducing emesis.

Table 2

Drug name	Clinical effect	Emesis	Inhibition of PDE4 catalytic activity	Formation of PDE4A4 spots	Removal of Rolipram- induced PDE4A4- spots	Removal of PDE4A1 spots	Reversal of Rolipram- induced removal of PDE4A1 spots
Rolipram	antidepressant anti- inflammatory	yes	yes	yes	-	yes	-
Denbufylline (BRL30892)	anti- inflammatory	yes	yes	yes	no	yes	no
RS25344	anti- inflammatory	yes	yes	yes	no	yes	no
RO 20-1724	anti- inflammatory	yes	yes	yes	no	yes	no
Piclamilast (RP73401)	anti- inflammatory	no	yes	no	yes	no	yes
Ariflo®	anti- inflammatory	no	yes	no	yes	no	yes

One aspect of the present invention is the number and type of uses to which this
 observation can be put. Examples of such uses are:

- 10 1) Screening of potential or newly discovered PDE4 inhibitors for Rolipram-like
 properties. Such a screen may be most useful as a counterscreen, for the

detection of compounds which cause undesirable side effects, such as emesis, nausea, headaches, and excess gastric acidity.

- 2) Screening for potential or newly discovered PDE4 inhibitors with the ability to reverse or prevent the change in PDE4 distribution caused by Rolipram. Such compounds should be useful inhibitors of PDE4 but lack Rolipram-like properties, such as emesis, nausea, headaches, and excess gastric acidity.
- 3) Screening for potential inhibitors of PDE4 cellular activity that have a defined and novel mode of action, inhibitors which work by dislocating specific PDE4 isoforms from their normal sites within cells thereby modulating their effectiveness to function in cellular signalling.

The uses can be applied also to gene families B, C and D of PDE4. Uses 1 and 2 can be applied e.g. through exchange of their catalytic domains into the structure of PDE4As. Such hybrid probes, when expressed in cells, bind Rolipram and other PDE4-specific inhibitors with affinities reflecting the particular properties of the substitute catalytic site (of PDE4B in the example above), but exhibit the redistribution behaviour of the chosen PDE4A. Measurement of the redistribution behaviour reflects the binding properties specific to the imported catalytic site.

Uses 1 and 2 may also be applied rather generally to any PDE families and subfamilies thereof that do not belong to the class of PDE4 enzymes, through construction of hybrid probes between a PDE4A and the catalytic domain of the chosen PDE type in the manner described. However with these mixed class hybrids, the choice of Rolipram-like reference compound, which for PDE4s may be Rolipram, will be made from those that are known to be specific inhibitors of the particular PDE class which contributes the catalytic domain to the hybrid probe.

Thus, one aspect of the invention relates to a method to monitor changes in intracellular distribution of phosphodiesterases of subtype 4 (PDE4s) in living cells, the method comprising the steps of:

- (a) recording the intracellular distribution of the PDE4;
- (b) adding a Rolipram-like reference compound to the cells in (a) or to similar cells, the Rolipram-like compound being able to bind to the catalytic cleft of the PDE4, or to some other part of the enzyme or an associated protein whereby it induces redistribution of the PDE4 probe;
- (c) recording the intracellular distribution of the PDE4 in the cells in step (b);

(d) determining the effect on the intracellular distribution of the PDE4 of the Rolipram-like reference compound by comparing the intracellular distribution recorded in step (a) with the intracellular distribution recorded in step (c).

- 5 In another aspect of the invention inclusion of multiple cell types allows tissue specific characterisation of specific PDE4 isoforms and/or mutations thereof.

In the present invention phosphodiesterases of class 4 (PDE4s) should be understood as enzymes which are inhibited by Rolipram with an IC_{50} of less than 5 μ M; enzymes capable
10 of reacting in this specific way being selected from the list of all protein products (including all splice variants derived from) the genes designated PDE4A, PDE4B, PDE4C and PDE4D. Throughout this application the PDE4 is selected from this group.

In the present invention the intracellular distribution should be understood as the
15 distribution of a gene product within the volume of the cell. More specifically, how it is disposed relative to other identifiable cellular features or compartments, or organelles such as the plasma membrane, the Golgi membranes, endosomal vesicles, nucleus, endoplasmic reticulum, mitochondria and so on. As such, the intracellular distribution indicates possible direct association with such features, or at least with components that
20 themselves are associated in some way with those features. It should be noted that a distinctive non-homogenous distribution may be maintained not only through static anchoring or tethering, but may also be maintained through dynamic interchange, where the rates of association and dissociation favour the state of association. In this context location, position, localisation and distribution can be used interchangeably.
25 In the invention the cell and/or cells are mechanically intact and alive throughout the experiment. In another embodiment of the invention, the cell or cells is/are fixed at a point in time after the application of the influence at which the response has been predetermined to be significant, and the recording is made at an arbitrary later time.

30

The mechanically intact living cell or cells could be selected from the group consisting of fungal cells, such as a yeast cells; invertebrate cells including insect cells; and vertebrate cells, such as mammalian cells. These cells are incubated at a temperature of 30°C or above, preferably at a temperature of from 32°C to 39°C, more preferably at a
35 temperature of from 35°C to 38°C, and most preferably at a temperature of about 37°C

during the time period over which the influence is observed. In one aspect of the invention the mechanically intact living cell is part of a matrix of identical or non-identical cells.

A cell used in the present invention may contain a nucleic acid construct encoding a fusion polypeptide as defined herein and be capable of expressing the sequence encoded by the construct. The cell is a eukaryotic cell selected from the group consisting of fungal cells, such as yeast cells; invertebrate cells including insect cells; vertebrate cells such as mammalian cells. The preferred cells are mammalian cells.

10 The term "mammalian cell" is intended to indicate any living cell of mammalian origin. The cell may be an established cell line, many of which are available from The American Type Culture Collection (ATCC, Virginia, USA) or a primary cell with a limited life span derived from a mammalian tissue, including tissues derived from a transgenic animal, or a newly established immortal cell line derived from a mammalian tissue including transgenic
15 tissues, or a hybrid cell or cell line derived by fusing different celltypes of mammalian origin e.g. hybridoma cell lines. The cells may optionally express one or more non-native gene products, e.g. receptors, enzymes, enzyme substrates, prior to or in addition to the fluorescent probe. Preferred cell lines include but are not limited to those of fibroblast origin, e.g. BHK, CHO, BALB, NIH-3T3 or of endothelial origin, e.g. HUVEC, BAE (bovine
20 artery endothelial), CPAE (cow pulmonary artery endothelial), HLMVEC (human lung microvascular endothelial cells), or of airway epithelial origin, e.g. BEAS-2B, or of pancreatic origin, e.g. RIN, INS-1, MIN6, bTC3, aTC6, bTC6, HIT, or of hematopoietic origin, e.g. primary isolated human monocytes, macrophages, neutrophils, basophils, eosinophils and lymphocyte populations, AML-14, AML-193, HL-60, RBL-1, U937, RAW,
25 JAWS, or of adipocyte origin, e.g. 3T3-L1, human pre-adipocytes, or of neuroendocrine origin, e.g. AtT20, PC12, GH3, muscle origin, e.g. SKMC, A10, C2C12, renal origin, e.g. HEK 293, LLC-PK1, or of neuronal origin, e.g. SK-N-DZ, SK-N-BE(2), HCN-1A, NT2/D1.

The examples of the present invention is based on CHO cells. Therefore fibroblast
30 derived cell lines such as BALB, NIH-3T3 and BHK cells are preferred.

In another aspect of the invention the cells could be from an organism carrying in at least one of its component cells a nucleic acid sequence encoding a fusion polypeptide as defined herein and be capable of expressing said nucleic acid sequence. The organism is

selected from the group consisting of unicellular and multicellular organisms, such as a mammal.

- 5 Recording the intracellular distribution of the PDE4 in cells can be performed in numerous ways, known to the person skilled in the art. One example is antibody staining of the PDE4s wherein antibodies are raised essentially as described by Shakur *et al.* (1995); the cells are treated and stained essentially as described by Pooley *et al.* (1997). In raising the antisera, it is desirable to use isoform-specific epitopes in order to allow the
- 10 distribution of specific PDE4 isoforms to be identified and recorded. The four families of PDE4s can be individually recognised using antisera raised against peptides that copy all or part of the C-terminal protein sequences that are unique to each family. Individual PDE4 isoforms may be recognised by antisera raised in the same way against the unique N-terminal portions of these enzymes.

15

A preferred way of recording the intracellular distribution of the PDE4 in the cells is by, prior to the initial recording, constructing a probe allowing the location of the PDE4 to be recorded and subsequently transfecting cells with the constructed probe.

- 20 Throughout this application a probe must be understood as a nucleotide sequence genetically encoding an identifiable protein comprising the PDE4 or part thereof.

Identification of the probe protein in a way which allows the location of the PDE4 to be recorded can be performed in several ways. Examples are:

- 25 - immunodetection, wherein an engineered antigenic tag is incorporated into the probe, such as the "flag" or "myc" tags which are foreign and therefore unique antigens within mammalian cells, and for which mass-produced antibodies are available so that the need to develop an antibody to each new probe produced is avoided.
- direct detection, wherein the probe is engineered to include a protein sequence able to
- 30 trap or chelate a luminophore, or to breakdown luciferin and thereby generate light, or to convert a substrate to a coloured product thereby directly revealing its own cellular distribution.

A preferred method of recording the localisation of the PDE4 is by fluorescence detection,

- 35 wherein the probe is a fusion of a luminophore and a PDE4, wherein the luminophore

encodes a fluorescent protein such as the fluorophore GFP. Using fluorescence detection methods the distribution of GFP can be visualised continuously.

In the present context, the term "green fluorescent protein" (GFP) is intended to indicate a protein which, when expressed by a cell, emits fluorescence upon exposure to light of the correct excitation wavelength (e.g. as described by Chalfie, M. *et al.* (1994) *Science* 263, 802-805). Such a fluorescent protein in which one or more amino acids have been substituted, inserted or deleted is also termed "GFP". "GFP" as used herein includes wild-type GFP derived from the jelly fish *Aequorea victoria*, or from other members of the Coelenterata, such as the red fluorescent protein from *Discosoma* sp. (Matz, M.V. *et al.* 1999, *Nature Biotechnology* 17: 969-973) or fluorescent proteins from other animals, fungi or plants, and modifications of GFP, such as the blue fluorescent variant of GFP disclosed by Heim *et al.* (Heim, R. *et al.*, 1994, *Proc.Natl.Acad.Sci.* 91:26, pp 12501-12504), and other modifications that change the spectral properties of the GFP fluorescence, or modifications that exhibit increased fluorescence when expressed in cells at a temperature above about 30°C described in PCT/DK96/00051, published as WO 97/11094 on 27 March 1997 and hereby incorporated by reference, and which comprises a fluorescent protein derived from *Aequorea* Green Fluorescent Protein or any functional analogue thereof, wherein the amino acid in position 1 upstream from the chromophore has been mutated to provide an increase of fluorescence intensity when the fluorescent protein of the invention is expressed in cells. Preferred GFP variants are F64L-GFP, F64L-Y66H-GFP, F64L-S65T-GFP, F64L-E222G-GFP. One especially preferred variant of GFP for use in all the aspects of this invention is EGFP (DNA encoding EGFP which is a F64L-S65T variant with codons optimized for expression in mammalian cells is available from Clontech, Palo Alto, plasmids containing the EGFP DNA sequence, cf. GenBank Acc. Nos. U55762, U55763). Another especially preferred variant of GFP is F64L-E222G-GFP.

Another method of recording the localisation of the PDE4 by fluorescence methods uses standard chemical means to label purified PDE4 proteins with fluorophores such as fluorescein, BODIPY or Cy dyes, or rhodamine. Labelling is performed using reactive forms of these dyes as supplied by, for example, Molecular Probes Inc. (Oregon, USA), and allowing the proteins to react under the conditions and protocols recommended by the manufacturers of these reagents. Following chemical labelling and appropriate purification the probes can be microinjected into cells by standard techniques known to the art, and their behaviour within cells observed by fluorescence techniques.

It is desirable, but not always necessary, that the identity of the probe, and hence its cellular distribution, can be followed in living cells. This allows for the progress of transient changes in distribution to be recorded. Thus a preferred aspect of the invention is a method as described, wherein the comparison between the effect of the reagent and the effect of the compound is based on a time series of measurements.

When an assay has been set up, one aspect of such assay is precise knowledge on when (in a time series) the effect is seen. Then, in order to optimise the screening and to minimise the data-output, the invention relates to a method as described, wherein the comparison between the effect of the reagent and the effect of the compound is based on an end-point measurement.

In general, a probe, i.e. a "GeneX"-GFP fusion or a GFP-"GeneX" fusion, is constructed using PCR with "GeneX"-specific primers followed by a cloning step to fuse "GeneX" in frame with GFP. The fusion may contain a short vector derived sequence between "GeneX" and GFP (e.g. part of a multiple cloning site region in the plasmid) resulting in a peptide linker between "GeneX" and GFP in the resulting fusion protein.

Some of the steps involved in the development of a probe include the following:
Identify the sequence of the gene. This is most readily done by searching a depository of genetic information, e.g. the GenBank Sequence Database, which is widely available and routinely used by molecular biologists. In the specific examples below the GenBank Accession number of the gene in question is provided.

25

Design the gene-specific primers. Inspection of the sequence of the gene allows design of gene-specific primers to be used in a PCR reaction. Typically, the top-strand primer encompasses the ATG start codon of the gene and the following ca. 20 nucleotides, while the bottom-strand primer encompasses the stop codon and the ca. 20 preceding nucleotides, if the gene is to be fused behind GFP, i.e. a GFP-"GeneX" fusion. If the gene is to be fused in front of GFP, i.e. a "GeneX"-GFP fusion, a stop codon must be avoided. Optionally, the full-length sequence of GeneX may not be used in the fusion, but merely the part that localizes and redistributes like GeneX in response to a signal. In addition to gene-specific sequences, the primers contain at least one recognition sequence for a restriction enzyme, to allow subsequent cloning of the PCR product. The sites are chosen

so that they are unique in the PCR product and compatible with sites in the cloning vector. Furthermore, it may be necessary to include an exact number of nucleotides between the restriction enzyme site and the gene-specific sequence in order to establish the correct reading frame of the fusion gene and/or a translation initiation consensus sequence.

- 5 Lastly, the primers always contain a few nucleotides in front of the restriction enzyme site to allow efficient digestion with the enzyme.

Identify a source of the gene to be amplified. In order for a PCR reaction to produce a product with gene-specific primers, the gene-sequence must initially be present in the reaction, e.g. in the form of cDNA. Information in GenBank or the scientific literature will usually indicate in which tissue(s) the gene is expressed, and cDNA libraries from a great variety of tissues or cell types from various species are commercially available, e.g. from Clontech (Palo Alto), Stratagene (La Jolla) and Invitrogen (San Diego). Many genes are also available in cloned form from The American Type Tissue Collection (Virginia).

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Optimise the PCR reaction. Several factors are known to influence the efficiency and specificity of a PCR reaction, including the annealing temperature of the primers, the concentration of ions, notably Mg^{2+} and K^+ , present in the reaction, as well as pH of the reaction. If the result of a PCR reaction is deemed unsatisfactory, it might be because the parameters mentioned above are not optimal. Various annealing temperatures should be tested, e.g. in a PCR machine with a built-in temperature gradient, available from e.g. Stratagene (La Jolla), and/or various buffer compositions should be tried, e.g. the OptiPrime buffer system from Stratagene (La Jolla).

Clone the PCR product. The vector into which the amplified gene product will be cloned and fused with GFP will already have been taken into consideration when the primers were designed. When choosing a vector, one should at least consider in which cell types the probe subsequently will be expressed, so that the promoter controlling expression of the probe is compatible with the cells. Most expression vectors also contain one or more selective markers, e.g. conferring resistance to a drug, which is a useful feature when one wants to make stable transfectants. The selective marker should also be compatible with the cells to be used.

The actual cloning of the PCR product should present no difficulty as it typically will be a one-step cloning of a fragment digested with two different restriction enzymes into a vector digested with the same two enzymes. If the cloning proves to be problematic, it

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may be because the restriction enzymes did not work well with the PCR fragment. In this case one could add longer extensions to the end of the primers to overcome a possible difficulty of digestion close to a fragment end, or one could introduce an intermediate cloning step not based on restriction enzyme digestion. Several companies offer systems for this approach, e.g. Invitrogen (San Diego) and Clontech (Palo Alto).

Once the gene has been cloned and, in the process, fused with the GFP gene, the resulting product, usually a plasmid, should be carefully checked to make sure it is as expected. The most exact test would be to obtain the nucleotide sequence of the fusion-gene.

Once a DNA construct for a probe has been generated, its functionality and usefulness may be evaluated by transfecting it into cells capable of expressing the probe.

Some of the advantages of using live cells in the design and operation of assays to screen for therapeutic drugs include the inherent ability of the assay to determine the availability of any compound to targets in the cell interior, and also an inherent assessment of the possible toxicity of a test compound or its cellular metabolites over the period of the assay, for example the PDE4A4 spot assay can involve up to 24 hours exposure of cells to test compounds before the readout, or measurement, is made, during which time any immediate toxic effects of test compounds or their cellular metabolites on cells can be observed.

Numerous cell systems for transfection exist. A few examples are *Xenopus* oocytes or insect cells, such as the sf9 cell line, or mammalian cells isolated directly from tissues or organs taken from healthy or diseased animals (primary cells), or transformed mammalian cells capable of indefinite replication under cell culture conditions (cell lines). However, it is preferred that the cells used are mammalian cells. This is due to the complex biochemical interactions specific for each cell type.

The fluorescence of the cell is inspected soon after, typically the next day. At this point, two features of cellular fluorescence are noted: the transfection intensity and the sub-cellular localisation.

The intensity should usually be at least as strong as that of unfused GFP in the cells. If it is not, the sequence or quality of the probe-DNA might be faulty, and should be carefully checked. Other causes of poor expression can often be corrected by linearising the plasmid DNA prior to transfection, or by increasing the concentration of DNA used for a
5 transfection process, or by choosing a different transfection agent or method, of which many are known to those skilled in the art.

The sub-cellular localisation is an indication of whether the probe is likely to perform well. If it localises as expected for the gene in question, e.g. form spots upon treatment with
10 Rolipram, it can immediately go on to a functional test. If the probe is not localised soon after the transfection procedure, it may be because of overexpression at this point in time, as the cell typically will have taken up very many copies of the plasmid, and localisation will occur in time, e.g. within a few weeks, as plasmid copy number and expression level decreases. If localisation does not occur after prolonged time, it may be because the
15 fusion to GFP has destroyed a localisation function, e.g. masked a protein sequence essential for interaction with its normal cellular anchor-protein. In this case the opposite fusion might work, e.g. if GeneX-GFP does not work, GFP-GeneX might, as two different parts of GeneX will be affected by the proximity to GFP. If this does not work, the proximity of GFP at either end might be a problem, and it could be attempted to increase
20 the distance by incorporating a longer linker between GeneX and GFP in the DNA construct. Lack of proper localisation may also be due to a lack of suitable anchorage or scaffold sites within the cell, which can often be corrected by co-transfection of genes coding for the the protein component or components responsible for providing the appropriate anchorage or scaffold sites.

25

If there is no prior knowledge of localisation, and no specific localisation is observed, it may be because the probe should not be localised at this point, because such is the nature of the protein fused to GFP. It should then be subjected to a functional test.

30 In a functional test, the cells expressing the probe are treated with at least one Rolipram-like reference compound. If a redistribution is observed and if prior knowledge suggests that it should translocate from location X to location Y, then the probe has passed the first critical test. In this case it can go on to further characterisation and quantification of the response.

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If it does not perform as expected, it may be because the cell lacks at least one component of the signalling pathway, e.g. a cell surface receptor, or the anchoring site is absent or saturated, or there is species incompatibility, e.g. if the probe is modelled on sequence information of a human gene product, and the cell is of hamster origin. In both instances one should identify other cell types for the testing process where these potential problems would not apply.

In the present invention preferred fusion probes are listed in Table 3. Most preferred probes are PDE4 probes of human origin, fusion probes like HSPDE4A1-EGFP, HSPDE4A4-EGFP, HSPDE4A4-H506N-EGFP, and HSPDE4A4- Δ LR2-EGFP. The construction and testing of the probes used in the scientific findings of the present invention are described in examples 1 and 2. Terminology used here for PDE4 proteins follows the recommendations of the nomenclature committee (Beavo *et al.*, 1994). The recommendations of the committee are that the first two letters should indicate the source species (HS, *Homo sapiens*; RN, *Rattus norvegicus*), that PDE be used to designate cyclic nucleotide phosphodiesterase, that an arabic numeral indicate the superfamily (4, in this case), that a single letter indicate the gene family (A, B, C, D for PDE4), that another arabic numeral indicate the splice variant, and finally a single letter be used for the report describing the enzyme in question (generally omitted after validation and full description of the isoform).

The behaviour and use of probes based on PDE4A may also be applied to gene families B, C and D of PDE4 through exchange of their catalytic domains into the structure of PDE4As. Since there exist identifiably homologous regions within the catalytic domains of the four gene families, hybrid molecules can conveniently be constructed as follows: A conserved amino acid is chosen within a stretch of the PDE4A protein's primary sequence, within the catalytic domain, that is homologous, or preferably identical, to the corresponding region in an enzyme from a different gene family of PDE4: one such region of amino acid identity is found for example between amino acids 457 and 467 (numbering from HSPDE4A4). Standard molecular biology techniques are then used to remove all codons for amino acids in the PDE4A gene sequence that are C-terminal to that position, and they are replaced with the corresponding coding sequence from a different PDE4 gene (say that of PDE4B). The PDE4 hybrid sequence is then fused to a sequence coding for a label/marker such as EGFP, such that the gene product will have EGFP attached to the C-terminal of the enzyme.

These hybrid probes, when expressed in cells, bind Rolipram and other PDE4-specific inhibitors with affinities reflecting the particular properties of the substitute catalytic site (of PDE4B in the example above), but exhibit the redistribution behaviour of the chosen PDE4A. Measurement of the redistribution behaviour reflects the binding properties
5 specific to the imported catalytic site.

It is published that the association of certain partner proteins with certain PDE4s will affect the affinity of the enzymes for Rolipram. It is presently contemplated that specific
10 compounds binding to the enzyme (e.g. Rolipram) will alter the mobility of the enzyme (that is the binding to an anchorage or docking partner) altering the freedom of the enzyme to move in 3 dimensional space.

One major aspect of this invention is based on the finding that incubation of cells with
15 Rolipram causes redistribution of PDE4A probes. This redistribution is not a consequence of an increase in cAMP brought about by inhibition of PDE4 activity, since the redistribution is not simply mimicked by treatment with IBMX, or Forskolin \pm IBMX (example 10).

Rather, it seems that binding of Rolipram and certain other PDE4 inhibitors induces
20 conformational changes in PDE4A leading to a change in the affinity of the PDE4A probes for their docking or anchorage partners, which results in their subsequent redistribution in the cell.

It is further speculated, from the known properties of Rolipram and the other compounds
25 that mimic the effects of Rolipram on PDE4A redistribution, that it is the binding of these compounds to what is commonly referred to as the "high affinity Rolipram binding site" (HARBS) or "Sr", or HPDE4 form, (Souness and Rao, 1997) that specifically triggers the changes that lead to redistribution of the PDE4A probes. Specific PDE4 inhibitors such as RP73401 and SB207499 do not recognise HARBS, but apparently recognise an
30 alternative site within the catalytic domain and inhibit the cAMP hydrolysing ability of the catalytic site (Sc) in a somewhat different way; Rolipram also binds in this second way, and in this case it is said to bind to a "low affinity site", or to the LPDE4 form (Souness and Rao, 1997). Since RP73401 does not cause redistribution of PDE4A probes, it is predicted that SB207499 will also fail to cause redistribution of these probes, and further,
35 that like RP73401, SB207499 will compete against and reverse the redistribution caused by Rolipram and other compounds that bind to the Sr. It is presently unknown whether

HARBS and the low affinity site (or sites), are truly distinct and separate positions within the catalytic cleft of PDE4 enzymes, or whether they represent different conformational states of the same site. The role of binding to the Sc or the Sr, in terms of elevation of cAMP and inhibition of cell responses, is not yet fully understood. The importance of the

5 Sr in influencing the pharmacological profile of PDE4 inhibitors may have implications in predicting not only efficacy but also the side-effects of these drugs, such as nausea, emesis, excess gastric acid secretion and headache, which have hampered the clinical development of PDE4 inhibitors to date. (Souness & Rao, 1997)

- 10 The majority of the experiments in the present application are based on the effects of Rolipram. Thus, it is preferred that the method of the invention is carried out using Rolipram as a reference compound. However, in another embodiment of the present invention, a Rolipram-like reference compound is used. A Rolipram-like reference compound is a compound, sharing the properties of Rolipram with regards to their ability
- 15 to cause redistribution of the PDE probe being used and with regards to the ability to inhibit the catalytic activity of the PDE4.

It is an important aspect of the present invention that a change in localisation of the PDE4 is detected as a consequence of the treatment with the Rolipram-like reference

20 compound. Examples 1-14 illustrates the formation of spots caused by Rolipram. Thus it is preferred that the method relates to a change in localisation of the PDE4 as formation of spots. Example 15 illustrates the dispersal of spots caused by Rolipram treatment. Thus, it is another preferred embodiment that the method relates to a change in localisation of the PDE4 as dispersal of spots.

- 25 One aspect of the present invention relates to the identification of a compound that produces a distinct change in intracellular distribution of the probe, such as a test compound that will mimic the distinct change in localisation caused by the Rolipram-like reference compound, and by inference that the test compound and the Rolipram-like
- 30 reference compound will share a common pharmacological profile.

This identification of compounds with an agonistic effect is preferably carried out as a method to monitor changes in intracellular distribution of PDE4s in living cells caused by a test compound, the method comprising the steps of:

- 35 O1) optionally constructing a probe allowing the location of the PDE4 to be recorded;
O2) optionally transfecting cells with the constructed probe of step (O1);
(a) recording the intracellular distribution of the PDE4;

- (b) adding a Rolipram-like reference compound to the cells in (a) or to similar cells, the Rolipram-like reference compound being able to bind to the catalytic cleft of the PDE4;
- (b1) adding the test compound to the cells in (a) or similar cells;
- (b2) recording the intracellular distribution of the PDE4 in the cells in step (b1);
- 5 (c) recording the intracellular distribution of the PDE4 in the cells in step (b);
- (d) determining the effect on the intracellular distribution of the PDE4 of the Rolipram-like reference compound by comparing the intracellular distribution recorded in step (a) with the intracellular distribution recorded in step (c);
- (d1) determining the effect of the test compound by comparing the intracellular distribution
- 10 recorded in step (b2) with the intracellular distribution recorded in step (a);
- the pharmacology of the test compound being established by comparing the determined effect in step (d1) with the determined effect in step (d) a substantial copy of the effect determined in step (d), in step (d1), being indicative of an agonistic effect of the test compound to the Rolipram-like reference compound in regards to the change in
- 15 intracellular distribution of the PDE4.
- Agonists induce the formation of very bright spots, often a single pair, in cells expressing the HSPDE4A4-EGFP probe. Certain agonists (RS25344, but not Rolipram) will induce formation of the same kind of spots in cells expressing the H506N mutant of this probe, indicating their ability to "bridge" or compensate for the mutation which removes Rolipram
- 20 agonism. In all cases, formation of bright spots requires protein synthesis and accumulation of probe: two hour incubation with Rolipram, or other agonists, is sufficient to determine that spot formation is under way, but the spots become larger and brighter, and therefore easier to measure, after a total of about 6 hours incubation with the test compound. Between 6 to 24 hours, spot numbers do not increase greatly, although their
- 25 size and brightness does continue to grow. Incubation of cells with test compounds for a period of 16 hours before fixation proves to be a convenient and reliable method to screen batches of compounds, allowing many plates of cells to be treated in the evening, incubated overnight, and fixed, stained and analysed the following morning.
- With the HSPDE4A1-EGFP probe, agonists induce dispersal of the bright spots that
- 30 normally lie in the perinuclear area of cytoplasm. Dispersal of spots is easily measurable after 60 to 90 minutes, and therefore is a faster process than spot formation with HSPDE4A4-EGFP.
- Agonists found through use of either probe may be expected to be specific PDE4 inhibitors, and a suitable secondary screen for PDE4 specific inhibition is desirable to
- 35 confirm this property.

Another aspect of the present invention relates to the identification of a test compound that will prevent and reverse the distinct change in localisation produced by action of the Rolipram-like reference compound e.g. by displacing the Rolipram-like reference compound.

This identification of test compounds with an antagonistic effect is preferably carried out as a method to monitor changes in intracellular distribution of PDE4s in living cells, the method comprising the steps of:

- 10 O1) optionally constructing a probe allowing the location of the PDE4 to be recorded;
O2) optionally transfecting cells with the constructed probe of step (O1);
(a) recording the intracellular distribution of the PDE4;
(b) adding a Rolipram-like reference compound to the cells in (a) or to similar cells, the Rolipram-like reference compound being able to bind to the catalytic cleft of the PDE4;
- 15 (b1) adding a test compound to the cells with the Rolipram-like reference compound in step (b) or similar cells;
(b2) recording the intracellular distribution of the PDE4 in the cells in step (b1);
(c) recording the intracellular distribution of the PDE4 in the cells in step (b);
(d) determining the effect on the intracellular distribution of the PDE4 of the Rolipram-like
20 reference compound by comparing the intracellular distribution recorded in step (a) with the intracellular distribution recorded in step (c).;
(d1) determining the effect of the test compound by comparing the intracellular distribution recorded in step (b2) with the intracellular distribution recorded in step (a);
the pharmacology of the test compound being established comparing the determined
25 effect in step (d1) with the determined effect in step (d) a reversal, in step (d1), of the effect determined in step (d) to an effect substantially identical to the effect determined in step (a) being indicative of an antagonistic effect of the test compound on the Rolipram-like reference compound in regards to the change in intracellular distribution.
- 30 The pharmacology of the test compound can also be established by comparing the determined effect in step (d1) with the determined effect in step (d) an increased effect, in step (d1), of the effect determined in step (d), comparable to an effect in step (d) obtained with a higher doses of the Rolipram-like reference compound, being indicative of the augmenting effect of the test compound on the Rolipram-like reference compound in
35 regards to the change in intracellular distribution.

Antagonists induce the dispersal of the very bright spots formed by the Rolipram-like reference compound in cells expressing the HSPDE4A4-EGFP probe. Dispersal of bright spots does not require protein synthesis, and is generally easily measurable after 30 to 60 minutes. Some compounds at higher concentrations, such as RP73401, can disperse spots very rapidly; spots formed by 2 μ M Rolipram over 16 hours will disperse within 10 minutes with 1 μ M RP73401. A screen for antagonists may involve incubating cells with Rolipram-like reference compound (say 3 μ M Rolipram, or 0.5 μ M RS25344) for a period of 16 hours, then adding the test compound and incubating further for a period of 60 minutes before being fixed, stained and analysed.

- 10 With the HSPDE4A1-EGFP probe, antagonists reverse the dispersal of perinuclear bright spots that normally results from treatment with Rolipram-like reference agonists such as Rolipram. Compounds may be added simultaneously with Rolipram-like reference compounds, or at some time later (such as after 60 to 90 minutes incubation with the Rolipram-like reference compound). Reappearance of spots is easily measurable after 15 240 minutes.

As detailed in examples 10, 11 and 12, certain treatments are known to disperse PDE4A4 Rolipram spots in CHO cells. These include [Forskolin + IBMX] (example 10) and [PMA \pm ionomycin] (example 11). Appropriate counterscreens will help to identify compounds that redistribute PDE4s through dislocation: dislocator compounds will not bind to the catalytic cleft, so will not inhibit catalytic activity of PDE4s, will not induce increase in cAMP/activation of PKA in cells (as forskolin + IBMX does), will not mimic the effects of PMA \pm ionomycin, i.e. directly stimulate PKC isoforms, perhaps through prolonged increase of intracellular Ca^{2+} or through increase in levels of diacyl glycerol.

Since the antagonist assay using the PDE4A4 probe detects compounds by their ability to disperse spots, this assay is also useful in detecting compounds that dislocate PDEs, or their anchor protein(s), from their preferred cellular location. If a compound is found to disperse spots in the PDE4A4 antagonist assay, AND causes spots to reform, or persist, in the 4A1 antagonist assay, that compound is most likely to be a PDE4 specific inhibitor with little affinity for the HARB site, and should have properties similar to RP73401 (and, as predicted, to Ariflo[®], or SB207499). If a compound found in the PDE4A4 antagonist assay fails to cause spots to reform or persist in the 4A1 antagonist assay, and does not screen as positive in the suggested counterscreens, that compound is likely to be a dislocator of PDE4A4, or it's anchor protein(s). By extension, a compound with activity in the 4A1 agonist assay, but no activity in the PDE4A4 agonist assay, and which does not

screen positive in a PDE4 inhibition assay, is likely to be a dislocator of PDE4A1 or it's anchor protein(s).

- Test compounds identified by the method of the present invention include specific
- 5 inhibitors of PDE4 enzymes, which can be categorised from their effect on the distribution of PDE4A probes as being either Rolipram-like, or non Rolipram-like inhibitors. It is speculated that the crucial property all Rolipram-like inhibitors share, is the ability to bind to the high affinity Rolipram binding site of PDE4 enzymes, and/or the ability to trigger a conformational change in PDE4A enzymes from an interaction within the catalytic cleft.
- 10 Test compounds identified by the method of the present invention are also predicted to include dislocator compounds, which either disrupt/enhance the association of PDE4 isoenzymes with particular anchor proteins or to disrupt/enhance machinery responsible for the trafficking of PDE4 proteins between different locations within the cell. In so doing, compounds are identified whose usage would be in disrupting or relocating the
- 15 placement of a PDE4 isoenzyme in its established place in the cell so as to enhance compartmentalised cAMP function. Through this novel approach and the derivation of appropriate assays an entirely new way of generating PDE4 isoform-selective therapeutics is envisaged.
- 20 It is preferred that the test compound identified, e.g. as an agonist or an antagonist, is a single substance composed of one or more chemical elements. An example of such a test compound is a peptide.

The term "compound" is intended to indicate any sample which has a biological function

25 or exerts a biological effect in a cellular system. The sample may be a sample of a biological material such as a sample of a body fluid including blood, plasma, saliva, milk, urine, or a microbial or plant extract, an environmental sample containing pollutants including heavy metals or toxins, or it may be a sample containing a compound or mixture of compounds prepared by organic synthesis or genetic techniques.

30

- In a further aspect of the invention it is preferred that the test compound binds to the catalytic cleft of the PDE4. The catalytic cleft of the PDE4 is the cleft within the protein macromolecule to which a substrate for the enzyme is introduced, and where conditions for a specific chemical (or physical) reaction involving the substrate are
- 35 thermodynamically optimised for the reaction to run in a particular direction. For the PDE4s it lies within the region recognised as the conserved catalytic domain, which (using

amino acid numbering from HSPDE4A4) has been determined from a combination of truncation and deletion experiments as comprising some 315 to 348 amino acids located between residues 332/365 and 680 (Houslay, Sullivan and Bolger, 1998). Mutations and deletions within this consensus region are likely to ablate or decrease cAMP binding and hydrolytic abilities of PDE4 enzymes. The terms catalytic site and active site have a similar meaning in this regard.

The affinity with which a test or Rolipram-like reference compound binds to the catalytic cleft may be determined through use of standard radioligand binding assays, wherein the test compound is radiolabelled and incubated with a more or less purified preparation of the target PDE4 enzyme. Such an enzyme preparation may be obtained from cells transfected with and expressing the PDE4 enzyme chosen. Typical systems used in such a procedure may be found described in Saldou *et al.* (1998). Alternatively displacement of tritiated Rolipram from brain-derived microsomal preparations can be used to determine the affinity of a test compound for the so-called high-affinity Rolipram binding site of PDE4 enzymes.

Binding affinity, effects on PDE4A probe redistribution and inhibitory effects on catalytic activity are not necessarily correlated. In certain aspects of the invention is preferred that test compounds found through such use of the invention also inhibit the catalytic activity of PDE4s. Effects of a test compound on the catalytic activity of a PDE4 can easily be determined by standard competitive binding experiments between PDE inhibitors and cAMP on enzyme activity for which known amounts of cAMP substrate and fixed amounts of enzyme are incubated together with various amounts of inhibitor substance for fixed periods of time, after which the reaction is stopped and the residual amount of unhydrolysed cAMP is measured. This may be done for any test sample by use of a scintillation proximity based assay (SPA) designed to measure the competition between cAMP in the test sample and a known amount of radiolabelled cAMP for binding to a cAMP-specific antibody attached to scintillant beads (Hancock *et al.*, 1995). The assay is read in a scintillation counter where the counts per sample are inversely related to the amount of cAMP present in the test sample. SPA kits for measurement of cAMP are available from Amersham Pharmacia Biotech (Amersham, UK).

Yet another class of compounds that can be detected by the method of the invention is described *inter alia* in example 13. These compounds inhibit the reappearance of spots in cells expressing the PDE4A4 probe, where the spot reappearance process is triggered by

various imposed conditions, and is specific to cells that have formed spots under the influence of a Rolipram-like PDE4 inhibitor (an agonist compound selected from Rolipram, Ro 20-1724, RS25344 etc.), and where the cells have then been cleared of spots by removal of the agonist compound. Thalidomide is an example of a compound that is a

5 spot reappearance inhibitor, as described in example 13.

Spot reappearance inhibitors may be compounds that inhibit cellular stress responses.

The identification of test compounds as spot reappearance inhibitors is preferably carried out as a method comprising the steps of:

- (1) treating PDE4A4 expressing cells with a reference agonist compound for a period to
10 induce spots (e.g. 7-24 hours);
- (2) checking that spots have formed;
- (3) washing away the Rolipram-like reference compound and leaving cells in incubator for spots to completely disappear (about 150 minutes);
- (4) checking that all spots have disappeared;
- 15 (5) adding the test compounds, keeping some wells as negative controls;
- (6) exposing all cells to 100 mM salt and 4°C for 4 hours, or leaving the cells at ambient conditions for 4 hours to allow cooling to 22°C, alkalisation of medium (pH=6.5 shifting to pH=8.2) and partial evaporation (about 20% decrease in volume);
- (7) determining the degree of spot reappearance compared to control wells that have not
20 been treated with any test compound.

As will be evident to the person skilled in the art, compounds capable of inhibition of the function of PDE4 are capable of preventing/decreasing inflammation and/or depression.

The present invention provides at least two novel approaches to identifying such
25 compounds. All approaches are based on the initial discovery that Rolipram, by binding to the catalytic cleft of the PDE4, induces a change in the cellular distribution of the PDE4.

One method is a method to determine if a compound is a dislocator of PDE4, comprising the steps of:

- 30 - testing if the compound removes PDE4-spots, where PDE4-spots may optionally be induced by a Rolipram-like reference compound and
 - testing if the compound inhibits the catalytic activity of the PDE4;
- the compound being a dislocator of PDE4, if the compound removes PDE4-spots and if the compound does not inhibit the catalytic activity of PDE4.

PDE4 dislocators will remove the PDE4 away from the native location in the cell and thereby increase the concentration of cAMP in said native location ('compartment') in the cell. Such increased concentration of cAMP is also seen upon inhibition of the catalytic activity of the PDE4, however, by dislocating the PDE4, and thereby not acting directly on
5 the catalytic well conserved site, the compound will act e.g. at the binding domain of the PDE4, thereby providing isoform-specific 'inhibitors' of PDE4.

One aspect of the present invention thus relates to a PDE4 dislocator obtainable by the method described. Such PDE4 dislocator is preferably included into a pharmaceutical
10 composition comprising a compound, the compound being a dislocator of PDE4, and the pharmaceutical composition having a market authorisation, the market authorisation being based on an application for market authorisation comprising data showing removal of PDE4-spots, optionally induced by a Rolipram-like reference compound, by the compound and lack of inhibition of the catalytic activity of PDE4 by the compound.

15

An example of the market authorisation is described in 65/65/EEC. The data required specified in Article 4,8. of said directive.

Preferred dislocators of PDE4 are dislocators of PDE4A isoforms, such as the PDE4A1
20 isoform and/or the PDE4A4 isoform.

PDE4A1 dislocators are identified by a method comprising the steps of:

- testing if the compound removes PDE4A1-spots, and
 - testing if the compound inhibits the catalytic activity of the PDE4A1;
- 25 the compound being a dislocator of PDE4A1, if the compound removes PDE4A1-spots and if the compound does not inhibit the catalytic activity of PDE4A1.

PDE4A1 dislocators obtainable by the method described are preferably included in a pharmaceutical composition wherein the indication on the market authorisation is
30 diseases in the central nervous system such as depression.

PDE4A4 dislocators are identified by a method comprising the steps of:

- testing if the compound removes PDE4A4-spots induced by a Rolipram-like reference compound and
- 35 - testing if the compound inhibits the catalytic activity of the PDE4A4;

the compound being a dislocator of PDE4A4, if the compound removes PDE4A4-spots and if the compound does not inhibit the catalytic activity of PDE4A4.

PDE4A4 dislocators obtainable by the method described are preferably included in a
5 pharmaceutical composition wherein the indication on the market authorisation is inflammatory diseases. Examples of inflammatory diseases are joint inflammation, Crohn's disease, inflammatory bowel disease, respiratory diseases, chronic obstructive pulmonary disease (COPD), including asthma, chronic bronchitis, pulmonary emphysema, endotoxic shock, toxic shock syndrome, systemic lupus erythematosus, psoriasis, bone
10 resorption diseases, reperfusion injury, cancer and HIV infection.

Another method according to the invention, is a method to determine if a compound is a low emesis PDE4 inhibitor comprising the steps of:

- testing if the compound causes PDE4A4-spots induced by a Rolipram-like reference
15 compound to dissolve,
 - testing if the compound induces re-appearance of PDE4A1-spots in cells exposed to a Rolipram-like reference compound, and
 - testing if the compound inhibits the catalytic activity of PDE4;
- the compound being a low emesis PDE4 inhibitor if the compound removes spots induced
20 by the Rolipram-like reference compound and induces re-appearance of PDE4A1 spots in cells exposed to the Rolipram-like reference compound and if the compound inhibits the catalytic activity of PDE4.

Low emesis PDE4 inhibitors will inhibit the catalytic activity of the PDE4, causing the anti-
25 inflammatory and anti-depressant effects without causing the side effects as emesis, nausea, headaches, and excess gastric acidity.

It is preferred that the low emesis PDE4 inhibitor is included in a pharmaceutical composition comprising a compound, the compound being a low emesis PDE4 inhibitor,
30 and the pharmaceutical composition having a market authorisation, the market authorisation being based on an application for market authorisation comprising data showing that the compound removes spots induced by the Rolipram-like reference compound, and that the compound induces re-appearance of PDE4A1 spots in cells exposed to the Rolipram-like reference compound, and that the compound inhibits the
35 catalytic activity of PDE4.

In one aspect the indication on the market authorisation is inflammatory diseases.

Another aspect relates to the use of a Low emesis PDE4 inhibitor or a PDE4 dislocator, or
5 a pharmaceutically acceptable salt, ester, amide or prodrug thereof, for the preparation of
medicament for the treatment of inflammatory diseases including joint inflammation,
Crohn's disease, and inflammatory bowel disease; respiratory diseases such as chronic
obstructive pulmonary disease (COPD) including asthma, chronic bronchitis, and
pulmonary emphysema; infections diseases including endotoxic shock and toxic shock
10 syndrome; immune diseases including systemic lupus erythematosus and psoriasis; and
other diseases including bone resorption diseases and reperfusion injury and conditions
associated with proliferating hematopoietic cells, such as cancer and HIV infection;
diseases in the central nervous system including depression.

15 Another important aspect of the present invention relates to the use of a Low emesis
PDE4 inhibitor, or a pharmaceutically acceptable salt, ester, amide or prodrug thereof,
capable of competing against and reversing the effect of the Rolipram-like reference
compound on the intracellular distribution of the PDE4 and being capable of inhibiting the
catalytic activity of the PDE4 for the preparation of medicament for the treatment of
20 inflammatory diseases, e.g. rheumatoid arthritis.

Yet another aspect of the invention is a method for treating inflammatory diseases, e.g.
asthma, or depression in an individual comprising administering to the individual an
effective amount of a compound, or a pharmaceutically acceptable salt, ester, amide or
25 prodrug thereof, the compound being capable of competing against and reversing or
mimicking the effect of the Rolipram-like reference compound on the intracellular
distribution of the PDE4 and the compound being capable of mimicking the effect of the
Rolipram-like reference compound on the catalytic activity of PDE4s.

30 The invention further relates to the test compound identified, or identifiable, by a method
according to the invention. E.g. a Low emesis PDE4 inhibitor or a PDE4 dislocator.

It is evident to the skilled person, that a PDE4 dislocator or a Low emesis PDE4 inhibitor
identified by the methods described will need further testing prior to human trials. Apart
35 from the toxicology requirements, the PDE4 dislocator is tested in functional assays for

relevant action, and counter indications, at both the cellular and organismal level.

Examples of such assays are the *in vitro* measurement of LPS-stimulated TNF α release from human peripheral blood mononucleocytes (e.g. as described by Barnett *et al.* 1998), and an *in vivo* measurement of anti-inflammatory action such as suppression of
5 antigen-induced eosinophilia and bronchoconstriction in rat lung (e.g. as described by Hughes *et al.* 1996, an Asthma model) or *in vivo* measurement of an anti-depressant function such as the induction of brain-derived neurotrophic factor (BDNF) in rat hippocampus (a depression model; Fujimaki *et al.* 2000) or *in vivo* measurement of the amelioration of collagen II-induced arthritis in rats (a rheumatoid arthritis model; Nyman *et al.* 1997). Additionally, PDE4 dislocator compounds are screened for unwanted potential
10 emetic properties in a ferret emesis test (e.g. as described by Robichaud *et al.* 1999).

The pharmaceutical compositions described herein may be formulated according to conventional pharmaceutical practice, see, e.g., "Remington's Pharmaceutical Sciences"
15 and "Encyclopedia of Pharmaceutical Technology".

It is likely that mutations in an individual within the PDE4 can be the cause of certain disease states e.g. immunological illnesses and depression. Those individuals can, in one aspect of the invention, be diagnosed for functional mutations in the the Rolipram binding
20 site, or in the anchor binding site

- by fishing the PDE4 subtype (e.g. by PCR)
- fuse the PDE4 subtype to GFP
- add Rolipram
- measure change in distribution of the PDE4.

25

Another important aspect of the invention is a method for providing the basis for diagnostic methods for the early and accurate detection and/or quantitation of PDE distribution associated with joint inflammation, Crohn's disease, and inflammatory bowel disease; respiratory diseases such as chronic obstructive pulmonary disease (COPD)
30 including asthma, chronic bronchitis, and pulmonary emphysema; infectious diseases including endotoxic shock and toxic shock syndrome; immune diseases including systemic lupus erythematosus and psoriasis; and other diseases including bone resorption diseases and reperfusion injury and conditions associated with proliferating hematopoietic cells, such as cancer and HIV infection.

35

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Examples

Example 1: Cloning and construction of the GFP-labelled PDE probes

Here is described the cloning and construction of specific PDE4A variants fused to a GFP.

Currently at least 5 PDE4A splice variants are known. These all share C-terminal

- 5 sequences but differ in their N-termini, where targeting sequences of PDE4As are believed to be located. To best preserve the normal distribution of PDE4As, the fusions are made between the C-terminus of the PDE4A species and the N-terminal of the GFP.

- To construct the HSPDE4A1-EGFP fusion, the ca. 1.95 kb coding region of HSPDE4A1
 10 (GenBank Acc.no. U97584) is amplified using PCR and primers 4A1-top and 4A-bottom described below. The top primer includes specific HSPDE4A1 sequences following the ATG, a Kozak sequence, and a Hind3 cloning site. The bottom primer includes the common PDE4A C-terminal sequence minus the stop codon, a BamH1 cloning site, and two extra nucleotides to preserve the reading frame when inserted into in pEGFP. The
 15 PCR product is digested with restriction enzymes hind3 and BamH1, and cloned into pEGFP (Clontech, Palo Alto; GenBank Accession number U55762) cut with Hind3 and BamH1. This produces a HSPDE4A1-EGFP fusion under the control of the CMV promoter. The resulting plasmid is referred to as PS461 and is deposited under the Budapest Treaty with Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH
 20 (DSMZ) on 17 April 2000 with DSM 13449.

4A1-top (SEQ ID NO: 9):

5'-GTAAGCTTAAGATGCCCTTGGTGGATTCTTC-3', specific for PDE4A1,

4A-bottom (SEQ ID NO: 10):

5'-GTGGATCCCAGGTAGGGTCTCCACCTGA-3'

25

- To construct the HSPDE4A4-EGFP fusion, the common ca. 1.9 kb C-terminal part of HSPDE4A4 (GenBank Acc.no. L20965) is amplified using PCR with primers 4A-Ct-top and 4A -bottom described below. The sequence of the top primer contains a silent mutation which introduces a Dra1 site exactly at the beginning of the shared 4A region.
 30 The bottom primer includes the common C-terminal sequence minus the stop codon, a BamH1 cloning site, and two extra nucleotides to preserve the reading frame when cloned into pEGFP. The unique ca. 0.8 kb N-terminal part of HSPDE4A4 is amplified using PCR in the presence of 5% DMSO with primers 4A4-top and 4A4N-bottom described below. The top primer includes specific HSPDE4A4 sequences following the ATG, a Kozak

sequence, and a Hind3 cloning site. The bottom primer spans the junction of the unique 4A4 N-terminal part and the common 4A C-terminal part, and it contains a silent mutation which introduces a Dra1 site exactly at the beginning of the shared 4A region. The PCR products are digested with the relevant restriction enzymes (Hind3 and Dra1 for the unique N-terminal part and Dra1 and BamH1 for the common C-terminal part), and ligated together into pEGFP (Clontech, Palo Alto; GenBank Accession number U55762) digested with Hind3 and BamH1. This produces a HSPDE4A4-EGFP fusion under the control of a CMV promoter. The resulting plasmid is referred to as PS462 and is deposited under the Budapest Treaty with Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ) on 17 April 2000 with DSM 13450.

4A-Ct-top (SEQ ID NO: 11): 5'-GTTTAAAAGGATGTTGAACCGTGAGCTC-3'

4A-bottom (SEQ ID NO: 12): 5'-GTGGATCCCAGGTAGGGTCTCCACCTGA-3'

4A4-top (SEQ ID NO: 13): 5'-GTAAGCTTGCGCCATGGAACCCCCGACC-3'

4A4N-bottom (SEQ ID NO: 14): 5'-GGTTTTAACTTGTGCGAGGCCATCTCGCTGAC-3'

Catalytically inactive PDE4 fusions that redistribute normally within the cell, can be constructed by introducing specific point mutations in the catalytic domain. Use of such fusions may be advantageous if a cell is sensitive to some overexpression of a catalytically active PDE4. Many mutations are known in PDE4As that greatly reduces catalytic activity, e.g. H506N in HSPDE4A4.

Plasmid PS535 (HSPDE4A4-H506N-EGFP) is a variant of PS462 (HSPDE4A4-EGFP) containing a substitution of His-506 to Asn in HSPDE4A4. This substitution is introduced using the PCR-based Quickchange mutagenesis kit (Stratagene, La Jolla). The PCR reaction leading to this substitution utilises plasmid PS462 as template and the complementary primers 4AH-N-forward and 4AH-N-reverse shown below. In addition to the substitution, these two primers contain a silent mutation that removes an Xho I restriction site, a feature that can be used to quickly distinguish mutants from the original template. Plasmid PS535 is deposited under the Budapest Treaty with Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ) on 17 April 2000 with DSM 13451.

Plasmid PS533 (HSPDE4A4-ΔLR2-EGFP) is a deletion mutant of plasmid PS462 (HSPDE4A4-EGFP). In plasmid PS533, 8 amino acid residues comprising the region Ala-313 to Gln-320 in the linker region 2 (LR2) of HSPDE4A4 is deleted using the PCR-based

Quickchange mutagenesis kit (Stratagene, La Jolla). The PCR reaction leading to this deletion uses plasmid PS462 as template and the primers 4AΔLR2-forward and 4AΔLR2-reverse shown below. In addition to the deletion, these two primers introduce an Acc65 I restriction site by a silent mutation, which can be used to quickly distinguish mutants from the original template. Plasmid PS533 is deposited under the Budapest Treaty with Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ) on 17 April 2000 with DSM 13452.

4AH-N-forward (SEQ ID NO: 15):

10 5'-GATGAGTCGGTGCTCGAAAATCACAACTGGCCGTGGGCTTCAAGCTGC

4AH-N-reverse (SEQ ID NO: 16):

5'-GCAGCTTGAAGCCCACGGCCAGGTTGTGATTTTCGAGCACCGACTCATC

4AΔLR2-forward primer (SEQ ID NO: 17):

5'-CCCATCACCCACGATGAAGGAACGAGAAAAACAGCAACCGCCCCCGCCCCCGGT
15 ACCACACTTACAGCCC

4AΔLR2-reverse primer (SEQ ID NO: 18):

5'GGGCTGTAAGTGTGGTACCGGGGGCGGGGGCGGTTGCTGTTTTTCTCGTTCCTTCATCGTG
GGTGATGGG

20 To construct HSPDE4A4catD-EGFP fusion, a 5'-end fragment of HSPDE4A4 and the common 3'-end catalytic region of HSPDE4D isoforms are amplified separately by PCR and ligated into pEGFP.

The 5'-end of HSPDE4A4 (nucleotides 1-1023; GenBank accession number L20965) is amplified by PCR in the presence of 5% DMSO using plasmid PS462 (HSPDE4A4 in
25 pEGFP) as template and primers 4A4-EcoRI-top and 4A4-HindIII-bottom described below.

The 4A4-BamHI-top primer contains an EcoRI restriction site followed by a Kozak sequence and the ATG of HSPDE4A4. The 4A4-HindIII-bottom primer contains a HindIII site that has been introduced into the HSPDE4A4 sequence of the primer by two silent mutations (underlined in the primer sequence). These mutations preserve correct
30 translation of the protein and allow ligation to the HSPDE4Dcat fragment.

For the amplification of the common 3'-end catalytic region of HSPDE4D isoforms, HSPDE4Dcat, the HSPDE4D3 isoform (GenBank accession number L20970) is amplified from a pool of placenta and fetal brain cDNA (Clontech, Palo Alto) using the primers 9855 and 9858 described below. Primer 9855 contains a HindIII restriction site and nucleotides
35 specific for the 5'-end of HSPDE4D3 including the ATG start codon. Primer 9858 contains

an EcoRI restriction site and nucleotides specific for the 3'-end of HSPDE4D except for the stop-codon. This fragment is digested with HindIII and EcoRI and ligated into the corresponding sites in pEGFP (Clontech, Palo Alto). The resulting plasmid is termed PS449. From this plasmid, the 3'-end catalytic region of HSPDE4D isoforms (nucleotides

5 700-2019 of HSPDE4D3 cDNA; GenBank accession number L20970) is amplified by PCR using primers 4Dcat-HinDIII-top and 4Dcat-SacII-bottom as described below. The 4Dcat-HinDIII-top contains a HindIII restriction site that has been integrated into the HSPDE4D sequence by two silent mutations (underlined in the primer sequence). The 4Dcat-SacII-bottom primer contains the 3'-end sequence of PDE4D isoforms except for the stop-

10 codon in order to make fusion to EGFP possible.

The HSPDE4A4 5'-end fragment is digested with EcoRI and HindIII. The HSPDE4Dcat fragment is digested with HindIII and SacII. These two fragments are ligated in a three-part ligation into pEGFP (Clontech, Palo Alto; GenBank accession number U55762) digested with EcoRI and SacII. This results in an HSPDE4A4catD-EGFP fusion under

15 control of a CMV promoter. The plasmid is referred to as PS687.

4A4-EcoRI-top (SEQ ID NO: 19)

5'-CCGGAATTCGCCCATGGAACCCCGACCGTCCCCTC

4A4-HindIII-bottom (SEQ ID NO: 20)

20 5'-GGCAAGCTTTTTCAACCCTGTGATTTGGGACATGGGCTGTAAGTG

4Dcat-HinDIII-top (SEQ ID NO: 21)

5'-GGCAAGCTTATGCACAGCTCTAGTCTGACTAATTCAAGTATCCCAAGGTTTGG

4Dcat-SacII-bottom (SEQ ID NO: 22)

5'-GCCCCGCGGCGTGTCTCAGGAGAACGATCATCTATGACACAGGCTTCAGGC

25 9855 (SEQ ID NO: 27):

5'-GTAAGCTTGCGAACATGATGCACGTGAAT

9858 (SEQ ID NO: 28):

5'-GTGAATTCCTCGTGTCTCAGGAGAACGATCAT

30 Plasmid PS716 expresses a fusion between PDE4A1 and a E222G derivative of GFP, and plasmid PS717 expresses a fusion between PDE4A4 and a E222G derivative of GFP. They were made by replacing a ca. 0.8 kb BamH1-Xba1 fragment containing EGFP sequence with a similar fragment containing the E222G derivative of GFP from plasmid PS699.

35 PS699 was constructed as described below.

Construction of GFP plasmid combining F64L and E222G and mammalian codon usage:

Plasmid pEGFP (GenBank accession number U55762) contain a derivative of GFP in
 5 which one extra amino acid has been added at position two to provide a better
 translational start sequence (a Kozak sequence) and so the total number of amino acids
 is increased by one to 239 instead of the 238 found in wildtype GFP. Therefore the
 denomination of mutations in GFP in these plasmids strictly should be referred to as e.g.
 F65L rather than F64L. However, to avoid this source of confusion and because the GFP
 10 community has adopted the numbering system of wildtype GFP in its communications, the
 numbers used here conform to the commonly used naming of mutations in wildtype GFP.
 The relevant mutations in this respect are F64L, S65T, and E222G.

Plasmid pEGFP contains the following mutations in the chromophore: F64L and S65T.
 The codon usage of the GFP DNA sequence has been optimized for expression in
 15 mammalian cells. N1 and refers to the position of multiple cloning sites relative to the
 GFP sequence.

To construct a plasmid combining F64L and E222G, pEGFP is first subjected to PCR with
 primers 9859 and 9860 described below. The primers are complementary to the DNA
 sequence around the chromophore region and introduce a point mutation changing the
 20 threonine at position 65 to serine. In addition the primers introduce a unique Spe1
 restriction site by silent mutation. The 4.7 kb PCR product is digested with Spe1,
 religated, and transformed into E.coli. The resulting plasmid is referred to as PS399. This
 plasmid contains the chromophore sequence 64-LSYG-67. Plasmid PS399 is subjected to
 Quick-Change mutagenesis (Stratagene) employing PCR with primers 0225 and 0226
 25 described below. These primers are complementary to sequences near the C-terminus of
 the GFP and change glutamate at position 222 to glycine, and in addition they introduce
 an Avr2 restriction site by silent mutation. The resulting plasmid is referred to as PS699. It
 combines an LSYG chromophore with E222G.

9859-top (SEQ ID NO: 33): 5'-TGTA CTAGTGACCACCTGTCTTACGGCGTGCA-3'
 30 9860-bottom (SEQ ID NO: 34): 5'-CTGACTAGTGTGGGCCAGGGCACGGGCAGC-3'
 0225-bottom (SEQ ID NO: 35):
 5'-CCCGGCGGCGGTCACGAACCCTAGGAGGACCATGTGATCGCG-3'
 0226-top (SEQ ID NO: 36):
 5'-CGCGATCACATGGTCCTCCTAGGGTTCGTGACCGCCGCGGG-3'

Table 3

Name	Nucleotide sequence	Predicted amino acid sequence
HSPDE4A1-EGFP	SEQ ID NO: 1	SEQ ID NO: 2
HSPDE4A4-EGFP	SEQ ID NO: 3	SEQ ID NO: 4
HSPDE4A4-delLR2-EGFP	SEQ ID NO: 5	SEQ ID NO: 6
HSPDE4A4-H506N-EGFP	SEQ ID NO: 7	SEQ ID NO: 8
HSPDE4A4catD-EGFP	SEQ ID NO: 23	SEQ ID NO: 24
HSPDE4D3-EGFP	SEQ ID NO: 25	SEQ ID NO: 26
HSPDE4A1-E222G	SEQ ID NO: 29	SEQ ID NO: 30
HSPDE4A4-E222G	SEQ ID NO: 31	SEQ ID NO: 32

Example 2: In vivo expression, visualisation and measurement of changes undergone by the probes

5 Transfection and cell culture:

Chinese hamster ovary cells (CHO), are transfected with the plasmids described in Example 1 above using the transfection agent FuGENE™ 6 (Boehringer Mannheim Corp, USA) according to the method recommended by the suppliers. Stable transfectants are selected using 1 mg/ml G418 sulphate (Calbiochem) in the growth medium (HAM's F12

- 10 nutrient mix with Glutamax-1, 10 % foetal bovine serum (FBS), 100 µg penicillin-streptomycin mixture ml⁻¹ (GibcoBRL, supplied by Life Technologies, Denmark). Cell are cultured at 37 °C in 100% humidity and conditions of normal atmospheric gases supplemented with 5% CO₂.

- 15 Clonal cell lines with particular properties are subcultured from mixed populations of stably transfected cells by isolating individual cells and removing them to sterile culture flasks containing fresh culture medium with 1 mg/ml G418 sulphate.

- For fluorescence microscopy, cells are allowed to adhere to Lab-Tek chambered coverglasses (Nalge Nunc International, Naperville USA) for at least 24 hours and are then cultured to about 80% confluence. Cells can also be grown in plastic 96-well plates
- 20 (Polyfiltronics Packard 96-View Plate or Costar Black Plate, clear bottom; both types tissue culture treated) for imaging purposes. Prior to experiments, the cells are cultured over night without G418 sulphate in HAM F-12 medium with glutamax, 100 µg penicillin-streptomycin mixture ml⁻¹ and 10 % FBS. This medium has low autofluorescence enabling fluorescence microscopy of cells straight from the incubator. For certain tests requiring
- 25 medium of defined composition, particularly with regard to the presence of specific cell

growth factors, the HAM's culture medium is replaced prior to imaging with a buffered saline solution (KRW buffer) containing (in mM) 3.6 KCl, 140 NaCl, 2 NaHCO₃, 0.5 NaH₂PO₄, 0.5 MgSO₄, 1.5 CaCl₂, 10 Hepes, 5 glucose, pH7.4.

Confocal imaging:

- 5 Confocal images are collected using a Zeiss LSM 410 microscope (Carl Zeiss, Jena, Germany) equipped with an argon ion laser emitting excitation light at 488 nm. In the light path are a FT510 dichroic beamsplitter and a 515 nm long-pass filter or a 510 to 525 nm bandpass emission filter. Images are typically collected with a Fluar 40X, NA: 1.3 oil immersion objective, the microscope's confocal aperture set to a value of 10 units
- 10 (optimum for this lens). Typical regions of CHO cells containing HSPDE4A1-EGFP, HSPDE4A4-EGFP, HSPDE4A4-ΔLR2-EGFP and HSPDE4A4-H506N-EGFP probes are shown in Figs 1, 2, 3, and 4 respectively.

Time lapse sequences and analysis:

- Image sequences of live cells over time (time lapse) are gathered using a Zeiss Axiovert
- 15 135M fluorescence microscope fitted with a Fluar 40X, NA: 1.3 oil immersion objective and coupled to a Photometrics CH250 charged coupled device (CCD) camera (Photometrics, Tucson, AZ USA). The cells are illuminated with a 100 W HBO arc lamp. In the light path are a 470±20 nm excitation filter, a 510 nm dichroic mirror and a 515±15 nm emission filter for minimal image background. The cells are maintained at 37°C with a
- 20 custom built stage heater.

- Time lapse response profiles are extracted from image sequences using a region of interest (ROI) defined over the same co-ordinates or pixels for each successive image in a sequence: pixel values are summed and averaged over the ROI in each image, and the resulting values plotted against image number to generate a time lapse response profile
- 25 for that defined region of the sequence. A ROI can include many cells, a single cell, or a region within a single cell.

Automated imaging and analysis:

- Changes in cellular distribution of a transfected probe can be imaged and quantified in an automated fashion. For this purpose cells are cultured to near 80% to 90% confluence in
- 30 coverglass chambers or plastic 96-well plates, given the relevant treatment and allowed to respond. At the end of the response period, cells are fixed in 4% formaldehyde buffer (Lillies fixative buffer, pH7.0: Bie and Berntsen A/S, Denmark) for 30 minutes to 2 hours, then washed in phosphate buffered saline (PBS, Life Technologies, Denmark). Nuclear DNA is stained with 1 μM Hoechst 33258 (Molecular Probes, Eugene, Oregon, USA) in
- 35 PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected

on a Nikon Diaphot 300 (Nikon, Japan) using a Nikon Plan Fluor 20X/0.5NA objective lens. The basic microscope is fitted with a motorised specimen stage and motorised focus control (Prior Scientific, Fulbourn, Cambridge UK), excitation filter wheel (Sutter Instruments, Novato CA USA) and Photometrics PXL series camera with a KAF1400 CCD chip (Photometrics, Tucson, AZ USA), each of these items being under the control of an Macintosh 7200/90 computer (Apple Computer, Cupertino, CA USA). Automation of stage positioning, focus, excitation filter selection, and image acquisition is performed using macros written in-house, running under IPLab Spectrum for Macintosh (Scanalytics, Fairfax, VA USA). Fluorescence illumination comes from a 100 W HBO lamp. Images are collected in pairs, the first using a 340/10 nm excitation filter, the second with a 475RDF40 excitation filter (Chroma, Brattleboro, Vermont). Both images are collected via the same dichroic and emission filters, which are optimised for EGFP applications (XF100 filter set, Omega Optical, Brattleboro, Vermont). While the choice of filters for imaging the nuclear stain (Hoechst 33258) is not well matched to that dye's spectral properties, resulting in lower image intensity, it greatly improves the throughput of the procedure by allowing both images to be collected using the same dichroic and emission filter. This eliminates any image registration problems and focus shifts which would result from using two different filter sets, which would require more steps in the acquisition procedure and more extensive image processing to overcome.

20

The necessary images are collected as follows: A holder containing four 8-well coverglass chambers, or a single 96-well plate, is loaded onto the microscope. The program is started, and the first well of cells is moved into position and manually coarse-focused by the operator. The image is fine-focused by an auto-focus routine using the 340/10 excitation. An image is captured and stored at this excitation wavelength (the nuclear image), and then a second image is captured and stored at the longer wavelength excitation (the GFP image). The stage is automatically repositioned and microscope automatically refocused to capture a second pair of images within the same well. This process is repeated a set number of times (typically 4 to 8) for the first well. The stage then advances the next well to the imaging position, and the process repeats itself until the set number of image pairs has been captured from each well of cells.

Image pairs are automatically analysed in the following way using a suite of macros running under the IPLab Spectrum software: First the nuclear image of a pair is filtered with a digital filter to simultaneously sharpen the edges of and suppress differences in

intensity of the nuclei. The choice of filter, and the filter constants, were arrived at through experimentation with various data sets. The filtered image is then segmented at a pre-determined intensity value, such that pixels below this threshold are very likely not within a nuclear region, and pixels above this threshold are very likely within a nuclear region.

- 5 The contiguous regions above the threshold are then counted, after rejecting contiguous regions that are larger than a certain area or smaller than a certain (different) area, the areas having been previously determined to provide a sufficiently accurate distinction between nuclei and other objects that are not nuclei. The final count is the estimated number of nuclei in the field. The GFP image of each pair is then digitally filtered with a
- 10 filter chosen experimentally to suppress the variation of intensity due to the typical non-localised distribution of GFP, while accentuating the intensity of any bright point-like objects relative to this background. This filtered image is then segmented at a threshold that has been experimentally determined to divide the image into pixels that are very likely to be in a spot (above the threshold) and pixels that are very likely not to be in a spot
- 15 (below the threshold). The contiguous regions of pixels that are above the threshold are counted, after rejecting regions that do not have certain morphological properties which were previously determined to be characteristic of spots. The ratio of spot count to nuclear count for each pair represents an estimate of the average number of spots per cell in that image pair. All image pairs are treated in this way, and the final table of values is used to
- 20 establish the cellular response to a given treatment. Data derived from automated imaging experiments are shown in Figs. 15 to 30, and 35 to 37.

Example 3: Redistribution of probe HSPDE4A4-EGFP caused by Rolipram treatment

- 25 This example illustrates how Rolipram affects the physical properties and behaviour of the HSPDE4A4-EGFP probe as expressed in stably transfected CHO cells.
Stably transfected (non-clonal) cells are allowed to adhere to chambered coverglasses in HAM's F12 medium containing 10% FBS. Once adhered, 2 μ M Rolipram (4-(3-cyclopentyloxy-4-methoxyphenyl)-2-pyrrolidone; Calbiochem) is added to the medium, and
- 30 cells incubated further at 37 °C with 5% CO₂ + air. At certain times after addition of Rolipram cells are checked on a fluorescence microscope for changes in cellular distribution of GFP fluorescence.

These experiments show that the general cytoplasmic distribution of fluorescence in most cells gradually changes to one consisting of bright concentrations of fluorescence located

at several distinct spots within the cytoplasm, some fluorescence remaining more evenly distributed within the rest of the (non nuclear) cytoplasm. A common pattern appears to be the presence of only 2 major accumulations of fluorescence diametrically separated across the nucleus of the cell (Fig. 5). Spots are stable in cells as long as Rolipram

5 continues to be present.

Spots begin to be visible about 3 hours after addition of 2 μ M Rolipram. The effect is qualitatively similar at concentrations spanning the range 100 μ M to 0.5 μ M Rolipram.

Pre-treatment of cells with 5 μ g/ml cycloheximide prevents formation of spots induced by Rolipram, indicating that protein synthesis is a necessary part of spot formation. Once

10 spots have formed, removal of Rolipram results in their rapid dissolution, within 60 minutes at 37 °C. However, replacement of Rolipram causes the bright spots to reform, also within 60 minutes. This is more rapid than is seen for *de novo* production of spots by Rolipram in these cells.

These experiments indicate that the spots are built around anchor proteins that require
15 time to be synthesised and to accumulate. Rolipram treated PDE4 appears to be necessary for this accumulation to occur. Once the accumulations of anchor proteins are formed, they remain stable within the cells, for periods of at least 4 hours.

Fig. 6 shows the homogenous response to 2 μ M Rolipram of a clonal population of cells derived from a single progenitor cell transfected with probe HSPDE4A4-EGFP – more
20 than 95% of cells have produced bright spots after 6.7 hours of exposure to Rolipram. The presence of 10% FBS is not necessary for the formation of bright spots in response to Rolipram treatment (Fig. 6).

Example 4: Redistribution of probes HSPDE4A4- Δ LR2-EGFP and

25 **HSPDE4A4-H506N-EGFP caused by Rolipram treatment**

This example illustrates how Rolipram affects the physical properties and behaviour of the HSPDE4A4- Δ LR2-EGFP and HSPDE4A4-H506N-EGFP probes as expressed in stably transfected CHO cells. The change in behaviour of the probe(s) is easily measurable by means of fluorescent imaging and allows this method to be used in the search for

30 compounds that have similar properties to the PDE4 inhibitor Rolipram. Comparison of the behaviours of the 4A4 variants to that of the wild-type enzyme indicates which regions of the molecule are important in effecting the Rolipram response.

Stably transfected (non-clonal) cells are allowed to adhere to chambered coverglasses in HAM's F12 medium containing 10% FBS. Once adhered, 2 μ M Rolipram (Calbiochem) is

added to the medium, and cells incubated further at 37 °C with 5% CO₂ + air. At certain times after addition of Rolipram cells are checked on a fluorescence microscope for changes in cellular distribution of GFP fluorescence.

Cells transfected with the HSPDE4A4-ΔLR2-EGFP probe had the same initial appearance 5 (Fig. 3) as those transfected with the HSPDE4A4-EGFP probe, and subsequently reacted to treatment with 3 μM Rolipram in a similar way to those transfected with the HSPDE4A4-EGFP probe (Fig. 7), indicating that deletion of the region of the enzyme designated LR2 did not ablate the Rolipram response. Cells transfected with the HSPDE4A4-H506N-EGFP probe had the appearance shown in Fig. 4. Subsequent 10 treatment with 100 μM Rolipram for 23.5 hours in HAM's F12 + 10% FBS causes spots to appear in only about 15% of the cells (Fig. 8). This result indicates that the histidine at position 506 in the primary sequence of the protein, located in the catalytic cleft of the enzyme, is essential to the Rolipram response. Mutation of this histidine is known to leave the Rolipram binding affinity of a shortened version of a recombinant human PDE4A 15 enzyme virtually unchanged (Jacobitz *et al.*, 1997), although the K_m of the enzyme for cAMP increases 11 fold, and hence its activity decreases by approximately 90%. The conclusion can therefore be drawn that the binding of Rolipram within the catalytic cleft of the enzyme initiates the change in the cellular distribution and behaviour of the HSPDE4A4-EGFP and HSPDE4A4-ΔLR2-EGFP probes via a mechanism that is 20 dependent on the presence of this critical histidine residue at position 506. The ability of the enzyme to simultaneously bind cAMP and Rolipram may also be important in the formation of spots in cells.

Example 5: Effects on the cellular distribution of HSPDE4A4-EGFP of 25 treatments with several compounds known either to inhibit PDE4 enzymes directly or processes that are known to be inhibited by Rolipram

This example illustrates how various compounds with either general or specific inhibitory activity against PDEs, and/or with proven anti-inflammatory or anti-depressive properties, affect the physical properties and behaviour of the HSPDE4A4-EGFP probe as expressed 30 in stably transfected CHO cells. The results of these experiments show how changes in cellular distribution of the HSPDE4A4-EGFP probe following treatment with a compound can be used to predict or evaluate the biological activity and therapeutic consequences of administering that compound to mammals, especially to humans.

Stably transfected (clonal and non-clonal) CHO cells are allowed to adhere to chambered coverglasses in HAM's F12 medium containing 10% FBS. Once adhered, compounds are added singly to each test chamber, and cells incubated further at 37 °C with 5% CO₂ + air. At certain times after addition of the compounds cells are checked on a fluorescence

- 5 microscope for changes in cellular distribution of GFP fluorescence. Different compounds cause different changes in the pattern of GFP fluorescence in these cells.

The compounds trequinsin (HL-725; 9,10-dimethoxy-2-mesitylimino-3-methyl-3,4,6,7-tetrahydro-2H-pyrimido[6,1-a]isoquinolin-4-one) and etazolate (SQ20009; 1-ethyl-4-[(1-methylethylidene)hydrazino]-1H-pyrazolo[3,4-b]pyridine-5-carboxylic acid ethyl ester, HCl;

- 10 Calbiochem) both have some inhibitory activity towards PDE4s, although they are not specific inhibitors of these enzymes; IC₅₀ values of trequinsin against PDE4 is 1 µM (Saldou *et al.*; 1998) and of etazolate is 2 µM (Ahluwalia *et al.*; 1982). Neither compound causes formation of spots in clonal CHO cells transfected with the HSPDE4A4-EGFP probe.

- 15 Denbufylline (BRL 30892, Beecham), a selective PDE4 inhibitor (IC₅₀ 1 µM against PDE4) which is structurally unrelated to Rolipram, produces spots in cells transfected with the HSPDE4A4-EGFP probe (Fig. 9). The spots it produces are indistinguishable from those induced by Rolipram treatment, although it is less effective; only 40% of cells develop spots after 24.5 hours with 10 µM of the compound.

- 20 RS 25344 is a specific inhibitor of PDE4 enzymes, also structurally unrelated to Rolipram, which has an IC₅₀ of 0.00028 µM against PDE4s (Saldou *et al.*; 1998). At 0.03 µM RS25344, spots appeared in about 40% of clonal cells transfected with the HSPDE4A4-EGFP probe after 24.5 hours (Fig 10a), the spots being indistinguishable from those caused by Rolipram. Under similar conditions, 1 µM of the compound produces very large
25 and bright spots in more than 95% of cells (Fig 10b).

RP 73401 (also known as Piclamilast; Rhone-Poulenc Rorer), a specific inhibitor of PDE4 enzymes, produces no spots in clonal cells transfected with the HSPDE4A4-EGFP probe when tested over the range 0.3 nM to 3 µM. The IC₅₀ of RP73401 for PDE4 is 0.3 nM (Saldou *et al.* 1998). Rolipram (2µM) plus 0.001 µM RP73401 will produce spots in less

- 30 than 5 % of cells (Fig. 11b) 7.5 hours. Under similar conditions, but without RP73401, more than 95% of the same cells respond to 2 µM Rolipram by producing bright spots in their cytoplasm (Fig 11a). With 0.003 µM RP73401, 2 µM Rolipram is unable to elicit the production of any spots in these cells (Fig 12). Furthermore, spots take over 4 hours to appear following subsequent replacement of the [Rolipram + RP73401] with 2 µM

Rolipram alone, indicating that no anchor proteins accumulate in the combined presence of Rolipram and RP73401.

The Rolipram-like compound Ro-20-1724 (Calbiochem) is a specific inhibitor of PDE4 enzymes with an IC_{50} of 2 μ M (Rubin *et al.*; 1991). At 10 μ M Ro-20-1724 spots appear in about 80% of clonal cells transfected with the HSPDE4A4-EGFP probe after 4.5 hours (Fig 13), the spots being indistinguishable from those caused by Rolipram.

Incubation of non-clonal cells with 500 μ M of the non-selective PDE inhibitor IBMX (Sigma Aldrich) causes spots to become visible in only about 5 to 10% of cells after 14 hours incubation. These spots are rather smaller and more numerous within each cell than those formed in the presence of Rolipram (Fig 14). IBMX is a general inhibitor of all PDEs, and its presence will therefore encourage cAMP levels to rise in treated cells, which is not the case for PDE4 selective inhibitors which leave the activity of other families of PDEs unaffected.

CHO cells transfected with the HSPDE4A4-EGFP probe do not produce spots when treated, in HAM's F12 with or without 10% FBS, with 500 μ M theophylline (a general PDE inhibitor), or 100 μ M caffeine (a weak and general PDE inhibitor), or 10 μ M milrinone (a strong PDE3 inhibitor but also reported to have IC_{50} for PDE4s of about 10 μ M), or 0.5 μ M cilostamide (a potent PDE3 inhibitor, IC_{50} 70 nM), or 100 μ M zaprinast (potent PDE5 inhibitor, IC_{50} 0.4 μ M), or 400 μ M thalidomide (an anti-inflammatory compound with unspecified mode of action); all these incubations are carefully observed over the period of 1 to 24 hours and none produces spots. Cells treated with 2 μ M Rolipram plus either theophylline, caffeine, milrinone, cilostamide or zaprinast (same concentrations as previously, same treatment times and conditions) form the same number and type of bright spots as they do when treated with 2 μ M Rolipram alone.

Together, these experiments where PDE inhibitors are simply incubated with cells transfected with the HSPDE4A4-EGFP probe show that the formation in these cells of spots similar to those produced by Rolipram is apparently associated only with the particular sub-class of PDE inhibitors that are potent and specific inhibitors of PDE4. The example shows how screening of compounds for their ability to form spots in these cells can be used to identify PDE4 inhibitors, and that the compounds identified will have similar properties to Rolipram. Further, the example shows how the HSPDE4A4-EGFP-transfected cells can be used to screen for compounds that will prevent Rolipram from forming spots, and that these compounds so identified, such as RP73401, will also be potent and specific inhibitors of PDE4 with certain properties different to those of Rolipram.

Example 6: Quantitative assessment of the effects of Rolipram, RS25344 and Ro 20 1724 on the cellular distribution of HSPDE4A4-EGFP probe in CHO cells.

- 5 This example shows how the number of spots per cell in CHO cells transfected with the HSPDE4A4-EGFP increases in a dose dependent fashion with certain PDE4-specific inhibitors, that this quantity is readily measurable by automated imaging, and that the dose response data from such measurements yield EC₅₀ values that are closely similar to the biological effectiveness of these compounds in therapeutic applications.
- 10 Fig. 15 shows dose response curves for spot formation in response to three different PDE4 inhibitors for a stable and clonal CHO cell line transfected with HSPDE4A4-EGFP probe (spot assays). The three inhibitors are Rolipram (▼), RS25344 (■) and Ro 20-1724 (●). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements ± sem, where each measurement is itself an average taken
- 15 from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 µM Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Data
- 20 sets are fitted to a 4-parameter Hill equation giving the following EC₅₀ values of 0.34 micromolar for Rolipram, 0.017 micromolar for RS25344 and 3.77 micromolar for Ro 20-1724.

Example 7: Quantitative assessment of the effects of Rolipram on the
25 **cellular distribution of HSPDE4A4-EGFP, HSPDE4A4-ΔLR2-EGFP and**
HSPDE4A4-H506N-EGFP probes in CHO cells.

- This example shows how measurement of the number of spots per cell induced by different concentrations of Rolipram, in CHO cells transfected with various N1 fusions of EGFP to wild-type and mutant forms of HSPDE4A4, can be used to quantify the
- 30 importance of different amino acids in the primary sequence of the enzyme in the sensitivity of the Rolipram response.

Fig. 16 shows dose response curves for spot formation in response to Rolipram in three stable and clonal cell lines of CHO cells transfected with HSPDE4A4-EGFP (●),

HSPDE4A4- Δ LR2-EGFP (∇) and HSPDE4A4-H506N-EGFP (\blacktriangledown). The number of spots per cell for each concentration is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of Rolipram for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Data sets are fitted to a 4-parameter Hill equation giving the following EC₅₀ values of 0.34 micromolar Rolipram for the HSPDE4A4-EGFP probe and 0.41 micromolar Rolipram for the HSPDE4A4- Δ LR2-EGFP probe. An EC₅₀ value can not be determined for the HSPDE4A4-H506N-EGFP probe, since the mutation makes it almost unresponsive to Rolipram in this clonal cell line.

The example shows that when 8 amino acid residues comprising the region Ala-313 to Gln-320 in the linker region 2 (LR2) of HSPDE4A4 are deleted, Rolipram-induced spot formation is not significantly changed relative to that of the wild-type probe. However, mutation of histidine 506 to asparagine (H506N) produces an almost total loss of sensitivity to Rolipram, indicating that this is an essential residue in the protein for transducing the spot formation activated by Rolipram.

Example 8: Quantitative assessment of the effects of Rolipram, RS25344 and Ro 20-1724 on the cellular distribution of HSPDE4A4-H506N-EGFP probe in CHO cells.

This example shows how the number of spots produced per cell by different PDE4 inhibitors in CHO cells transfected with the HSPDE4A4-H506N-EGFP is useful in discovering compounds that interact with an ensemble of amino acid residues in HSPDE4A4 that are different to those with which Rolipram interacts.

Fig. 17 shows dose-response curves for spot formation in response to three different PDE4 inhibitors for a stable and clonal CHO cell line transfected with the HSPDE4A4-H506N-EGFP probe. The three inhibitors are Rolipram (\blacktriangledown), RS25344 (\blacksquare) and Ro 20-1724 (\bullet). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst

33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The data set for RS25344 is fitted to a 4-parameter Hill equation giving an EC₅₀ value of 0.125 micromolar. These clonal cells containing the H506N mutant of HSPDE4A4-EGFP are

5 almost unresponsive to the other two inhibitors over the concentrations tested.

The data show that RS25344 is very different to Rolipram and Ro 20-1724 in that it does not require the presence of a histidine at position 506 to effect the production of spots, and this indicates that the RS compound interacts with an ensemble of amino acids that are different to those with which Rolipram and Ro 20-1724 interact. A spot production

10 assay using the HSPDE4A4-H506N-EGFP probe is therefore able to identify other compounds that differ in this respect from Rolipram and Rolipram-like compounds.

Example 9: Quantitative assessment of the effects of RP73401 on the ability of Rolipram to produce spots in CHO cells transfected with the HSPDE4A4-EGFP probe.

This example shows that the spot assay can be run in a competitive way to identify compounds that are specific PDE4 inhibitors which interfere with the ability of Rolipram to form spots, and that the spot assay can be used to quantify the competitive strength of such compounds.

20 Fig. 18 shows a competitive dose-response curve for Rolipram-induced spot formation in a stable and clonal CHO cell line transfected with HSPDE4A4-EGFP probe. The cells are challenged with a fixed concentration of 2 µM Rolipram and varying concentrations of the specific PDE4 inhibitor RP73401 (Piclamilast). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements ± sem, where
25 each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of RP73401 plus 2 µM Rolipram for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 µM Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the
30 number of spots per cell as described in Example 2.

Approximately 0.003 µM RP73401 is sufficient to inhibit 50% of the spot formation response of these cells that normally results from treatment with 2 µM Rolipram.

Example 10: Effect upon Rolipram-induced distribution of HSPDE4A4-EGFP following treatment with 3-isobutyl-1-methylxanthine (IBMX) plus forskolin measured as time response profiles in low and high throughput.

This example illustrates how the Rolipram-stimulated accumulations, or large spots, of the HSPDE4A4-EGFP probe may be mobilised and dispersed through the action of compounds that increase cAMP levels in transfected cells. Such treatments may be useful as positive controls in screening assays designed to search for novel compounds able to dislocate bound forms of the HSPDE4A4-EGFP probe. The example also illustrates how changes in the distribution of the HSPDE4A4-EGFP probe may be quantified with standard imaging techniques using a fluorescence microscope or a Fluorescence Imaging Plate Reader device (FLIPR, Molecular Devices, Sunnyvale, California, USA). Furthermore, the example provides some evidence for the involvement of cAMP-dependent protein kinase in the dispersal of Rolipram-induced spots. This example also suggests that either assays of PKA activity or of cellular cAMP concentration are useful as secondary screens in conjunction with this PDE-dislocation assay based on the dispersal of Rolipram-induced spots, to rule out compounds causing dispersal through elevation of cAMP and possible activation of PKA.

For the microscopic assessment of spot dispersal, stably transfected (non-clonal) cells transfected with the HSPDE4A4-EGFP probe are allowed to adhere to chambered coverglasses in HAM's F12 medium containing 10% FBS. Once adhered, 2 μ M Rolipram is added to the medium, and cells incubated further at 37 °C with 5% CO₂ + air until spots form in about 80% of cells (approx. 12 hours or more). Chambered coverglasses are then transferred to a Zeiss 135 inverted microscope for time lapse imaging as described in Example 2, above. At a given point after starting an experiment, a mixture of IBMX and forskolin is added to give final concentrations of 1 mM and 50 μ M, respectively. Images are captured at regular intervals to form a time lapse sequence of the response, also as described in Example 2. Individual frames from the sequence are shown in Figs 19 a, b, c and d. Analysis of the sequence generated the response profiles shown in Figs 20a and b. Fig. 20a shows how the fluorescence intensity increases in the cytoplasm with time following application of forskolin plus IBMX; contemporaneously, the fluorescence intensity of each bright spot decreases (Fig. 20b). The average intensity for the entire image does not change significantly over the period of the time lapse sequence (data not shown). These measurements taken together confirm that the bright spots disperse into the cytoplasm under the influence of forskolin plus IBMX, treatments that serve to elevate cytosolic levels of cAMP.

For the FLIPR measurements, clonal CHO cells transfected with the HSPDE4A4-EGFP probe are cultured in a 96-well black microtitre plate (Packard Polyfiltronics ViewPlate-96, Packard Instrument Co.) to near confluency, then treated with 2 μ M Rolipram for 24 hours. The plate is washed with KRW buffer plus 2 μ M Rolipram. Half the plate is treated with 2 μ M H-89 (Calbiochem), a kinase inhibitor especially potent against the cAMP-dependent protein kinases (PKA, IC_{50} about 50 nM) and incubated a further 20 minutes. The plate is then run in the FLIPR system at 37 °C, with addition of IBMX and forskolin to all wells at final concentrations of 500 μ M and 50 μ M respectively after the first minute. The experiment is continued for a further 45 minutes, readings being taken at intervals of 1 minute. Curves A and B in Fig. 21 represent averages over 8 wells each for responses to IBMX and forskolin, where wells for curve B are treated with compound H-89, and those for curve A are not.

The difference in the levels of response indicates that the inhibitor of PKA has a significant effect on the dispersal of spots induced by elevated cAMP, suggesting a role for PKA in this process.

A PKA-GFP redistribution assay, or SPA-based assay of cAMP, would be useful adjuncts to the Rolipram-induced spot-dispersal assay based on the HSPDE4A4-EGFP probe since they could counterscreen compounds that induced dispersal through elevation of cAMP.

Example 11: Effect upon Rolipram-induced distribution of HSPDE4A4-EGFP following treatment with phorbol-12-myristate-13-acetate (PMA) and/or ionomycin

This example illustrates how the Rolipram-stimulated accumulations, or large spots, of HSPDE4A4-EGFP probe may be mobilised and dispersed through the action of compounds that increase the concentration of cytosolic calcium ($[Ca^{2+}]_{cyt}$) and activate C-type protein kinases (PKC) in transfected cells. Such treatments may be useful as positive controls in screening assays designed to search for novel compounds able to dislocate bound forms of the HSPDE4A4-EGFP probe. This example also suggests that either assays of PKC activity or of changes in $[Ca^{2+}]_{cyt}$ are useful as secondary screens in conjunction with this PDE-dislocation assay based on the dispersal of Rolipram-induced spots, to rule out compounds causing dispersal through elevation of $[Ca^{2+}]_{cyt}$ and possible activation of PKC.

Non-clonal CHO cells stably transfected with the HSPDE4A4-EGFP probe are allowed to adhere to chambered coverglasses in HAM's F12 medium containing 10% FBS and to

grow for 24 hours. After 24 hours, 2 μ M Rolipram (Calbiochem) is added to the medium, and cells incubated at 37 °C with 5% CO₂ + air for a further 42 hours. Approximately 70% of the cells contain bright spots when viewed on a fluorescence microscope.

Different wells are then treated with fresh HAM's F12 medium plus 10% FBS plus 2 μ M

- 5 Rolipram containing either 0.2% DMSO, 200 nM PMA, 2 μ M ionomycin or 200 nM PMA plus 2 μ M ionomycin and returned to the incubator for 55 minutes prior to imaging.

Figures 22, 23, 24 and 25 show that the number of cells with bright spots is unaffected in the DMSO control (70%), reduced to between 20-40% in the ionomycin treatment and that all spots dissolve entirely in the other two treatments.

- 10 In summary, 2 μ M ionomycin alone is capable of dispersing Rolipram-induced spots of the HSPDE4A4-EGFP probe in FBS-replete cells, but slowly and in an incomplete manner, whereas 200 nM PMA, \pm ionomycin, will disperse all spots of HSPDE4A4-EGFP probe rapidly and completely.

A PKC-GFP redistribution assay, and/or assay to detect changes in $[Ca^{2+}]_{cyt}$, for example

- 15 a fluorescence based assay with a cell permeable Ca^{2+} -sensitive probe such as Fura 2-AM or Fluo 3-AM (both available from Molecular Probes, Eugene, Oregon, USA), are useful adjuncts to the Rolipram-induced spot-dispersal assay based on the HSPDE4A4-EGFP probe since they rule out compounds that induce dispersal through elevation of $[Ca^{2+}]_{cyt}$ and/or activation of PKC.

20

Example 12: Effect upon Rolipram-induced distribution of HSPDE4A4-EGFP following treatment with phorbol-12-myristate-13-acetate (PMA) plus ionomycin within serum-depleted cells

This example illustrates that in transfected cells which are serum depleted, the Rolipram-stimulated accumulations, or large spots, of the HSPDE4A4-EGFP probe resist the mobilisation or dispersal that is normally elicited by compounds which either increase the concentration of cytosolic calcium ($[Ca^{2+}]_{cyt}$) and/or activate C-type protein kinases (PKC).

This example demonstrates that accumulation and dispersal of the HSPDE4A4-EGFP probe involves yet another control switch in addition to the Rolipram-, cAMP- and [PMA \pm

- 30 ionomycin]-sensitive behaviours described in the preceding examples. However, this switch affects only the behaviour governed by [PMA \pm ionomycin]. As such, the system has considerable in-built complexity, analysis of which in a drug-screening setting demands assays of the highest information content, and secondary screens able to unambiguously identify compounds with the desired mode of action.

Clonal CHO cells stably transfected with the HSPDE4A4-EGFP probe are allowed to adhere to chambered coverslips in HAM's F12 medium containing 10% FBS and to grow for 5 days without change of medium, which by this time is serum-depleted. After 4 days, 2 μ M Rolipram is added to the same medium, and cells incubated at 37 °C with 5% CO₂ + air for a further 22 hours. Approximately 95% of the cells contain bright spots when viewed on a fluorescence microscope.

Cells are then washed to KRW buffer, with no added FBS but containing 2 μ M Rolipram. Individual wells are then treated with either 50 μ M forskolin plus 500 μ M IBMX or 200 nM PMA plus 2 μ M ionomycin. Spots started to disperse in cells treated with forskolin plus IBMX within 10 to 20 minutes (Figs. 26a and 26b). In cells treated with PMA plus ionomycin there is little or no change in the number or size of spots present in the cells, even after 40 minutes (Figs. 27a and 27b)

In summary, Rolipram-induced spots of probe HSPDE4A4-EGFP resist dispersal by agents that should activate PKC and increase $[Ca^{2+}]_{cyt}$ when cells are depleted of certain substances normally found in foetal bovine serum. Spots in serum depleted cells remain sensitive to dispersal by agents that increase cAMP. Run as a primary screen for compounds that disperse Rolipram-induced spots, the assay does not require a counterscreen for PMA-like compounds or ones that increase $[Ca^{2+}]_{cyt}$ if the cells are serum depleted, as when grown in the same medium for 5 days, without addition of fresh serum, in the manner described in this example. A secondary screen for agents that act in the same manner as [IBMX + forskolin] remains a useful adjunct to such a spot disappearance assay, for example a cell-based screen for increased cAMP.

Example 13: Description of treatments found to cause reappearance of spots in CHO cells transfected with HSPDE4A4-EGFP from which spots have been cleared by the removal of Rolipram, and use of the HSPDE4A4-EGFP probe in an assay to identify compounds which inhibit the reappearance of bright cytoplasmic spots under such conditions.

This example describes conditions found to activate the reappearance of Rolipram-like spots in cells that have previously been treated with Rolipram, but then cleared of spots by removal of Rolipram. The example further shows how the reappearance of spots in cells given these appropriate conditions is sensitive to the presence of thalidomide, and therefore how such an assay can be used to screen for compounds with similar properties.

First, CHO cells stably transfected with probe HSPDE4A4-EGFP are grown in HAM's F12 medium with 10% FBS, and with 2 micromolar Rolipram for 15.5 hours. Bright spots are present in more than 95% of all cells. Rolipram is then washed from the cells, and fresh HAM's F12 + 10% FBS is added. Cells are returned to conditions of 37 °C + 5% CO₂ in humidified air (i.e. standard incubator conditions). After 150 minutes, all GFP-bright spots disappear from cells. Spots are induced to reappear in these ways:

A) NaCl is added to the cells to increase the final concentration of salt in the medium.

The cells in Figs. 28a, 28b and 28c are all in HAM's F12 + 10% FBS to which NaCl has been added to increase the concentration of NaCl by 100 mM. Cells in 28c are additionally treated with 5 micromolar SB203580, a specific inhibitor of p38 mitogen activated protein kinases (p38 MAPK). The cells in Fig. 28b are returned to conditions of 37 °C + 5% CO₂ in humidified air (i.e. standard incubator conditions), while those in 28a and 28c are chilled to 4 °C. After 4 hours of these treatments cells were fixed with 4% formaldehyde pH 7.0 at room temperature for 1 hour, and washed with PBS buffer ready for imaging. Many small GFP-bright spots form in more than 90% of the chilled cells, but of those returned to incubator conditions (Fig 28b) less than 5% of cells contain spots. Chilled and SB 203580-treated cells (Fig. 28c) contain significantly fewer, but larger, bright spots per cell than those in Fig. 28a. Fig. 30 shows the response of these cells to various amounts of NaCl where the concentration of salt in the medium has been increased by 0 mM, 5 mM, 50 mM or 100 mM. A second group of the same cells are treated similarly but with the addition of 5 µM SB203580. All treatments are then chilled, fixed and then stained with 1 µM Hoechst 33258 in PBS for 10 minutes at 25 °C, and washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The number of spots per cell increases in a dose-dependent fashion with increasing salt concentration. SB203580 decreases the number of spots per cell. Examination of the images from the SB203580 treatment (e.g. Fig. 28c) suggests that decreased spot number is accompanied by increased spot size.

B) Cells are left at ambient conditions. The cells in Fig. 29a are pre-treated with Rolipram as above, and washed to remove spots as described. They are then left under ambient conditions (normal air, 22 to 25 °C) rather than in a cell incubator, for a period of 4 hours. During this time the medium evaporates by about 20%, and the pH of the medium shifts from pH6.5 to pH8.1 as the CO₂ in the medium equilibrates with ambient conditions. After 4 hours, spots reappear in the cytoplasm which are indistinguishable from those induced by the original Rolipram treatment. As time

continues the proportion of cells containing spots increases as does the size of spots in cells. Return of cells to the incubator after 4 to 6 hours under ambient conditions results in complete reversal of this effect.

The cells in Fig. 29b are treated according to the protocol described in (B) above, and also given 400 μ M thalidomide at the time of removal of the Rolipram. The thalidomide appears to hasten disappearance of the spots, but also inhibits return of spots under ambient conditions. Fig. 31 is a dose-response curve for this effect, for which a set of cells are treated with a range of thalidomide concentrations at the time of removal of Rolipram. After 4 hours under ambient conditions the cells are fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, and washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The data set for this experiment is fitted to a 4-parameter Hill equation (the curve in Fig. 31) indicating an IC_{50} value of 33 micromolar thalidomide against the reappearance of spots under these conditions.

Thalidomide also inhibits the reappearance of spots under the NaCl treatment protocol. Cells treated with an addition of 100 mM NaCl as described in (A) above produce a mean spot count per cell (\pm sem) of 0.856 ± 0.195 after 2 hours at 4 °C. In similar cells treated with 400 μ M thalidomide at the time of removal of Rolipram, the spot count is 0.364 ± 0.047 after 2 hours at 4 °C.

Spots do not reappear in CHO cells stably transfected with probe HSPDE4A4-EGFP under ambient or chilled and salt-supplemented conditions unless the cells are pre-treated with Rolipram. These observations allow the reappearance of spots to be used as a procedure to screen for compounds similar to thalidomide which cannot, or do not, form spots themselves in CHO cells transfected with probe HSPDE4A4-EGFP, and cannot or do not compete directly with Rolipram in the prevention of spot formation in these cells. Such compounds may share certain properties and therapeutic uses in common with thalidomide and related compounds, many of which are known to have useful anti-inflammatory properties together with mild to strong inhibitory actions against PDE4 enzymes.

Example 14: Effect upon distribution of prob HSPDE4A4-EGFP following treatment with ionomycin within serum-depleted Rolipram-treated cells

This example demonstrates yet another behaviour of the Rolipram-treated HSPDE4A4-EGFP probe, which is restricted to cells grown for long periods in serum-depleted media, or starved of serum in KRW buffer. This behaviour involves only the fluorescence seen more or less evenly distributed within the cytoplasm of Rolipram-treated cells, and does not involve the large fluorescent accumulations characteristic of these cells. This example provides evidence that more than one component may be involved in anchoring the HSPDE4A4-EGFP probe in Rolipram-treated cells, and that direct or indirect sensitivity to changes in $[Ca^{2+}]_{cyt}$ is a characteristic of that component (or components).

Clonal CHO cells stably transfected with the HSPDE4A4-EGFP probe are allowed to adhere to chambered coverglasses in HAM's F12 medium containing 10% FBS and to grow for 4 days without change of medium. Rolipram at 2 μ M is added to the medium 32 hours before the cells are used, and incubation continued at 37 °C with 5% CO₂ + air. At the end of this time approximately 95% of the cells contain bright spots when viewed on a fluorescence microscope.

Cells are then washed to KRW buffer, with no added FBS but containing 2 μ M Rolipram. Individual wells are then treated first with 1 or 2 μ M ionomycin, and then a short time later with 50 μ M forskolin plus 500 μ M IBMX. Within 1 minute of treatment with ionomycin, at either 1 or 2 μ M, small spots form in the cytoplasm of the cells (Fig. 32a and 32b). This occurs in nearly all cells, whether they contain large Rolipram-induced spots or not. Large spots are not affected in any way during the formation of the smaller spots. The small spots will spontaneously disappear within 10 to 20 minutes. Application of forskolin plus IBMX clears them within minutes (Fig. 33a). The larger spots also disperse in time with this treatment, but more slowly (Fig. 33b). The formation of small spots will not occur in serum-replete cells, or in starved cells that then have been given 10% serum for more than 45 minutes. The response does not occur in cells transfected with the HSPDE4A4-EGFP probe if Rolipram is not present. The time course of transient appearance of the small spots of the HSPDE4A4-EGFP probe is consistent with that of the Ca²⁺ transient generally elicited by ionomycin in treated cells.

This example suggests that the Rolipram-inhibited HSPDE4A4-EGFP probe, or some other anchoring component it is attached to in the Rolipram-inhibited state, is Ca²⁺ sensitive. Since there is no obvious effect on the large accumulations, it is also possible that the HSPDE4A4-EGFP probe distributed throughout the cytoplasm is anchored to a different component than that or those found in the large accumulations or spots. The

behaviour of the HSPDE4A4-EGFP probe under these conditions may be used to screen for compounds that remove the calcium sensitivity of the enzyme complex, or which simply prevent the formation of these minor spots. Such compounds may be useful in controlling inflammatory responses of pro-inflammatory cells such as eosinophils and
5 other leucocytes.

Example 15: Redistribution of HSPDE4A1-EGFP caused by treatment with Rolipram

This example shows that the HSPDE4A1-EGFP probe redistributes within cells when
10 treated with Rolipram, but in a way that is very different to the behaviour of the HSPDE4A4-EGFP probe. HSPDE4A1-EGFP accumulates as small perinuclear spots in otherwise untreated CHO cells transfected with the plasmid PS461 (Figs. 1 and 34a). Rolipram induces these spots to disperse into the cytoplasm (Fig. 34b).
The HSPDE4A1-EGFP probe is useful in the search for dislocators of this isoform, and to
15 discover compounds that mimic or antagonise the effect of Rolipram on the probe. Such compounds will likely be therapeutically useful in the treatment of depressive disorders and inflammatory reactions in the central nervous system.
In Fig. 34a the cells are growing in only HAM's F12 medium with 10% FBS; the GFP fluorescence is restricted to bright granule-like spots within the perinuclear cytoplasm of
20 each cell. The spots may be clustered around, in or on the Golgi membranes. In Fig. 34b similar cells to those seen in 34a have been treated with 2 micromolar Rolipram for 2 hours. The majority of GFP-bright spots disappear in all cells under Rolipram treatment, and the cytoplasm becomes generally brighter. Larger spots may not disperse completely in some cells. When Rolipram is washed away, the spots reform within 1.75 hours.
25 Certain other compounds also reduce PDE4A1 spot numbers, and these include Ro 20-1724, RS25344 and to a lesser extent, denbufylline and IBMX, but the latter compound only starts to have an effect after 100 μ M. RP73401 does not disperse spots, and it is anticipated that other such compounds with affinity only for the "low affinity binding site" of PDE4s, such as SB207499 or CDP840 (CellTech/Chiroscience) will also fail to disperse
30 spots of PDE4A1.

This example shows that the HSPDE4A1-EGFP probe does not share the same responses or behaviour demonstrated by the HSPDE4A4-EGFP probe. Since the 4A4 and 4A1 probes share much of the same genomic and therefore primary protein sequence, behavioural differences can be ascribed with some confidence to those regions

of the two enzymes that differ. Specifically, these are from amino acid 1 to 22 of probe HSPDE4A1-EGFP and from amino acid 1 to 261 of probe HSPDE4A4-EGFP. The remaining primary sequence of these proteins is identical, as coded for in the plasmids described in Example 1 above.

5

Example 16: Quantitative assessment of the effects of Rolipram, RS25344, Ro 20-1724, Trequinsin and RP73401 on the cellular distribution of HSPDE4A1-EGFP probe in CHO cells.

This example shows how different PDE4 inhibitors, and one PDE3 inhibitor with some
10 PDE4 inhibitory activity, either affect the distribution of the 4A1 probe in a dose dependent fashion or have no significant effect on the distribution, that this distribution and any change thereof is readily measurable by automated imaging.

Fig. 35 shows dose response curves for spot dispersal in response to three different PDE4 inhibitors for a stable and clonal CHO cell line transfected with HSPDE4A1-EGFP
15 probe. The three inhibitors are Rolipram, RS25344 (▼) and Ro 20-1724 (O). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5 hours. The cells are then fixed with 4% formalin buffer
20 (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Estimated EC₅₀ values are 0.35 micromolar for Rolipram, 0.005 micromolar for RS25344 and 3.5 micromolar for Ro 20-1724. These values are very closely similar to the EC₅₀ values each
25 of these compounds has in causing redistribution of HSPDE4A4-EGFP probe in CHO cells, except in such an experiment spots are formed, and not dispersed as here (example 6).

Data in Fig. 36 has also been obtained by automated imaging from CHO cells stably transfected with HSPDE4A1-EGFP probe, but treated with various concentrations of
30 RP73401 (●), a specific and potent PDE4 inhibitor, and Trequinsin (▽), a PDE3 inhibitor with some action on PDE4. Again, the number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5 hours. The cells

are then fixed with 4% formalin buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25°C, then washed twice in PBS.

Automated images are collected and analysed for the number of spots per cell as described in Example 2. There is no significant spot dispersal over the concentration

5 ranges tested for either compound, in agreement with the lack of agonist activity of these compounds upon CHO cells stably transfected with HSPDE4A4-EGFP probe.

Example 17: Quantitative assessment of the effects of RP73401 plus Rolipram on the cellular distribution of HSPDE4A1-EGFP probe in CHO

10 cells.

This example shows that RP73401 can overcome the effect of Rolipram, and prevent the Rolipram-induced disappearance of spots, in a dose dependent fashion. This example describes how antagonists to the Rolipram effect on 4A1 may be found.

Fig. 37 shows a competitive dose response curve for Rolipram-induced spot dispersal in a
15 stable and clonal CHO cell line transfected with. The cells are challenged with a fixed concentration of 3 μ M Rolipram and then varying concentrations of the specific PDE4 inhibitor RP73401 (Piclamilast). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12
20 medium plus 10% FBS plus 3 μ M Rolipram for 20 hours. This treatment removes most spots from all cells. Various concentrations of RP73401 are then added and incubation continued for a further 6 hours. When there is sufficient RP73401 compound present to compete against Rolipram, spots re-form within the cells. The process of spot re-appearance in CHO cells expressing the HSPDE4A1-EGFP probe can be measured as
25 little as 60 minutes after addition of the test compound (RP73401 in this example), or as much as 24 hours after addition, if the test compound is sufficiently stable. Alternatively, both Rolipram and the test compound can be added simultaneously to these cells, and incubation continued for a period in the range 1 hour to 24 hours, after which the spot count per cell will again be indicative of possible antagonistic action.

30 After the test period, cells are fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25°C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. In all cases, if increasing concentrations of a test compound give increasing spot count per cell, then that compound is by definition

having an antagonistic effect on the action of Rolipram on 4A1. The data in Fig. 37 are fitted to a 4-parameter Hill equation, giving an IC_{50} of 0.01 μ M for RP73401 against 3 μ M Rolipram.

Example 18: Quantitative assessment of the effects of SB207499 (Ariflo®) on the ability of Rolipram and RS25344 to produce spots in CHO cells transfected either with the HSPDE4A4-EGFP, HSPDE4A4- Δ LR2-EGFP or HSPDE4A4-H506N-EGFP probes.

This example shows that Ariflo® (SB207499), a specific PDE4 inhibitor with excellent therapeutic properties and minimal side effect profile is able to prevent or reverse the usual spot forming activity of Rolipram or RS25344 in cells transfected with HSPDE4A4-EGFP, HSPDE4A4- Δ LR2-EGFP or HSPDE4A4-H506N-EGFP probes in a dose dependent way. This example demonstrates that the spot assay can be run in a competitive way to identify compounds that are specific PDE4 inhibitors and which interfere with the ability of Rolipram-like compounds to form spots, and that the spot assay can be used to quantify the competitive strength of such compounds.

Ariflo® does not by itself produce spots in any CHO cells transfected with either HSPDE4A4-EGFP, HSPDE4A4- Δ LR2-EGFP or HSPDE4A4-H506N-EGFP probes over the concentration range 30 to 0.01 μ M.

Fig. 38 shows competitive dose-response curves for Rolipram- and RS25344-induced spot formation in a stable and clonal CHO cell line transfected with the HSPDE4A4-EGFP probe. The cells are challenged with fixed concentrations of either Rolipram or RS25344 and various concentrations of Ariflo®. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of RP73401 plus 2 μ M Rolipram for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2.

Approximately 3.5 μ M SB207499 is sufficient to inhibit 50% of the spot formation response of these cells to 5 μ M Rolipram, indicating approximately equal competitive affinity or strengths of the two compounds for the catalytic cleft of the enzyme. A higher concentration of Ariflo®, approximately 20 μ M, is needed to produce the same effect against only 0.5 μ M of RS25344, confirming the greater affinity of RS25344 for position in the catalytic cleft.

Fig. 39 shows that the affinity of RS25344, as shown by the amount of Ariflo® needed to halve the spot formation response, is the same for the three different variants of the HSPDE4A4 probes used. This results also indicates that H506 is not involved in binding either RS25344 or Ariflo® to the catalytic cleft, nor is the LR2 region important in this regard.

Example 19: Redistribution of probe HSPDE4A4catD-EGFP caused by Rolipram and RS25344 treatment

This example illustrates how Rolipram and RS25344 affect the physical properties and behaviour of the HSPDE4A4catD-EGFP probe as expressed in stably transfected CHO cells.

Transiently transfected or stably transfected (non-clonal) cells are allowed to adhere to chambered coverglasses in HAM's F12 medium containing 10% FBS. Once adhered, either Rolipram (4-(3-cyclopentyloxy-4-methoxyphenyl)-2-pyrrolidone; Calbiochem) or RS25344 is added to the medium, and cells incubated further at 37 °C with 5% CO₂ + air.

At certain times after addition of Rolipram cells are checked on a fluorescence microscope for changes in cellular distribution of GFP fluorescence.

These experiments show that the general cytoplasmic distribution of cellular fluorescence gradually changes to one consisting of bright concentrations of fluorescence located at several distinct spots within the cytoplasm, some fluorescence remaining more evenly distributed within the rest of the (non nuclear) cytoplasm. A common pattern appears to be the presence of only 2 major accumulations of fluorescence diametrically separated across the nucleus of the cell (Fig. 40). Spots are stable in cells as long as Rolipram or RS25344 continues to be present.

Spots begin to be visible about 6 hours after addition of 1 µM RS25344. The effect is qualitatively similar at concentrations spanning the range 10 µM to 0.1 µM RS25344, 100 µM to 10 µM Rolipram. Pre-treatment of cells with 5 µg/ml cycloheximide prevents formation of spots induced by Rolipram and RS25344, indicating that protein synthesis is a necessary part of spot formation. Once spots have formed, removal of either compound results in their rapid dissolution, within 60 minutes at 37 °C. However, replacement of either Rolipram or RS25344 causes the bright spots to reform, also within 60 minutes. This is more rapid than is seen for *de novo* production of spots by Rolipram or RS25344 in these cells.

These experiments indicate that the HSPDE4A4catD-EGFP probe responds to Rolipram or RS25344 in qualitatively the same way as the HSPDE4A4-EGFP probe, and shows

that a cassette substitution of PDE4 catalytic regions into the PDE4A4 enzyme is a feasible method to search for isoform-specific catalytic inhibitors of PDE4. By extension from the methods described for the uses of HSPDE4A4-EGFP and HSPDE4A1-EGFP probes, the cassette substitution of other PDE4 isoform catalytic regions into these

- 5 PDE4A probes will allow discovery of isoform-specific catalytic inhibitors which belong either to the Rolipram group of compounds, or to the group of PDE4 inhibitors with low potential for causing emesis in humans (members of this latter group being Ariflo® and RP73401, for example).

- Fig. 40 shows the response to 1 μ M RS25344 of a population of CHO cells stably
10 transfected with probe HSPDE4A4cat4D-EGFP. Cells have been treated with RS25344 for 32 hours. Many cells in this stable population respond by forming pairs of bright spots in their cytoplasm. Fig. 41 shows CHO cells transiently transfected with HSPDE4A4cat4D-EGFP and treated with 10 μ M Rolipram for 26 hours. A fraction of cells in the heterogenous population respond by forming bright spots of fluorescence in their
15 cytoplasm.

Example 20: Quantitative assessment of the effects of RP73401 on the reappearance of stress-induced spots in CHO cells stably transfected with HSPDE4A4- E222G probe.

- This example describes the behaviour of the HSPDE4A4-E222G probe in a CHO cell
20 clone treated to produce stress-induced spots (see Example 13), but in the presence of various concentrations of RP73401 compound. The example demonstrates (1) that the E222G version of the PDE4A4 probe behaves in the same way as does the EGFP version, namely that PDE4A4 spots reappear under stress conditions, and (2) that the number of spots per cell that reappear under stress conditions is inhibited in a dose-
25 dependent way by RP73401 compound.

- CHO cells stably transfected with the HSPDE4A4-E222G probe are grown in HAM's F12 medium with 10% FBS, and with 3 micromolar Rolipram for 20 hours. Bright spots, usually paired, are present in more than 95% of all cells. Rolipram is then washed from the cells, and fresh HAM's F12 (with no additions) is added. Cells are returned to conditions of 37
30 °C + 5% CO₂ in humidified air (i.e. standard incubator conditions). After 4 hours, all GFP-bright spots disappear from cells.

Cells are then treated with various concentrations of RP73401 in HAM's F12 and left at ambient conditions (normal air, 22 to 25 °C) for a period of 3 hours (stress treatment). During this time the medium evaporates by about 15%, and the pH of the medium shifts

from pH6.5 to pH8.1 as the CO₂ in the medium equilibrates with ambient conditions. After 3 hours, spots reappear in the cytoplasm. The cells are then fixed with 4% formalin buffer (pH7) for 15 minutes, washed with PBS and stained with 10 µM Hoechst 33258 in PBS for 15 minutes at 25 °C, then washed twice in PBS. Automated images are collected and

5 analysed for the number of spots per cell as described in Example 2.

Spots do not reappear in these CHO cells under ambient conditions unless the cells are pre-treated with Rolipram. Spot reappearance behaviour is indistinguishable from that of cells stably transfected with HSPDE4A4-EGFP probe (Example 13).

Fig. 42 shows a dose response curves for spot reappearance under stress treatment.

10 Estimated IC₅₀ value is 0.3 nanomolar for RP73401. This value is equal to the IC₅₀ value determined for inhibition of PDE4 enzyme by this compound (Saldou *et al.* 1998). This result indicates that in the absence of Rolipram, the action of RP73401 opposes spot formation with a kinetic determined by simple reversible binding of the compound to the catalytic site.

15 **Example 21: Quantitative assessment of the effects of Rolipram on the cellular distribution of HSPDE4A1- E222G probe in CHO cells.**

This example shows how Rolipram affects the distribution of the PDE4A1-E222G probe in a dose dependent fashion, as measured by automated imaging, and that the response of this probe is indistinguishable from that of the HSPDE4A1- EGFP probe.

20 Fig. 43 shows a dose response curves for spot dispersal in a clonal line of CHO cells stably transfected with the HSPDE4A1- EGFP probe treated with Rolipram. The number of spots per cell for each concentration of Rolipram is the mean of 3 measurements ± sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of

25 Rolipram for 25 hours. The cells are then fixed with 4% formalin buffer (pH7) for 15 minutes, washed with PBS and stained with 10 µM Hoechst 33258 in PBS for 15 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Estimated EC₅₀ value is 0.1 micromolar for Rolipram. This value is very closely similar to the EC₅₀ value determined

30 for the HSPDE4A1-EGFP probe in CHO cells (0.35 micromolar, Example 16).

Figures

Fig. 1

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A1-EGFP growing in HAM's F12 medium with 10% FBS. The transfected cells are a clonal population derived from a single parent cell. GFP fluorescence is almost entirely restricted to bright granule-like spots within the perinuclear cytoplasm of each cell. The probe is not visible in the nuclei of these cells.

Fig. 2

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS. The transfected cells are a mixed, non-clonal population. GFP fluorescence is more or less evenly distributed throughout the non-nuclear cytoplasm, darker regions within this area are probably mitochondria from which the probe is apparently excluded.

Fig. 3

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-ΔLR2-EGFP growing in HAM's F12 medium with 10% FBS. The transfected cells are a clonal population derived from a single parent cell. GFP fluorescence is more or less evenly distributed throughout the non-nuclear cytoplasm, darker regions within this area are probably mitochondria from which the probe is apparently excluded.

Fig. 4

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-H506N-EGFP growing in HAM's F12 medium with 10% FBS. The transfected cells are a clonal population derived from a single parent cell. GFP fluorescence is more or less evenly distributed throughout the non-nuclear cytoplasm, darker regions within this area are probably mitochondria from which the probe is apparently excluded.

Fig. 5

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram. The transfected cells are a mixed, non-clonal population, and have been
5 treated with Rolipram for 42 hours. GFP fluorescence concentrates in bright spots in approximately 70% of the cell population.

As a scale to guide, nuclei sizes are generally in the range of 8 to 15 μm (mean of 11 μm s.d. 2.5 μm (n=15)).

Fig. 6

- 10 Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium without FBS but with 2 micromolar Rolipram. The transfected cells have been derived from a single cell isolated from a non-clonal population. The cells have been treated with Rolipram for 6.7 hours. GFP fluorescence concentrates in bright spots in more than 95% of the cells.
- 15 As a scale to guide, nuclei sizes are generally in the range of 8 to 15 μm (mean of 11 μm s.d. 2.5 μm (n=15)).

Fig. 7

- Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4- Δ LR2-EGFP growing in HAM's F12 medium with 10% FBS plus 3 micromolar
20 Rolipram. The transfected cells are a clonal population, and have been treated with Rolipram for 23.5 hours. In approximately 90% of the cells GFP fluorescence concentrates in bright spots, which are indistinguishable from those in seen in Rolipram-treated cells transfected with the "wild-type" probe HSPDE4A4-EGFP.

Fig. 8

- Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-H506N-EGFP growing in HAM's F12 medium with 10% FBS plus 100 micromolar Rolipram. The transfected cells are a clonal population, and have been treated with Rolipram for 23.5 hours. In only approximately 15% of the cells GFP
30 fluorescence concentrates in bright spots, which are indistinguishable from those in seen in Rolipram-treated cells transfected with the "wild-type" probe HSPDE4A4-EGFP.

Fig. 9

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS and 10 micromolar denbufylline (BRL30892). The transfected cells have been derived from a single cell isolated from a non-clonal population. The cells have been treated with denbufylline for 24.5 hours. GFP fluorescence concentrates in bright spots in approximately 40% of the cells.

Fig. 10a and 10b

Confocal fluorescence images showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS and two concentrations of RS 25344 for 24.5 hours : cells in 10a are treated with 0.03 micromolar RS 25344, cells in 10b with 1 micromolar. The transfected cells have been derived from a single cell isolated from a non-clonal population. GFP fluorescence concentrates in bright spots in approximately 40% of the cells in Fig 10a. In Fig 10b the accumulations of GFP fluorescence are considerably more massive, and present in more than 95% of cells.

Fig. 11a, b

Confocal fluorescence images showing CHO cells stably transfected with probe HSPDE4A4-EGFP. The transfected cells are a clonal population. In Fig. 11a the cells are grown in HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram for 6.7 hours. The cells in Fig. 11b have been treated with a combination of Rolipram plus 0.001 micromolar of the specific PDE4 inhibitor RP 73401 for 7.5 hours. RP73401 inhibits the Rolipram-induced production of spots in these CHO cells; GFP fluorescence concentrates in bright spots in less than 5% of the cell population.

Fig. 12

Confocal fluorescence images showing CHO cells stably transfected with probe HSPDE4A4-EGFP. The transfected cells are a clonal population. The cells are grown in HAM's F12 medium with 10% FBS with a combination of 2 micromolar Rolipram plus 0.003 micromolar of the specific PDE4 inhibitor RP 73401 for 7.5 hours. RP73401 inhibits the Rolipram-induced production of spots in these CHO cells; there are no spots formed in any of the cells.

Fig. 13

Wide-field fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS plus 10 micromolar of the specific PDE4 inhibitor Ro-20-1724. The transfected cells are a clonal population, and are been treated with Ro-20-1724 for 4.5 hours. GFP fluorescence concentrates in bright spots in approximately 80% of the cell population.

Fig. 14

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS plus 500 micromolar of the general PDE inhibitor IBMX. The transfected cells are a mixed, non-clonal population, and are treated with IBMX for 14 hours. GFP fluorescence forms small bright spots in about 10% of cells. In the remaining cells, the distribution is uniformly cytoplasmic, indistinguishable from untreated cells (Fig. 2). Those cells that contain spots are dissimilar to Rolipram treated cells (Figs. 5 and 6) in that they each contain more than two major bright spots.

Fig. 15

Dose response curves for spot formation in response to three different PDE4 inhibitors for a stable and clonal CHO cell line transfected with HSPDE4A4-EGFP probe. The three inhibitors are Rolipram (▼), RS25344 (■) and Ro 20-1724 (●). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Data sets are fitted to a 4-parameter Hill equation giving the following EC₅₀ values of 0.34 micromolar for Rolipram, 0.017 micromolar for RS25344 and 3.77 micromolar for Ro 20-1724.

Fig. 16

Dose response curves for spot formation in response to Rolipram in three stable and clonal cell lines of CHO cells transfected with HSPDE4A4-EGFP (), HSPDE4A4-ΔLR2-EGFP (∇) and HSPDE4A4-H506N-EGFP (▼). The number of spots per cell for each concentration is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of Rolipram for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Data sets are fitted to a 4-parameter Hill equation giving the following EC₅₀ values of 0.34 micromolar Rolipram for the HSPDE4A4-EGFP probe and 0.41 micromolar Rolipram for the HSPDE4A4-ΔLR2-EGFP probe. An EC₅₀ value can not be determined for the HSPDE4A4-H506N-EGFP probe, since the mutation makes it almost unresponsive to Rolipram.

Fig. 17

Dose response curves for spot formation in response to three different PDE4 inhibitors for a stable and clonal CHO cell line transfected with the HSPDE4A4-H506N-EGFP probe. The three inhibitors are Rolipram (▼), RS25344 (■) and Ro 20-1724 (●). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The data set for RS25344 is fitted to a 4-parameter Hill equation giving an EC₅₀ value of 0.125 micromolar. These clonal cells containing the H506N mutant of HSPDE4A4-EGFP are almost unresponsive to the other two inhibitors over the concentrations tested.

Fig. 18

A competitive dose response curve for Rolipram-induced spot formation in a stable and clonal CHO cell line transfected with HSPDE4A4-EGFP probe. The cells are challenged with a fixed concentration of 2 micromolar Rolipram and varying concentrations of the specific PDE4 inhibitor RP73401 (Piclamilast). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of RP73401 plus 2 micromolar Rolipram for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Approximately 0.003 micromolar RP73401 is sufficient to inhibit 50% of the spot formation response of these cells that normally results from treatment with 2 micromolar Rolipram.

15

Fig. 19a to d

The figures show four fluorescence images from a time lapse sequence for non-clonal CHO cells transfected with the HSPDE4A4-EGFP probe. Cells are pre-treated with 2 micromolar Rolipram for 24 hours, then given 50 micromolar forskolin plus 1 millimolar IBMX (all in HAM's F12 medium plus 10% FBS). Fig. 15a is taken immediately before addition of IBMX plus forskolin, Fig. 15b, c and d at 6, 9 and 24 minutes after that addition. Two regions of interest, marked A and B in Fig. 15a, are used to generate the time profiles shown in Fig 16a and Fig 16b respectively (according to the method in Example 2).

25

Fig. 20a

Time profile derived from a region of interest (ROI) delineating an area of cytoplasm, marked as A in Fig. 15a. IBMX and forskolin are added 2 minutes prior to the start of imaging. The curve is derived by averaging pixel values within the ROI for each image in the sequence. Images are taken at intervals of 30 seconds.

30

Fig. 20b

Time profile derived from a ROI delineating a single bright spot, marked as B in Fig. 15a. IBMX and forskolin are added 2 minutes prior to the start of imaging. The curve is derived by averaging pixel values within the ROI for each image in the sequence. Images are
5 taken at intervals of 30 seconds.

Fig. 21

This figure shows results from a FLIPR™ (Molecular Devices) 96-well plate reader. The plate contains clonal CHO cells transfected with the HSPDE4A4-EGFP probe that are
10 treated with 2 micromolar Rolipram for 24 hours, then washed to KRW buffer plus 2 micromolar Rolipram just prior to running the experiment. Time traces A and B represent averages over 8 wells each for responses to 500 micromolar IBMX plus 50 micromolar forskolin, where wells for curve B are pre-treated with 2 micromolar compound H-89 for 20 minutes, and those for curve A are not. Curves are normalised and corrected to a buffer +
15 DMSO control. The experiment is run at 37 °C, and addition of the test compounds occurs after the first minute. Readings after the addition are made at intervals of 1 minute. The difference in the levels of response indicates that the inhibitor of PKA has a significant effect on the dispersal of spots that is induced by IBMX plus forskolin, suggesting a role for PKA in this process.

20

Fig. 22

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram. The transfected cells are a mixed, non-clonal population, and are treated with
25 Rolipram for 42 hours, then further treated with fresh HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram plus 0.2% DMSO for 50 minutes. All treatments are under standard incubator conditions. GFP fluorescence remains concentrated in bright spots in approximately 70% of the cell population.

30 Fig. 23

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram. The transfected cells are a mixed, non-clonal population, and are treated with

Rolipram for 42 hours, then further treated with fresh HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram plus 2 micromolar ionomycin for 50 minutes. All treatments are under standard incubator conditions. GFP fluorescence remains concentrated in bright spots in approximately 20 to 40% of the cell population.

5

Fig. 24

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram. The transfected cells are a mixed, non-clonal population, and are treated with

10 Rolipram for 42 hours, then further treated with fresh HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram plus 200 nM PMA for 50 minutes. All treatments are under standard incubator conditions. GFP fluorescence is no longer concentrated in bright spots in any of the cell population.

15 **Fig. 25**

Confocal fluorescence image showing CHO cells stably transfected with probe HSPDE4A4-EGFP growing in HAM's F12 medium with 10% FBS plus 2 micromolar Rolipram. The transfected cells are a mixed, non-clonal population, and have been treated with Rolipram for 42 hours, then further treated with fresh HAM's F12 medium with

20 10% FBS plus 2 micromolar Rolipram plus 200 nM PMA plus 2 micromolar ionomycin for 50 minutes. All treatments are under standard incubator conditions. GFP fluorescence is no longer concentrated in bright spots in any of the cell population.

Fig. 26a and 26b

25 Confocal fluorescence images showing clonal CHO cells stably transfected with probe HSPDE4A4-EGFP in KRW buffer with no FBS plus 2 micromolar Rolipram. The cells are depleted of serum for more than 22 hours. Fig 26a shows the cells before treatment, Fig. 26b 18 minutes after addition of 50 micromolar forskolin plus 500 micromolar IBMX. This treatment is under ambient conditions on the microscope stage. After 18 minutes, most

30 large spots have dispersed within the cells.

Fig. 27a and 27b

Confocal fluorescence images showing clonal CHO cells stably transfected with probe HSPDE4A4-EGFP in KRW buffer with no FBS plus 2 micromolar Rolipram. The cells have been depleted of serum for more than 22 hours. Fig 27a shows the cells before treatment, Fig. 27b 38 minutes after addition of 200 nanomolar PMA plus 2 micromolar ionomycin. This treatment is under ambient conditions on the microscope stage. There is no significant dispersal of the large fluorescent spots under this protocol.

Fig. 28a, 28b, 28c

10 Confocal fluorescence images showing CHO cells stably transfected with probe HSPDE4A4-EGFP. The transfected cells have been derived from a single cell isolated from a non-clonal population. These cells are grown in HAM's F12 medium with 10% FBS, and with 2 micromolar Rolipram for 15.5 hours. Bright spots are present in more than 95% of all cells. Rolipram is then washed from the cells, and fresh HAM's F12 + 10%
15 FBS is added. After 150 minutes all GFP-bright spots disappear from cells. A volume of 1 molar NaCl is then added to the cells to increase the final concentration of salt in the medium by 100 mM. The cells in Fig. 28c are additionally treated with 5 micromolar SB203580, a specific inhibitor of p38 mitogen activated protein kinases (p38 MAPK). The cells in Fig. 28b are returned to conditions of 37 °C + 5% CO₂ in humidified air (i.e.
20 standard incubator conditions), while those in 28a and 28c are chilled to 4 °C. After 4 hours of these treatments cells were fixed with 4% formaldehyde pH 7.0 at room temperature for 1 hour, and washed with PBS buffer ready for imaging. Many small GFP-bright spots form in more than 90% of the chilled cells, but of those returned to incubator conditions (Fig 28b) less than 5% of cells contain spots. Chilled and SB 203580-treated
25 cells (Fig. 28c) contain significantly fewer, but larger, bright spots per cell than those in Fig. 28a.

Fig. 29a, 29b

Confocal fluorescence images showing CHO cells stably transfected with probe
30 HSPDE4A4-EGFP. The transfected cells have been derived from a single cell isolated from a non-clonal population. These cells are grown in HAM's F12 medium without FBS, but with 2 micromolar Rolipram for 12 hours. Bright spots are present in more than 95% of all cells. Rolipram is then washed from the cells, and fresh HAM's F12 (no FBS) added.

The cells in Fig. 29b are additionally treated with 400 micromolar thalidomide. Cells are returned to conditions of 37 °C + 5% CO₂ in humidified air (i.e. standard incubator conditions). After 60 minutes all GFP-bright spots disappear from cells. The coverglass chambers or 96-well plates containing the cells are then left at ambient room conditions for a further 4 hours to cool during which the growth medium evaporates by about 20%. During this time GFP-bright spots reappear in about 50% of the cells which are not treated with thalidomide (Fig. 29a). Spots reappear in less than 5% of cells under these conditions in the presence of 400 micromolar thalidomide (Fig. 29b).

10 **Fig. 30**

Dose response curves for spot reappearance in response to different concentrations of added NaCl for a stable and clonal CHO cell line transfected with HSPDE4A4-EGFP probe. The number of spots per cell for each concentration of NaCl is the mean of 2 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. These cells are grown in HAM's F12 medium with 10% FBS, plus 2 micromolar Rolipram for 15.5 hours. Bright spots are present in more than 95% of all cells. Rolipram is then washed from the cells, and fresh HAM's F12 + 10% FBS is added. Cells are returned to conditions of 37 °C + 5% CO₂ in humidified air (i.e. standard incubator conditions). After 150 minutes all GFP-bright spots disappear from cells. Various amounts of 1 molar NaCl is then added to different populations of cells to increase the final concentration of salt in the medium by 0 mM, 5 mM, 50 mM or 100 mM. Another group of cells are treated similarly but with the addition of 5 micromolar SB203580, an inhibitor of p38 MAPK. All treatments are then chilled to 4 °C, in normal air, for 4 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The number of spots per cell increases in a dose-dependent fashion with increasing salt concentration. SB203580 decreases the number of spots per cell. Examination of the images from the SB203580 treatment (e.g. Fig. 28c) suggests that decreased spot number is accompanied by increased spot size.

Fig. 31

A dose response curve for the inhibition of spot reappearance under ambient conditions by thalidomide for a stable and clonal CHO cell line transfected with HSPDE4A4-EGFP

probe. The number of spots per cell for each concentration of thalidomide is the mean of 2 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. These cells are grown in HAM's F12 medium with 10% FBS, plus 2 micromolar Rolipram for 12 hours. Bright spots are present in more than 95% of all cells.

5 Rolipram is then washed from the cells, and fresh HAM's F12 + 10% FBS is added together with different concentrations of thalidomide. Cells are returned to conditions of 37 °C + 5% CO₂ in humidified air (i.e. standard incubator conditions). After 60 minutes all GFP-bright spots disappear from cells. The coverglass chambers (or 96-well plates) containing the cells are then left at ambient room conditions for a further 4 hours to cool

10 during which the growth medium evaporates by about 20%. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The data set is fitted to a 4-parameter Hill equation giving an IC₅₀ value of 33

15 micromolar thalidomide under these conditions.

Fig. 32a and 32b

Confocal fluorescence images showing clonal CHO cells stably transfected with probe HSPDE4A4-EGFP treated with 2 micromolar Rolipram. The cells are depleted of serum

20 then washed to KRW buffer with no added FBS for over 3 hours. Fig 32a shows the cells before treatment, Fig. 32b 3 minutes after addition of 2 micromolar ionomycin. Very many smaller spots appear in the cytoplasm, without changing the size or number of the larger Rolipram-induced spots.

25 Fig. 33a and 33b

These images continue from the treatment shown in Fig 32a and 32b. The cells are further treated with 50 micromolar forskolin plus 500 micromolar IBMX 7 minutes prior to the image shown in Fig. 33a. There is significant dispersal of the smaller spots by this time. By 24 minutes after forskolin and IBMX treatment (Fig. 33b), the larger spots are

30 beginning to disperse as normal.

Fig. 34a and 34b

Confocal fluorescence images showing CHO cells stably transfected with probe HSPDE4A1-EGFP. Images are recorded at the same microscope settings for direct comparison of intensities. The transfected cells are a clonal population derived from a single parent cell. In Fig. 34a the cells are growing in only HAM's F12 medium with 10% FBS; the GFP fluorescence is restricted to bright granule-like spots within the perinuclear cytoplasm of each cell. In Fig. 34b similar cells to those seen in 34a have been treated with 2 micromolar Rolipram for 2 hours. The majority of GFP-bright spots disappear in all cells under Rolipram treatment, and the cytoplasm becomes generally brighter. Larger spots do not disperse in some cells. When Rolipram is washed away, the spots reform within 1.75 hours.

Fig. 35

Dose response curves for spot dispersal in response to three different PDE4 inhibitors for a stable and clonal CHO cell line transfected with HSPDE4A1-EGFP probe. The three inhibitors are Rolipram (●), RS25344 (▼) and Ro 20-1724 (○). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5 hours. The cells are then fixed with 4% formalin buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Estimated EC50 values are 0.35 micromolar for Rolipram, 0.005 micromolar for RS25344 and 3.5 micromolar for Ro 20-1724.

Fig. 36

Dose response curves for spot dispersal in response to two different PDE inhibitors for a stable and clonal CHO cell line transfected with HSPDE4A1-EGFP probe. The two inhibitors are RP73401 (●), aspecific and potent PDE4 inhibitor, and Trequinsin (▼), a PDE3 inhibitor with some action on PDE4. The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of inhibitors for 23.5

hours. The cells are then fixed with 4% formalin buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. There is no significant spot dispersal over the concentration ranges tested for either compound.

Fig. 37

A competitive dose response curve for Rolipram-induced spot dispersal in a stable and clonal CHO cell line transfected with HSPDE4A1-EGFP probe. The cells are challenged with a fixed concentration of 3 μ M Rolipram and then varying concentrations of the specific PDE4 inhibitor RP73401 (Piclamilast). The number of spots per cell for each concentration of the different inhibitors is the mean of 4 measurements \pm sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus 3 μ M Rolipram for 20 hours. Various concentrations of RP73401 are then added and incubation continued for a further 6 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The data are fitted to a 4-parameter Hill equation, giving an IC_{50} of 0.01 μ M for RP73401 against 3 μ M Rolipram.

Fig. 38

Competitive dose response curves for Rolipram- and RS25344-induced spot formation in a stable and clonal CHO cell line transfected with HSPDE4A4-EGFP probe. The cells are challenged with a fixed concentration of either 5 micromolar Rolipram (\diamond) or 0.5 micromolar RS25344 (\blacklozenge) and varying concentrations of the specific PDE4 inhibitor SB207499 (Ariflo[®]). Cells are grown in HAM's F12 medium plus 10% FBS plus the inhibitors for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μ M Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. The data set for Rolipram is fitted to a 3-parameter Hill equation giving an IC_{50} value of 3.37 micromolar for SB207499 in this competition.

Fig. 39

Competitive dose response curves for RS25344-induced spot formation in three stable and clonal CHO cell lines separately transfected with HSPDE4A4-EGFP (•), HSPDE4A4-H506N-EGFP (▽) or HSPDE4A4-ΔLR2-EGFP (□) probes. The cells are challenged with a fixed concentration of 0.5 micromolar RS25344 and varying concentrations of the specific PDE4 inhibitor SB207499 (Ariflo®). Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of SB207499 plus 2 micromolar Rolipram for 23.5 hours. The cells are then fixed with 4% formaldehyde buffer (pH7) for 1 hour, washed with PBS and stained with 1 μM Hoechst 33258 in PBS for 10 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Approximately 20 micromolar SB207499 is needed to reduce by 50% the spot formation response to 0.5 micromolar RS25344.

Fig. 40

Confocal fluorescence image shows the response to 1 μM RS25344 of a population of CHO cells stably transfected with probe HSPDE4A4cat4D-EGFP. Cells have been treated with RS25344 for 32 hours. Many cells in this stable population respond by forming pairs of bright spots in their cytoplasm.

Fig. 41

Confocal fluorescence image shows CHO cells transiently transfected with HSPDE4A4cat4D-EGFP and treated with 10 μM Rolipram for 26 hours. A fraction of cells in the heterogenous population respond by forming bright spots of fluorescence in their cytoplasm.

Fig. 42

Shows a dose response curves for spot reappearance under stress treatment in the presence of various concentrations of RP73401 in a clonal line of CHO cells stably transfected with the HSPDE4A4-E222G probe. The number of stress-induced spots per cell for each concentration of RP73401 is the mean of 3 measurements ± sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus 3 μM Rolipram for 20 hours.

Rolipram is then washed from the cells, and fresh HAM's F12 (with no additions) is added. Cells are returned to conditions of 37 °C + 5% CO₂ in humidified air (i.e. standard incubator conditions). After 4 hours, all GFP-bright spots disappear from cells. Cells are then treated with various concentrations of RP73401 in HAM's F12 and left at ambient conditions (normal air, 22 to 25 °C) for a period of 3 hours (stress treatment). During this time the medium evaporates by about 15%, and the pH of the medium shifts from pH6.5 to pH8.1 as the CO₂ in the medium equilibrates with ambient conditions. After 3 hours, spots reappear in the cytoplasm.

The cells are then fixed with 4% formalin buffer (pH7) for 15 minutes, washed with PBS and stained with 10 µM Hoechst 33258 in PBS for 15 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Estimated IC₅₀ value is 0.3 nanomolar for RP73401. This value is equal to the IC₅₀ value determined for inhibition of PDE4 enzyme by this compound (Saldou *et al.* 1998).

15 **Fig. 43**

Shows a dose response curves for spot dispersal in a clonal line of CHO cells stably transfected with the HSPDE4A1- EGFP probe treated with Rolipram. The number of spots per cell for each concentration of Rolipram is the mean of 3 measurements ± sem, where each measurement is itself an average taken from not less than 100 cells. Cells are grown in HAM's F12 medium plus 10% FBS plus various concentrations of Rolipram for 25 hours. The cells are then fixed with 4% formalin buffer (pH7) for 15 minutes, washed with PBS and stained with 10 µM Hoechst 33258 in PBS for 15 minutes at 25 °C, then washed twice in PBS. Automated images are collected and analysed for the number of spots per cell as described in Example 2. Estimated EC₅₀ value is 0.1 micromolar for Rolipram.

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B. IDENTIFICATION OF DEPOSIT Further deposits are identified on an additional sheet <input checked="" type="checkbox"/>	
Name of depositary institution DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH	
Address of depositary institution (including postal code and country) Mascheroder Weg 1B D-38124 Braunschweig Germany	
Date of deposit 17 April 2000	Accession Number DSM 13449
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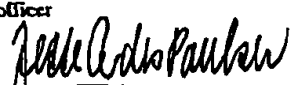
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Claims

1. A method to determine if a compound is a dislocator of PDE4, comprising the steps of:
 - testing if the compound removes PDE4-spots, where PDE4-spots may optionally be induced by a Rolipram-like reference compound and
- 5 - testing if the compound inhibits the catalytic activity of the PDE4;
the compound being a dislocator of PDE4, if the compound removes PDE4-spots and if the compound does not inhibit the catalytic activity of PDE4.
2. A method according to any of the preceding claims, wherein the Rolipram-like
10 reference compound is Rolipram.
3. A method according to any of the preceding claims, wherein the PDE4 is a PDE4A isoform.
- 15 4. A method according to any of the preceding claims, wherein the PDE4 is the PDE4A1 isoform.
5. A method according to any of the preceding claims, wherein the PDE4 is the PDE4A4
20 isoform.
6. A method according to any of the previous claims, comprising the steps of:
 - testing if the compound removes PDE4A1-spots, and
 - testing if the compound inhibits the catalytic activity of the PDE4A1;the compound being a dislocator of PDE4A1, if the compound removes PDE4A1-spots
25 and if the compound does not inhibit the catalytic activity of PDE4A1.
7. A method according to any of the previous claims, comprising the steps of:
 - testing if the compound removes PDE4A4-spots induced by a Rolipram-like reference compound and
- 30 - testing if the compound inhibits the catalytic activity of the PDE4A4;
the compound being a dislocator of PDE4A4, if the compound removes PDE4A4-spots and if the compound does not inhibit the catalytic activity of PDE4A4.
8. A PDE4 dislocator obtainable by the method according to any of the previous claims.

9. A pharmaceutical composition comprising a compound, the compound being a dislocator of PDE4, and the pharmaceutical composition having a market authorisation, the market authorisation being based on an application for market authorisation comprising data showing removal of PDE4-spots, optionally induced by a Rolipram-like reference compound, by the compound and lack of inhibition of the catalytic activity of PDE4 by the compound.
10. A pharmaceutical composition according to the previous claim, wherein the PDE4 is a PDE4A isoform.
11. A pharmaceutical composition according to any of the previous claims, wherein PDE4 is the PDE4A1 isoform and wherein the indication on the market authorisation is diseases in the central nervous system.
12. A pharmaceutical composition according to the previous claim, wherein the indication is depression.
13. A pharmaceutical composition according to any of the previous claims, wherein PDE4 is the PDE4A4 isoform and wherein the indication on the market authorisation is inflammatory diseases.
14. A pharmaceutical composition according to the previous claim, wherein the indication is selected from the group consisting of joint inflammation, Crohn's disease, inflammatory bowel disease, respiratory diseases, chronic obstructive pulmonary disease (COPD), including asthma, chronic bronchitis, pulmonary emphysema, endotoxic shock, toxic shock syndrome, systemic lupus erythematosus, psoriasis, bone resorption diseases, reperfusion injury, cancer and HIV infection.
15. A method to determine if a compound is a low emesis PDE4 inhibitor comprising the steps of:
- testing if the compound causes PDE4A4-spots induced by a Rolipram-like reference compound to dissolve,
 - testing if the compound induces re-appearance of PDE4A1-spots in cells exposed to a Rolipram-like reference compound, and

- testing if the compound inhibits the catalytic activity of PDE4;

the compound being a low emesis PDE4 inhibitor if the compound removes spots induced by the Rolipram-like reference compound and induces re-appearance of PDE4A1 spots in cells exposed to the Rolipram-like reference compound and if the compound inhibits the

5 catalytic activity of PDE4.

16. A pharmaceutical composition comprising a compound, the compound being a low emesis PDE4 inhibitor, and the pharmaceutical composition having a market authorisation, the market authorisation being based on an application for market

10 authorisation comprising data showing that the compound removes spots induced by the Rolipram-like reference compound, and that the compound induces re-appearance of PDE4A1 spots in cells exposed to the Rolipram-like reference compound, and that the compound inhibits the catalytic activity of PDE4.

15 17. A pharmaceutical composition according to any of the previous claims, wherein the indication on the market authorisation is inflammatory diseases.

18. A pharmaceutical composition according to the previous claim, wherein the indication is selected from the group consisting of joint inflammation, Crohn's disease, inflammatory

20 bowel disease, respiratory diseases, chronic obstructive pulmonary disease (COPD), including asthma, chronic bronchitis, pulmonary emphysema, endotoxic shock, toxic shock syndrome, systemic lupus erythematosus, psoriasis, bone resorption diseases, reperfusion injury, cancer and HIV.

25 19. A method to monitor changes in intracellular distribution of phosphodiesterases (PDEs) in living cells, the method comprising the steps of:

(a) recording the intracellular distribution of the PDE;

(b) adding a Rolipram-like reference compound to the cells in (a) or to similar cells;

(c) recording the intracellular distribution of the PDE in the cells in step (b);

30 (d) determining the effect on the intracellular distribution of the PDE of the Rolipram-like reference compound by comparing the intracellular distribution recorded in step (a) with the intracellular distribution recorded in step (c).

20. A method according to the previous claim, further comprising the following steps prior
35 to step (a):

- (O1) constructing a probe allowing the location of the PDE to be recorded;
- (O2) transfecting cells with the constructed probe of step (a1);

21. A method according to the previous claim, wherein the probe is constructed such that
5 the location of the PDE can be recorded continuously.

22. A method to identify a reagent, capable of interfering with the intracellular distribution of PDEs, comprising the method according to any of the preceding claims further comprising the following steps after step (b):

- 10 (b1) adding a reagent to the compound treated cells in step (b) or similar cells;
- (b2) recording the intracellular distribution of the PDE in the cells in step (b1);
- and the method comprising the following steps after step (d):
- (d1) determining the effect of the reagent by comparing the intracellular distribution recorded in step (b2) with the intracellular distribution recorded in step (a);
- 15 (d2) establish the pharmacology of the reagent by comparing the determined effect in step (d1) with the determined effect in step (d)
- a reversal of the effect determined in step (d) to the effect substantially identical to the effect determined in step (a) in step (d1) being indicative of an antagonistic effect of the reagent on the compound with affinity for the catalytic site of the PDE in regards to the
- 20 change in intracellular distribution.

23. A method according to any of the preceding claims, further comprising the step of:
(e) determining the effect of the reagent on the catalytic activity of the PDE in an assay capable of measuring the catalytic activity of PDEs.

25

24. A method according to any of the preceding claims, further comprising the following steps after step (b):
- (b1) adding a reagent to similar cells of those in step (a);
 - (b2) recording the intracellular distribution of the PDE in the cells in step (b1);
 - 30 and the method comprising the following steps after step (d):
 - (d1) determining the effect of the reagent by comparing the intracellular distribution recorded in step (b2) with the intracellular distribution recorded in step (a);
 - (d2) establish the pharmacology of the reagent by comparing the determined effect in step (d1) with the determined effect in step (d)

a copy of the effect determined in step (d) to the effect in step (a) in step (d1) being indicative of an agonistic effect of the reagent on the compound with affinity for the catalytic site of the PDE in regards to the change in intracellular distribution.

5 25. A method according to any the two previous claims, wherein the reagent does not bind to the docking site of the PDE4.

26. A method according to any of the three previous claims, wherein the reagent binds to the catalytic site of the PDE4.

10

27. A method according to any of the preceding claims, wherein the reagent inhibits the catalytic activity of the PDE.

28. A method according to any of the preceding claims, wherein the reagent is a peptide
15 or a polypeptide.

29. A method according to any of the preceding claims, wherein the reagent is a small molecule.

20 30. A method according to any of the preceding claims, wherein PDE is a PDE4.

31. A method according to any of the preceding claims, wherein the Rolipram-like reference compound is Rolipram.

25 32. A method according to any of the preceding claims, wherein the comparison between the effect of the reagent and the effect of the compound is based on a time series of measurements.

33. A method according to any of the preceding claims, wherein the comparison between
30 the effect of the reagent and the effect of the compound is based on an end-point measurement.

34. A reagent obtainable by the method according to any of the preceding claims.

35. Use of a reagent being able to mimic the effect of the compound with affinity for the catalytic site on intracellular distribution of the PDE for the preparation of a medicament.
36. Use of a reagent being able to reverse the effect of the compound with affinity for the
5 catalytic site on intracellular distribution of the PDE for the preparation of a medicament.
37. A method for treating asthma in an individual comprising administering to the individual an effective amount of a compound, or a pharmaceutically acceptable salt, ester, amide or prodrug thereof, the compound being capable of reversing the effect of a
10 compound with affinity for the catalytic site of a PDE on the intracellular distribution of the PDE and mimicking the effect of the compound with affinity for the catalytic site of the PDE on the catalytic activity of the PDE.
38. The nucleic acid HSPDE4A1-EGFP construct.
15
39. The nucleic acid HSPDE4A4-EGFP construct.
40. The nucleic acid HSPDE4A4-H506N-EGFP construct.
- 20 41. The nucleic acid HSPDE4A4- Δ LR2-EGFP construct.
42. The nucleic acid HSPDE4A4-EGFP construct.
43. The nucleic acid HSPDE4A4catD-EGFP construct.
25
44. The nucleic acid HSPDE4D3-EGFP construct.
45. The nucleic acid HSPDE4A1-E222G construct.
- 30 46. The nucleic acid HSPDE4A4-E222G construct.

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Fig. 1

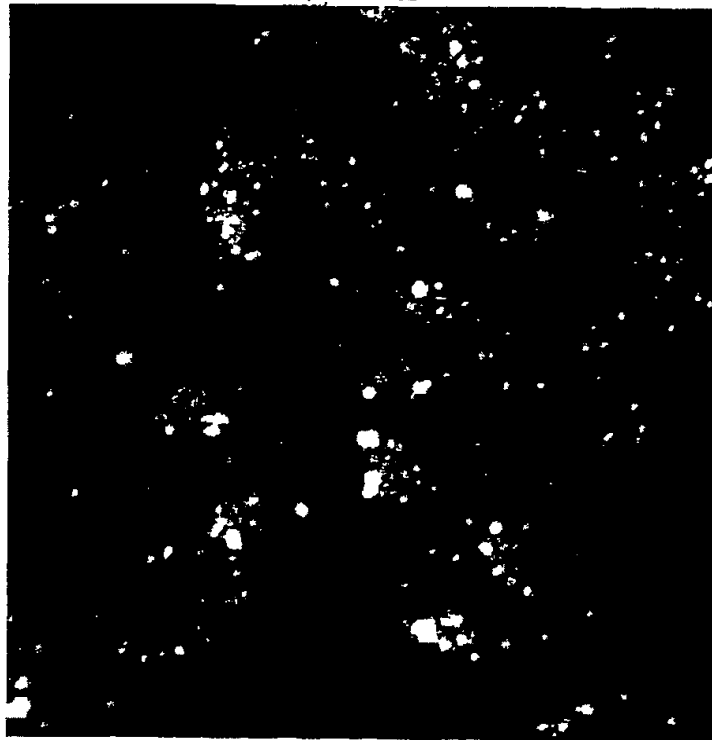


Fig. 2



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Fig. 3

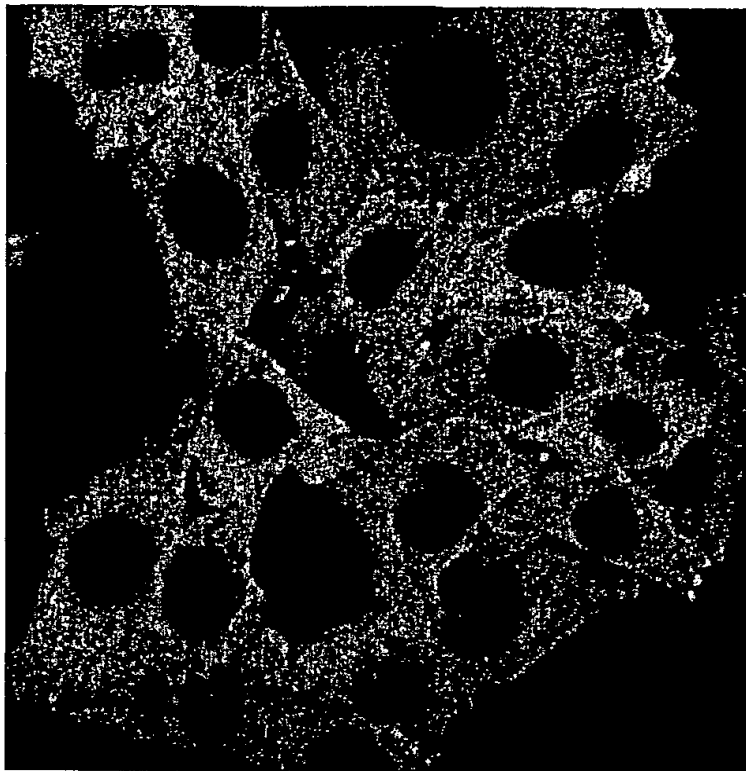
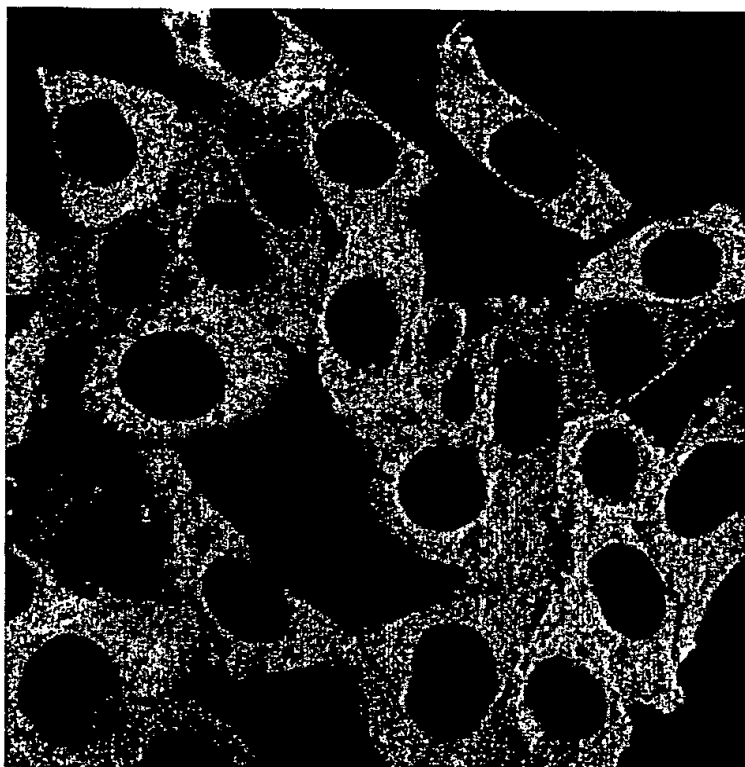


Fig. 4



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Fig. 5

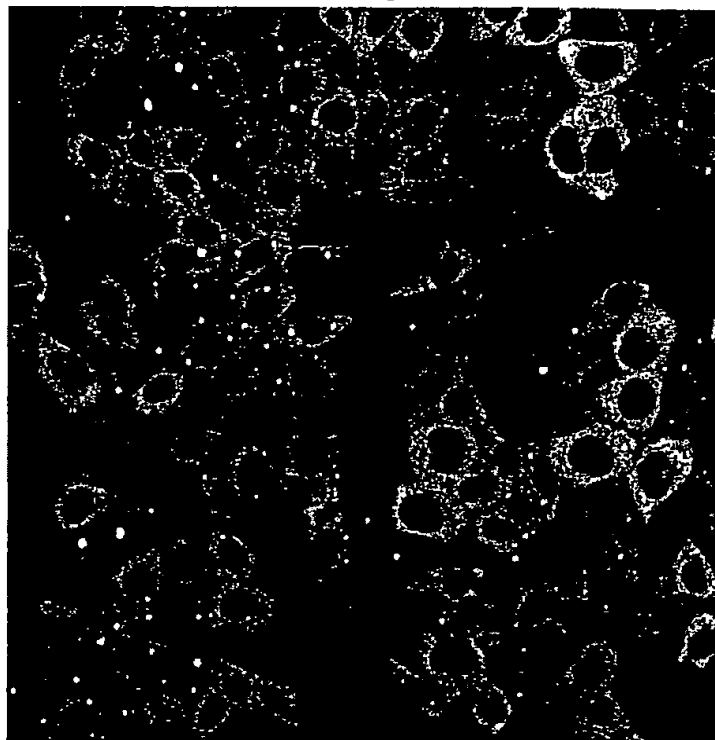
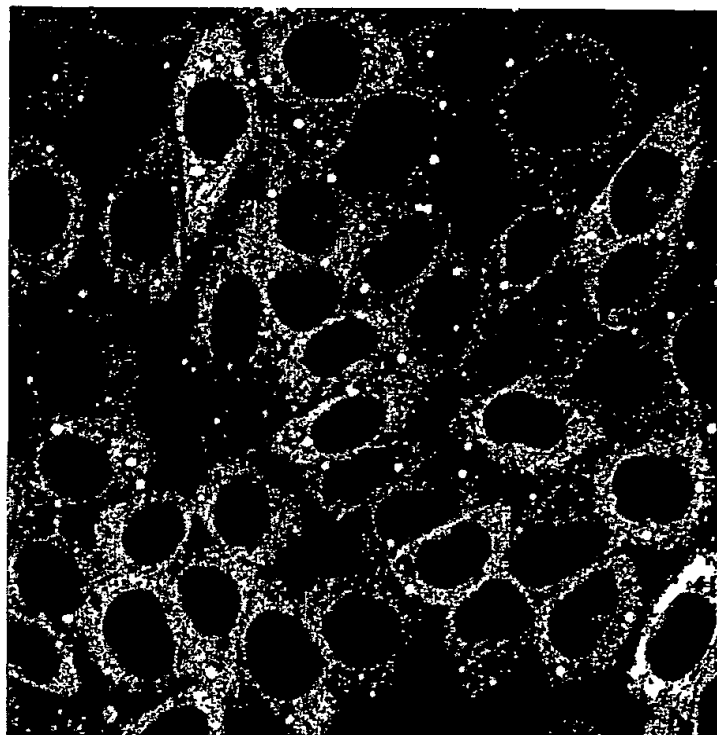


Fig. 6



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Fig. 7

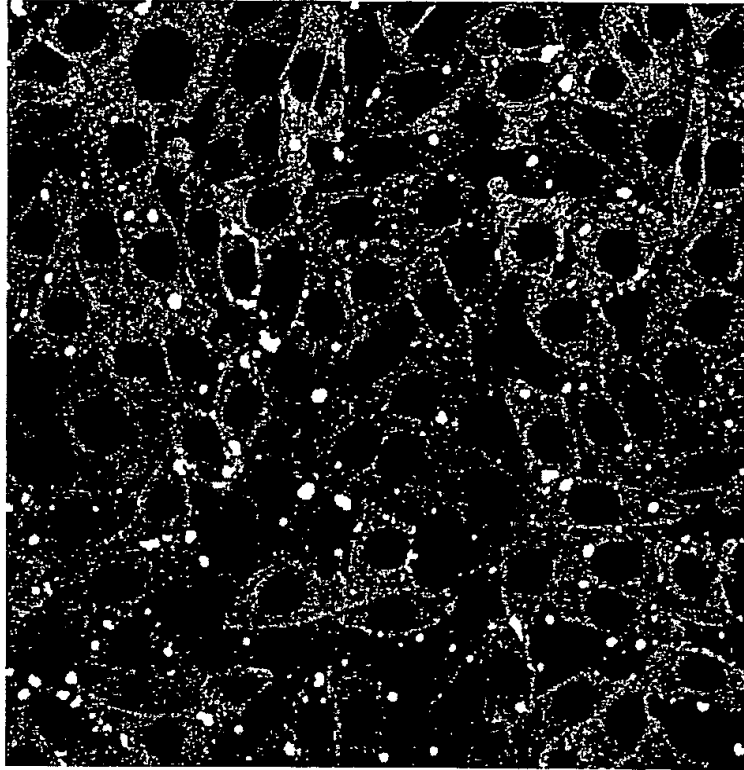
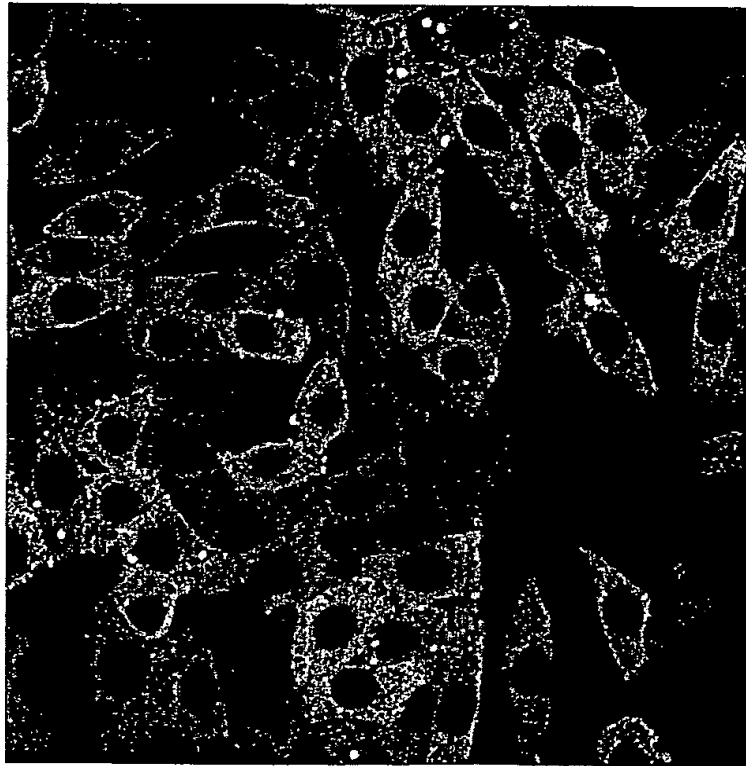


Fig. 8



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Fig. 9

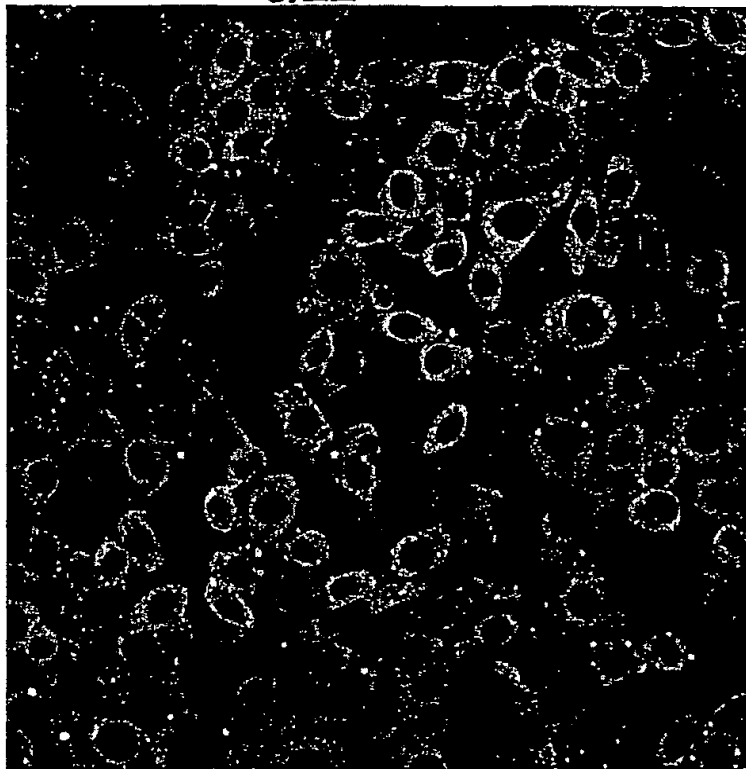
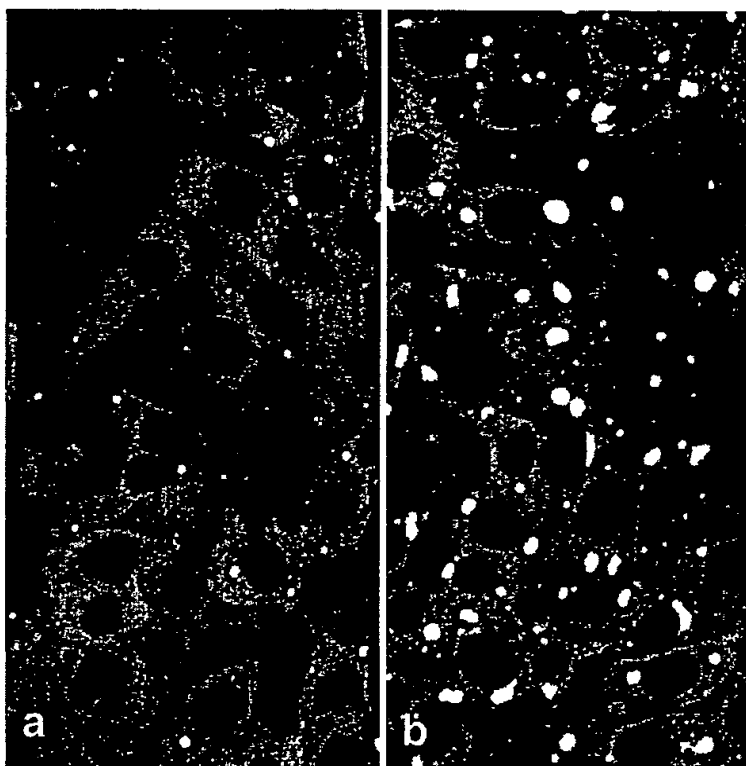


Fig. 10



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Fig. 11

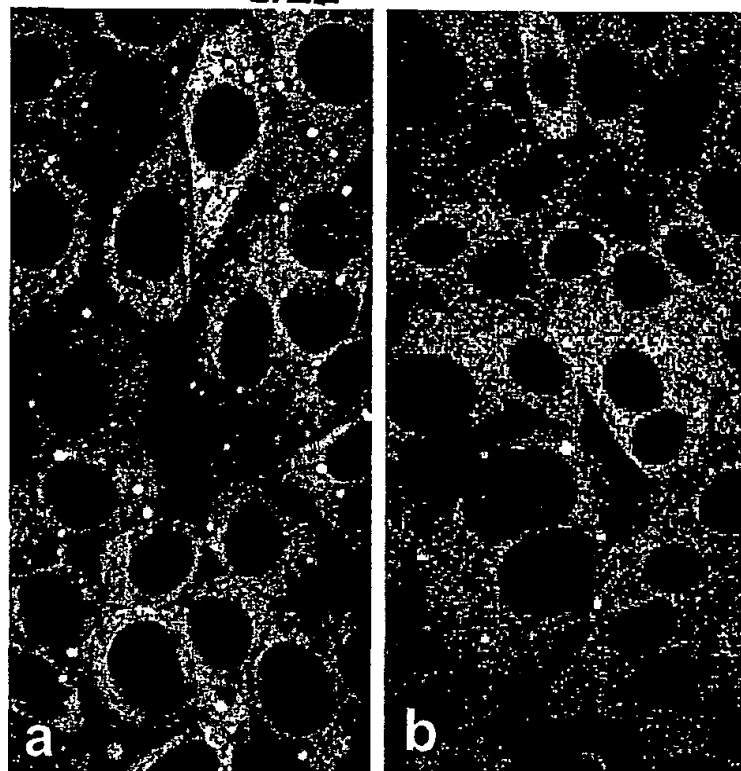


Fig. 12

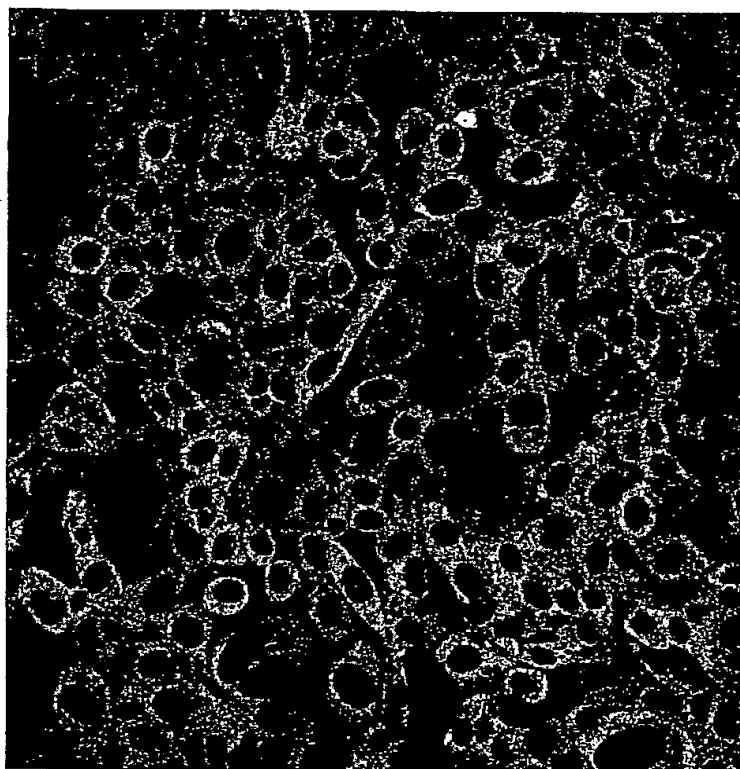
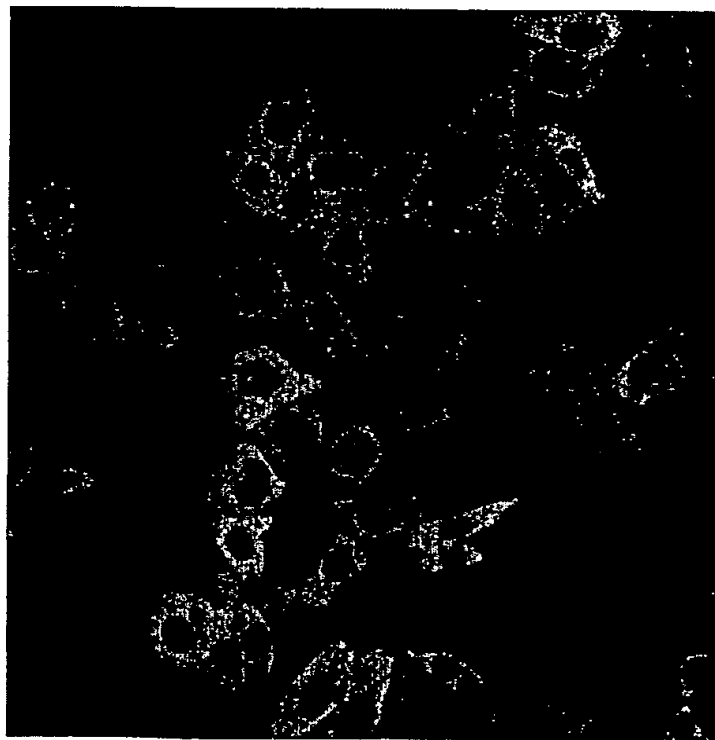


Fig. 13

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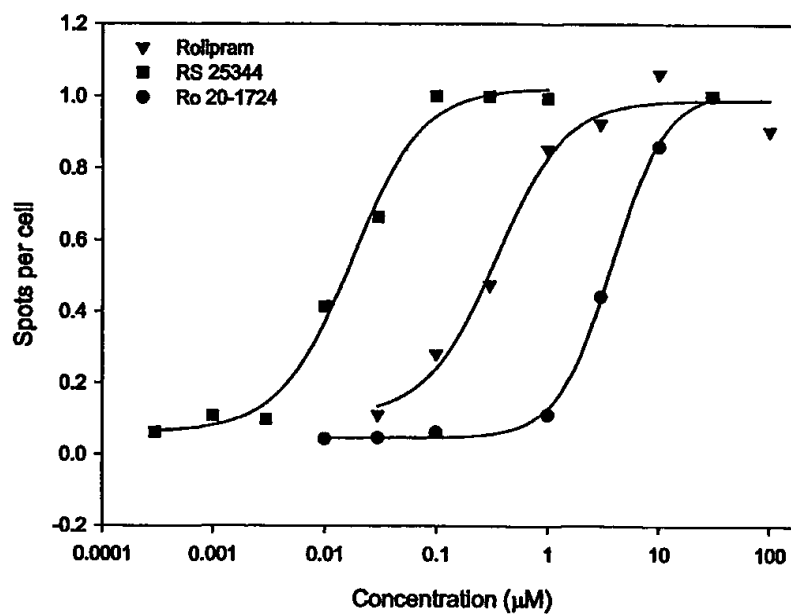
Fig. 14



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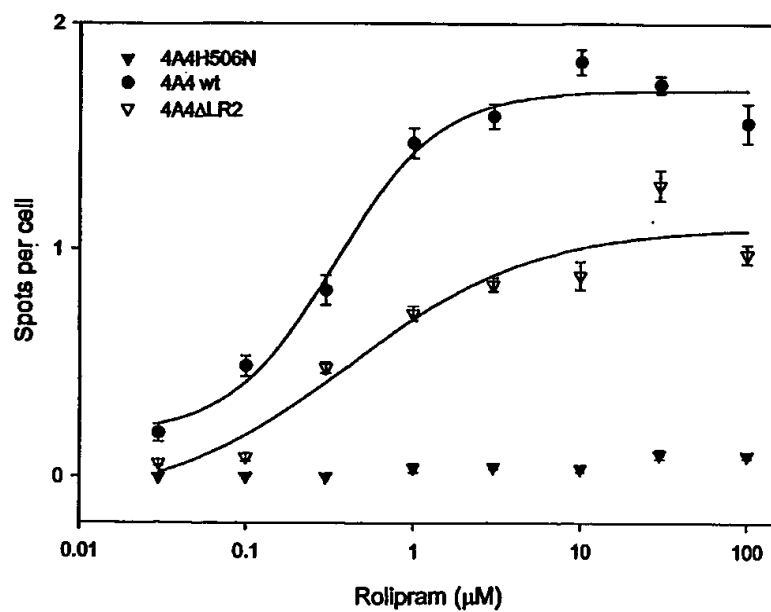
Fig.15

4A4 wt dose-responses



Rolipram dose response (23.5 hr)

Fig.16



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4A4H506N variant

Fig.17

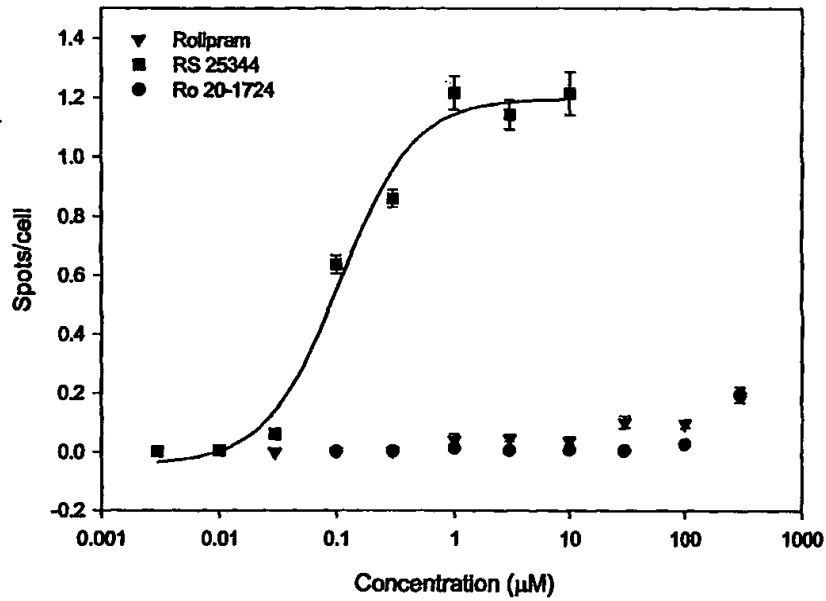
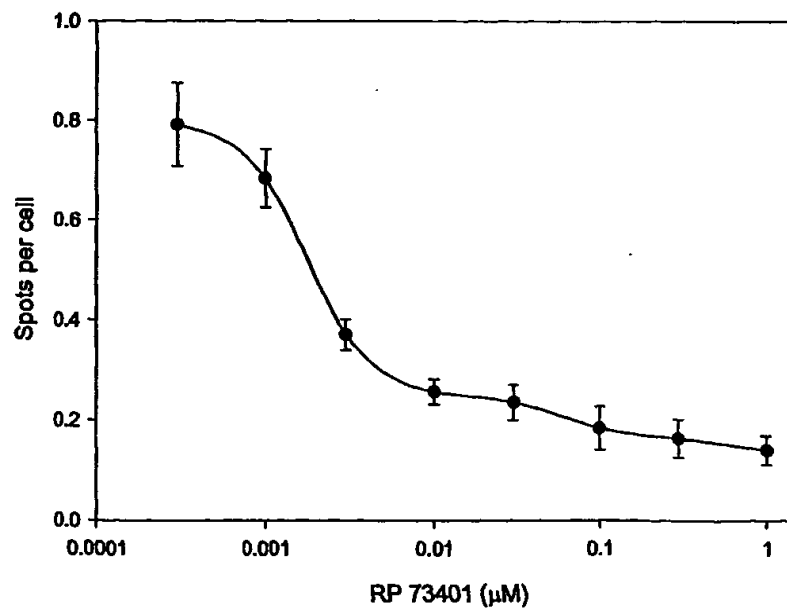


Fig.18 RP 73401 vs. rolipram dose response (23.5 hr)



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Fig. 19a

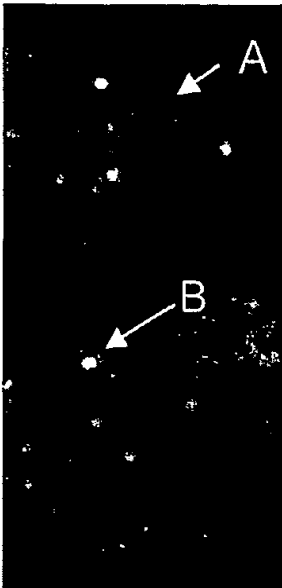


Fig. 19b



Fig. 19c



Fig. 19d



Fig. 20a

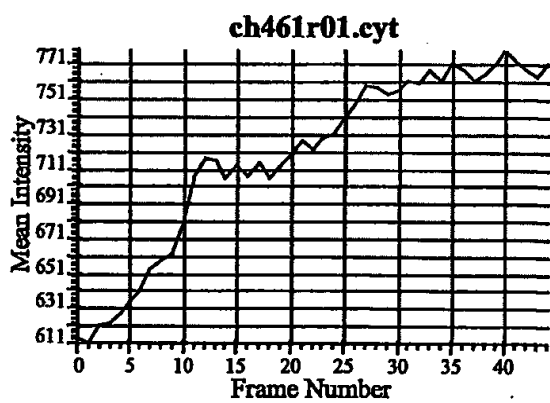
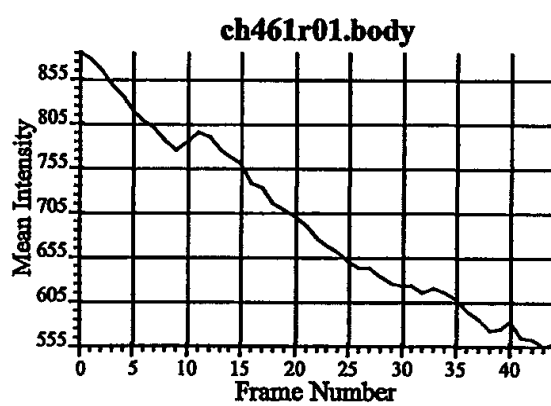
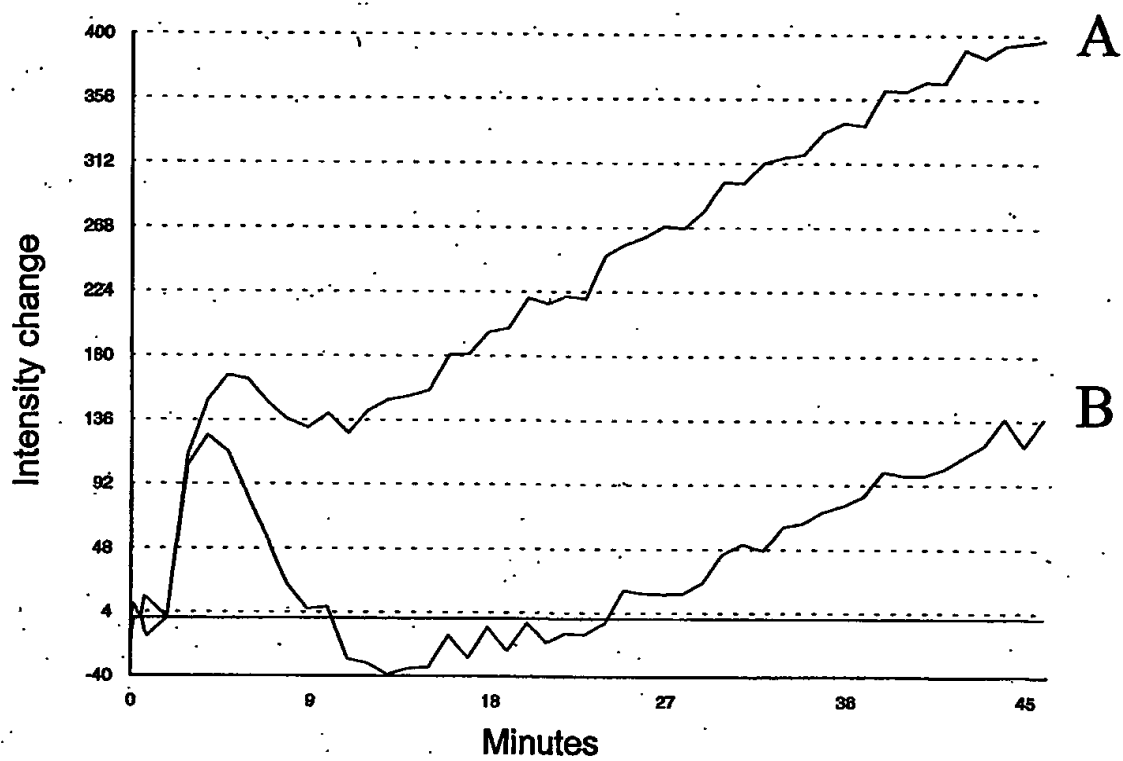


Fig. 20b



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Fig. 21



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Fig. 22

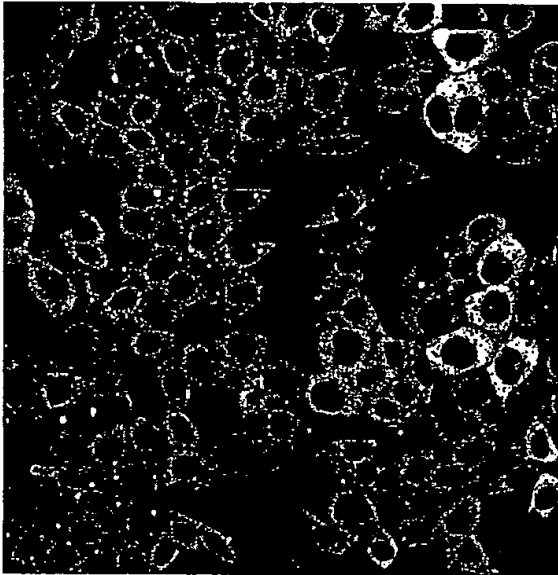


Fig. 23

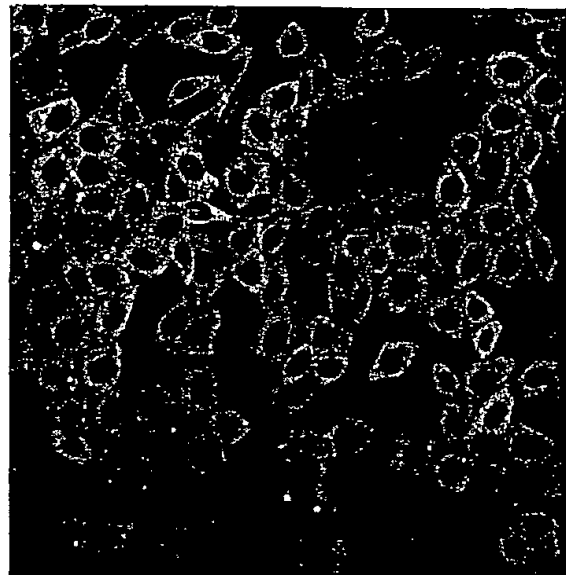


Fig. 24

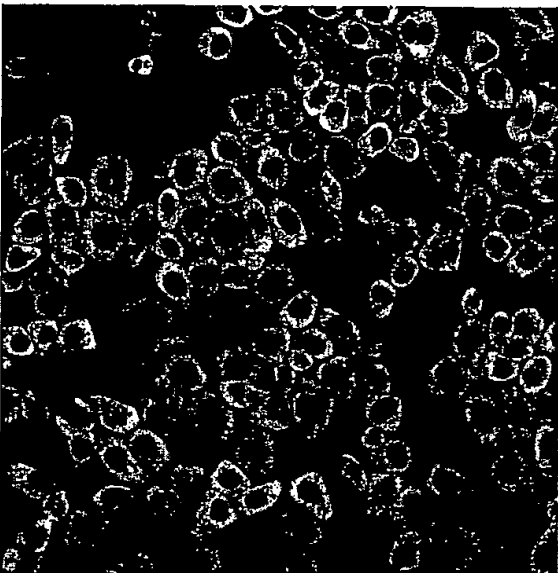
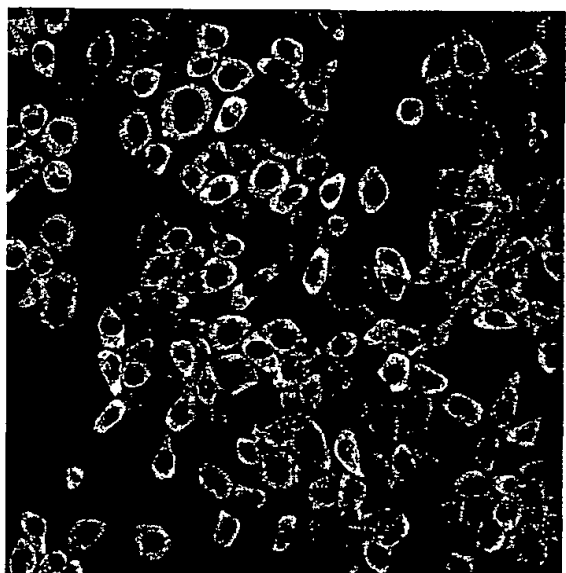


Fig. 25



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Fig. 26a

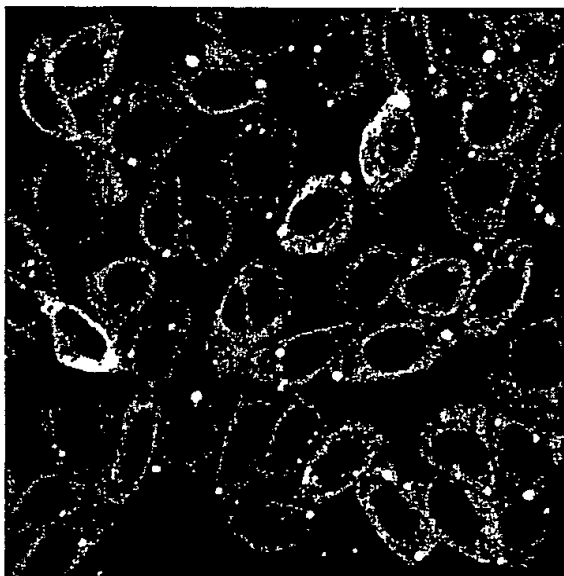


Fig. 26b

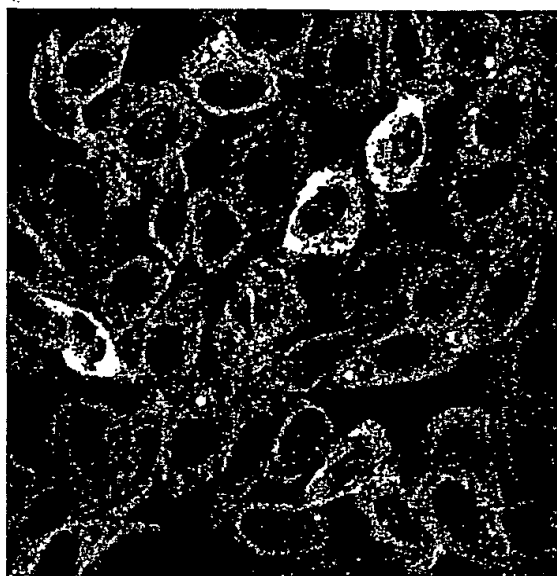


Fig. 27a

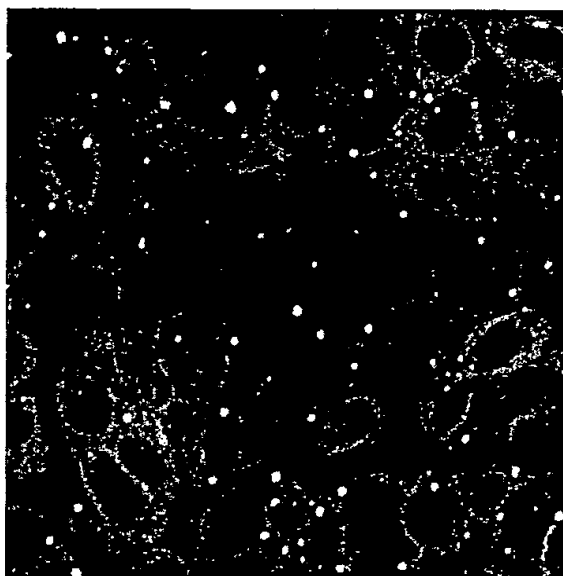


Fig. 27b

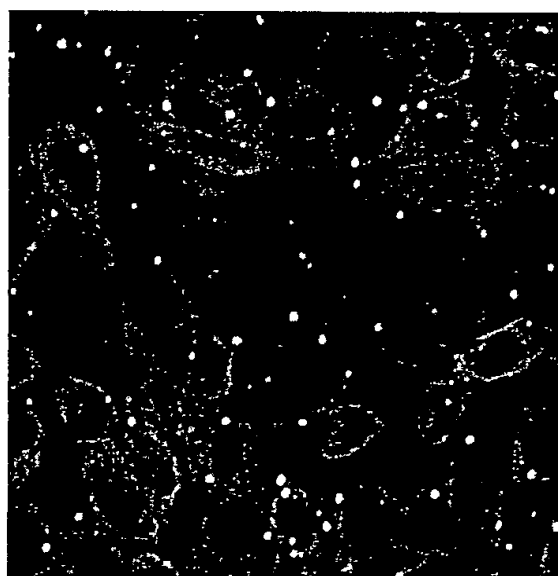


Fig. 28

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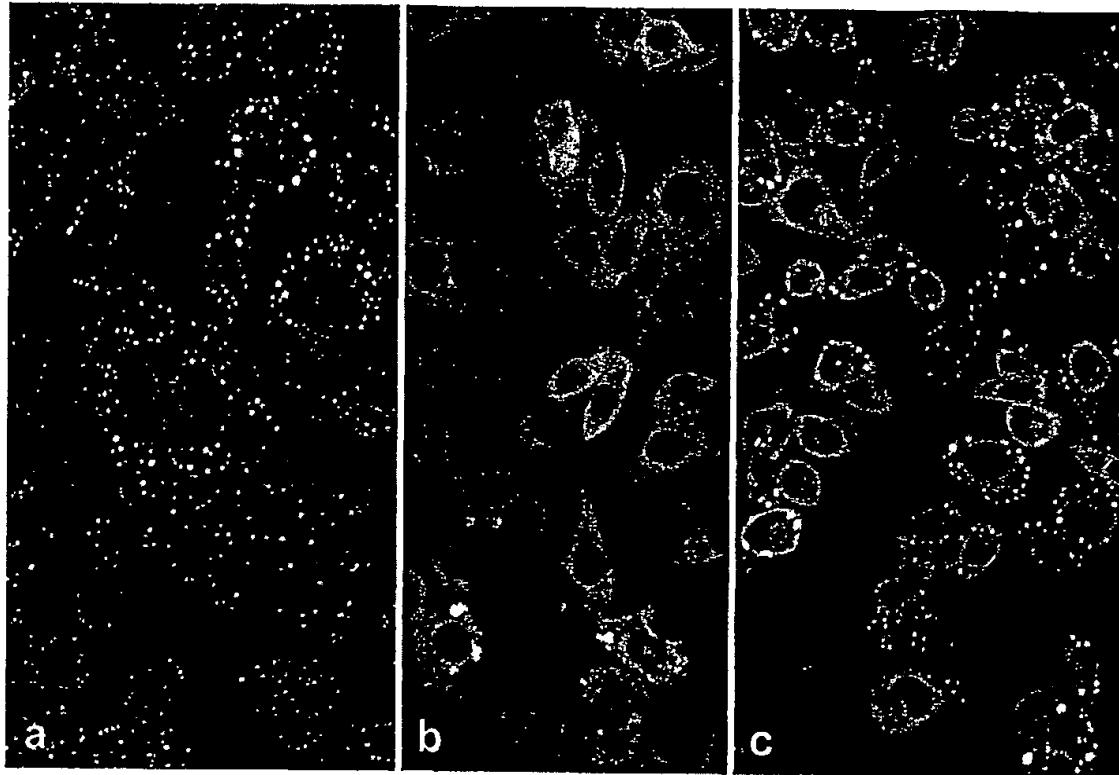
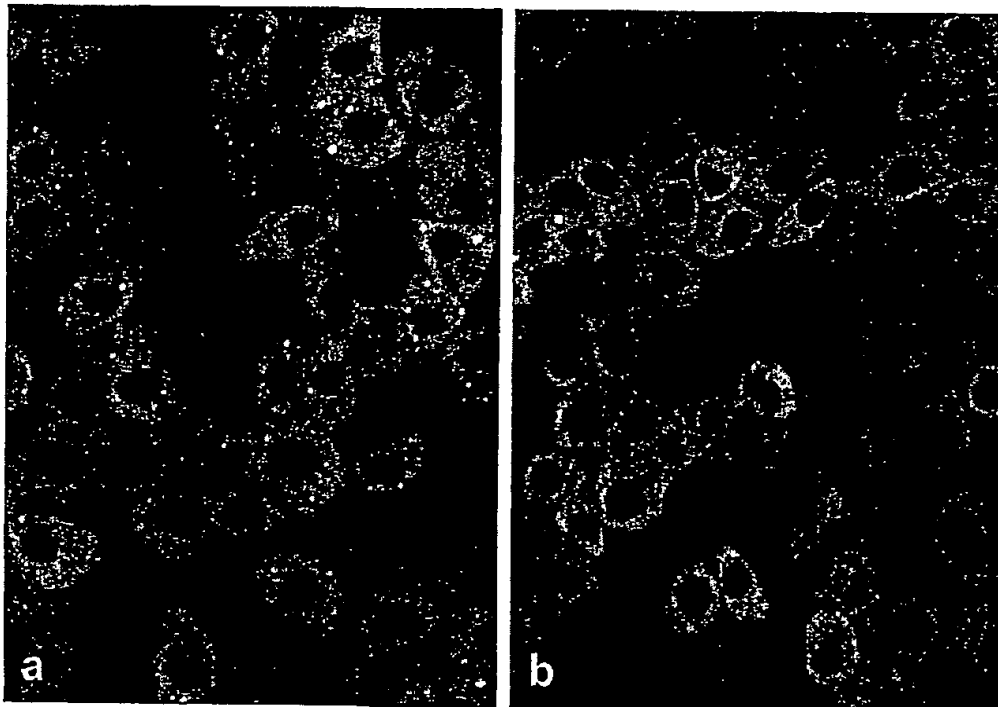
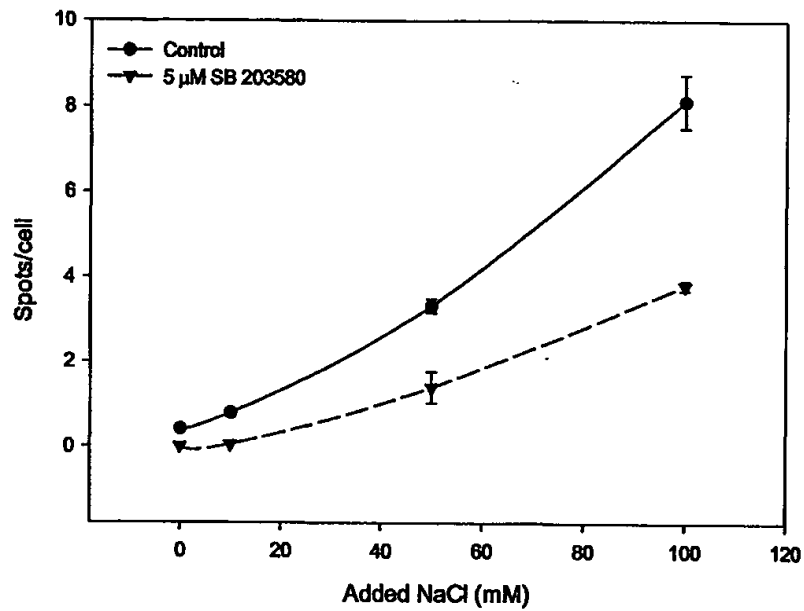


Fig. 29

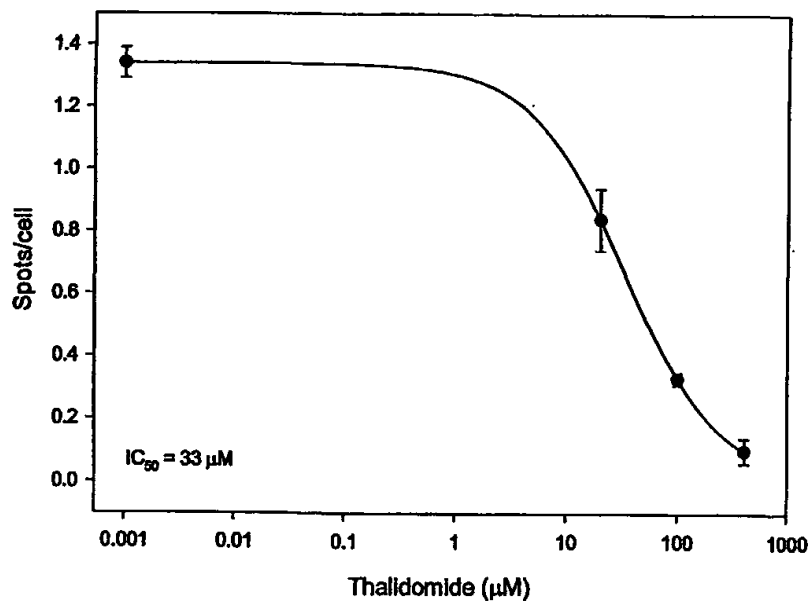


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Spot reappearance with step-increased osmotic strength
CHO462-0, 4 °C 4 hrs

Fig. 30**Fig. 31**

Thalidomide effect on spot reappearance
4 hr at ambient



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Fig. 32a

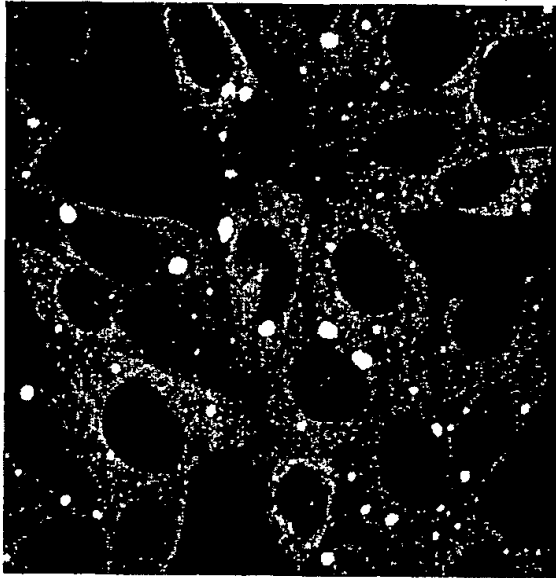


Fig. 32b

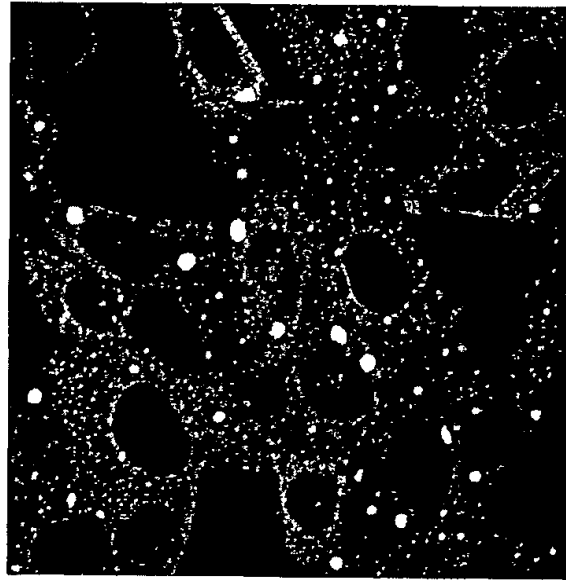


Fig. 33a

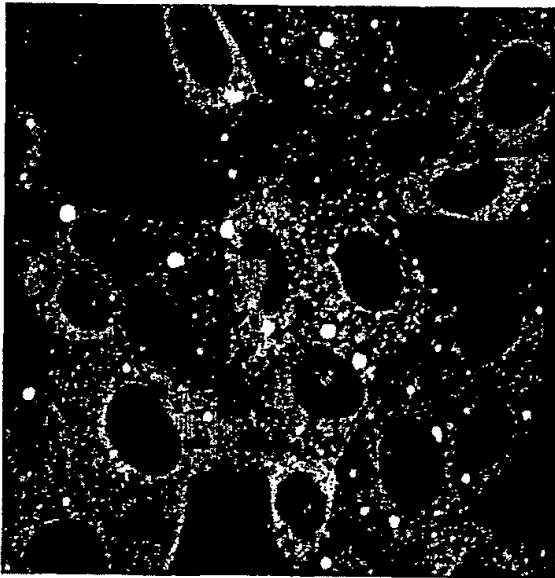
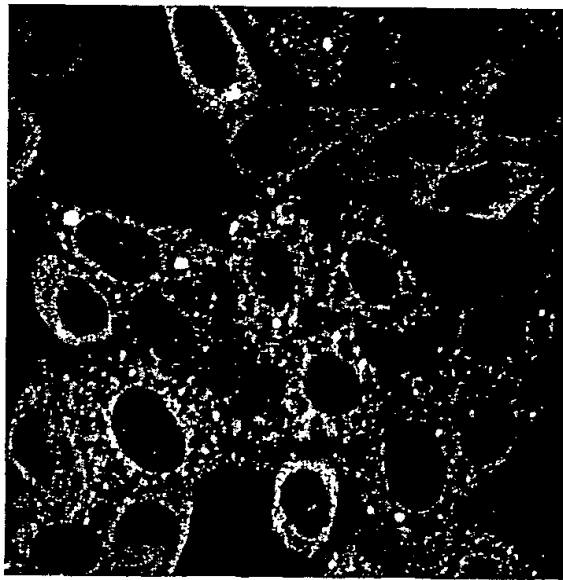


Fig. 33b



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Fig. 34a

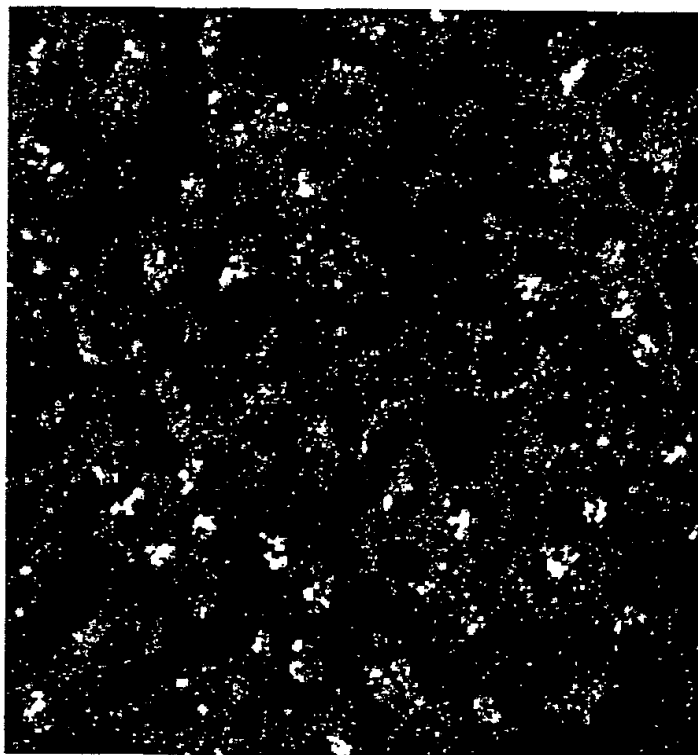
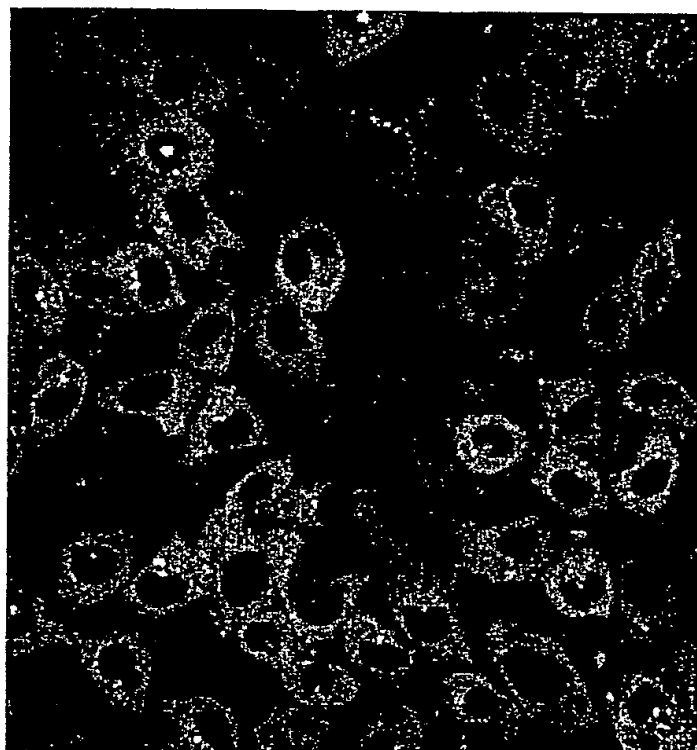


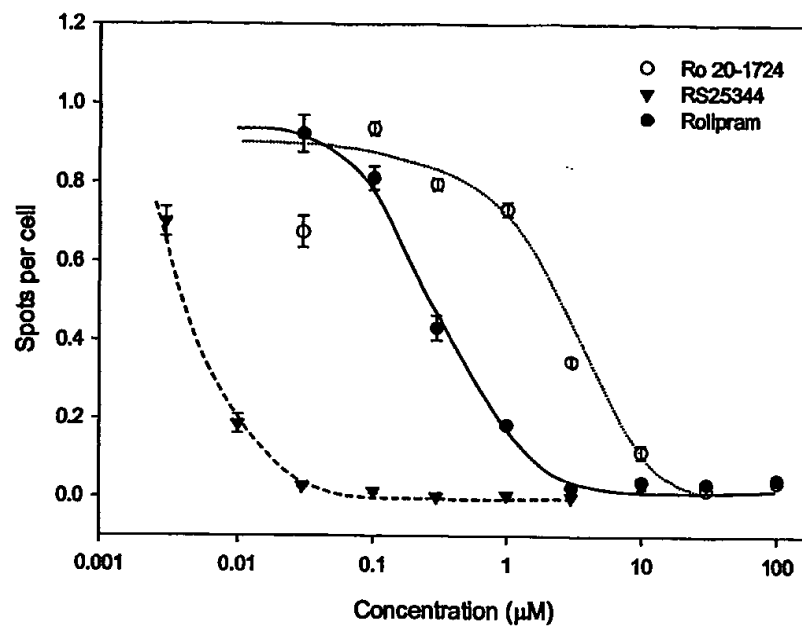
Fig. 34b



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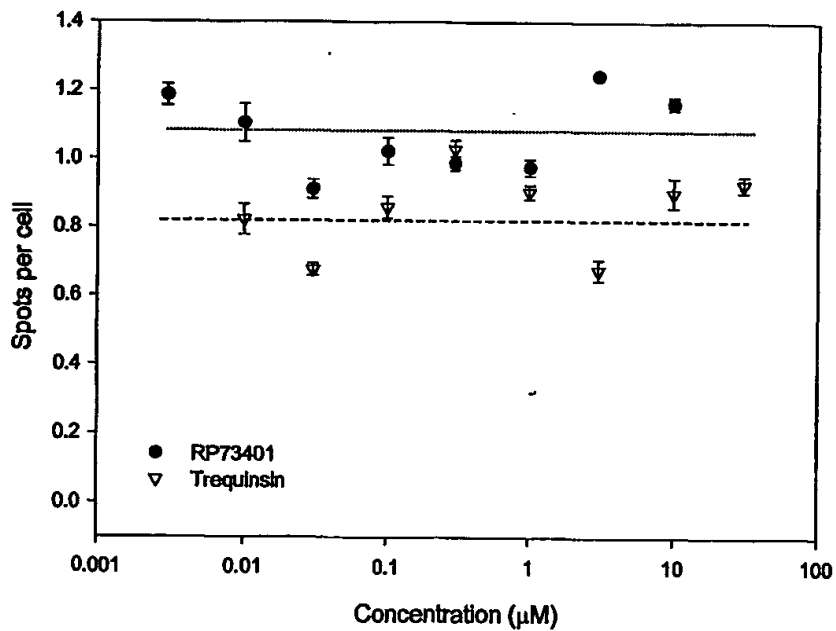
Redistribution of 4A1-eGFP in CHO

Fig. 35



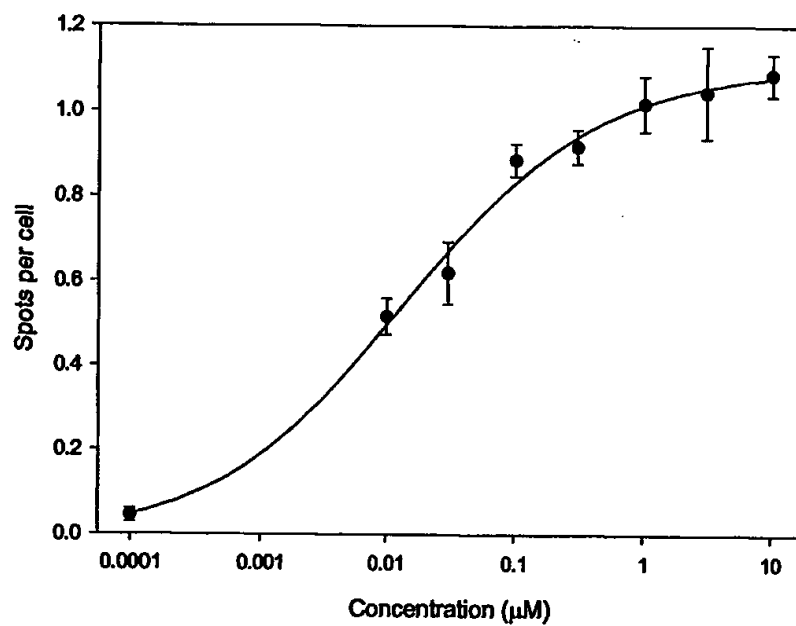
Redistribution of 4A1-eGFP in CHO

Fig. 36



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Fig. 37

RP73401 vs 3 μ M Rolipram (6 hr)

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Fig.38

4A4 wt: Ariflo vs. RS25344 and rolipram

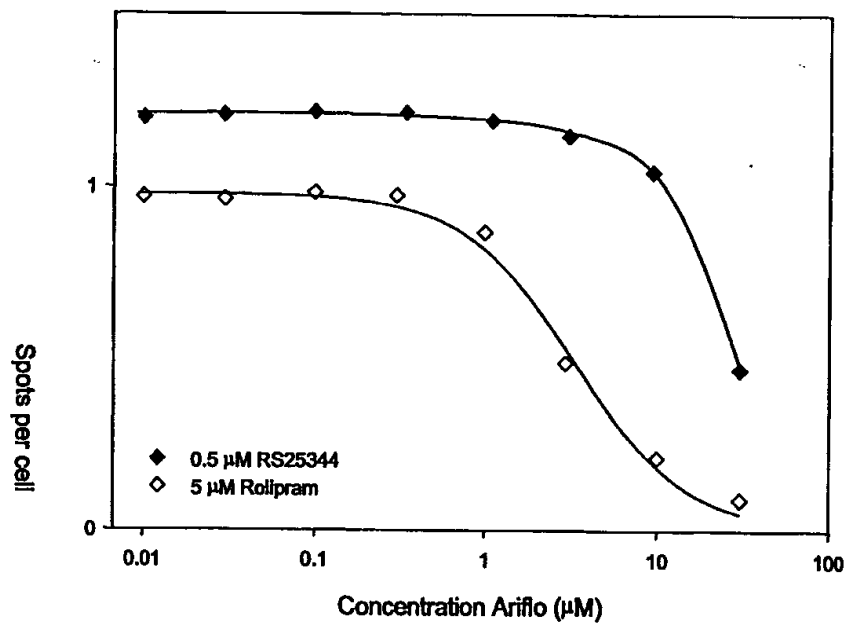
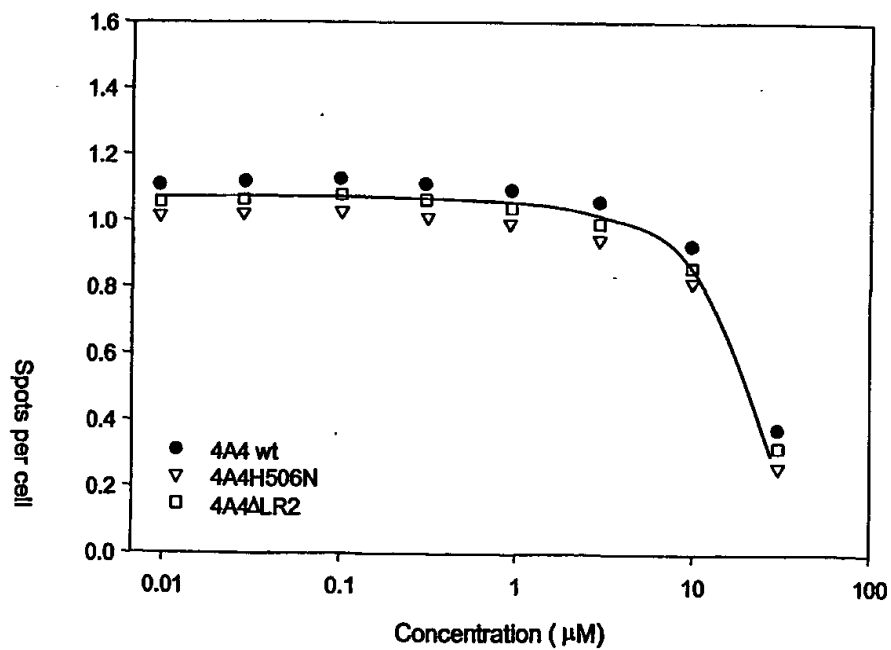


Fig.39

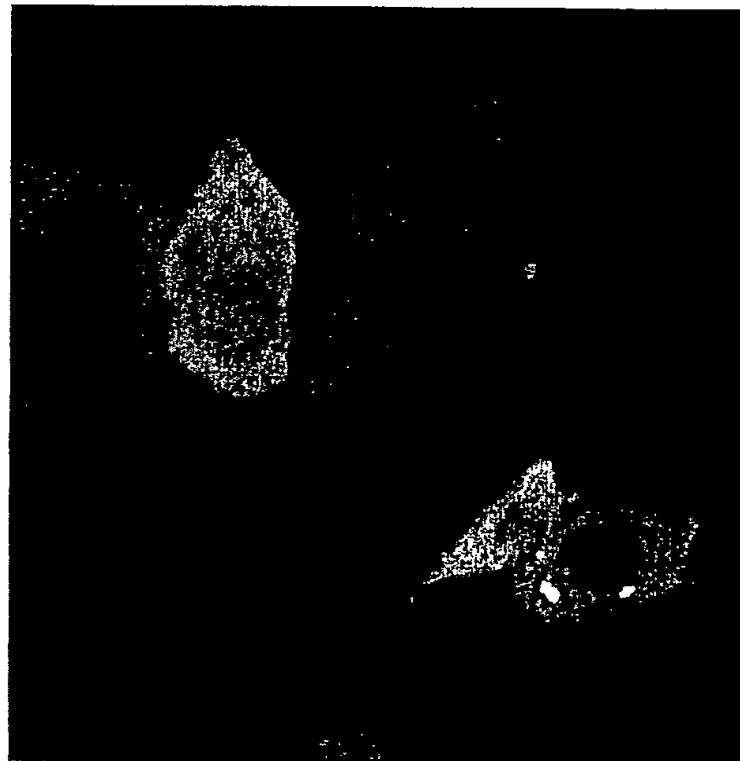
Ariflo vs 0.5 μM RS25344

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Fig. 40



Fig. 41



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PDE4A4-E222G
Stress Spots vs. RP73401

Fig. 42

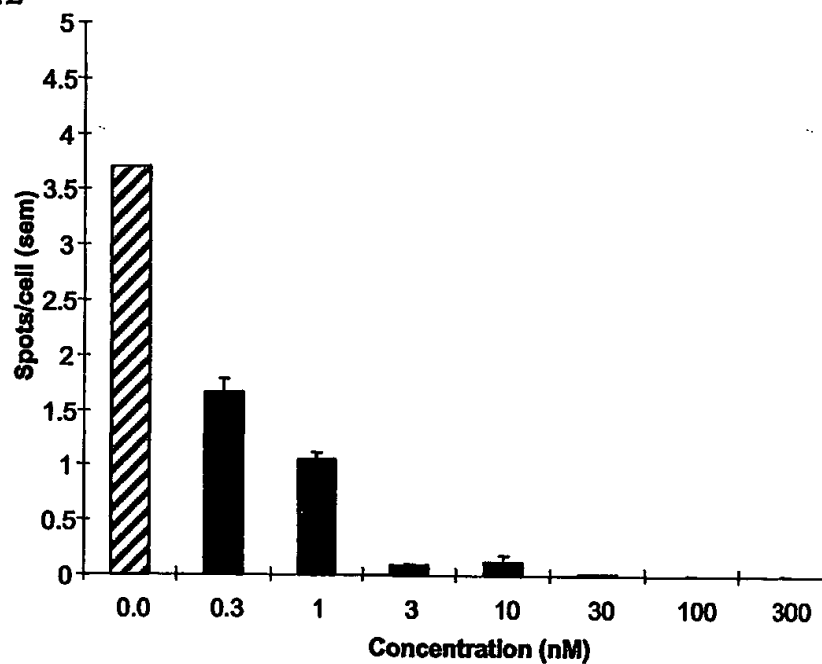
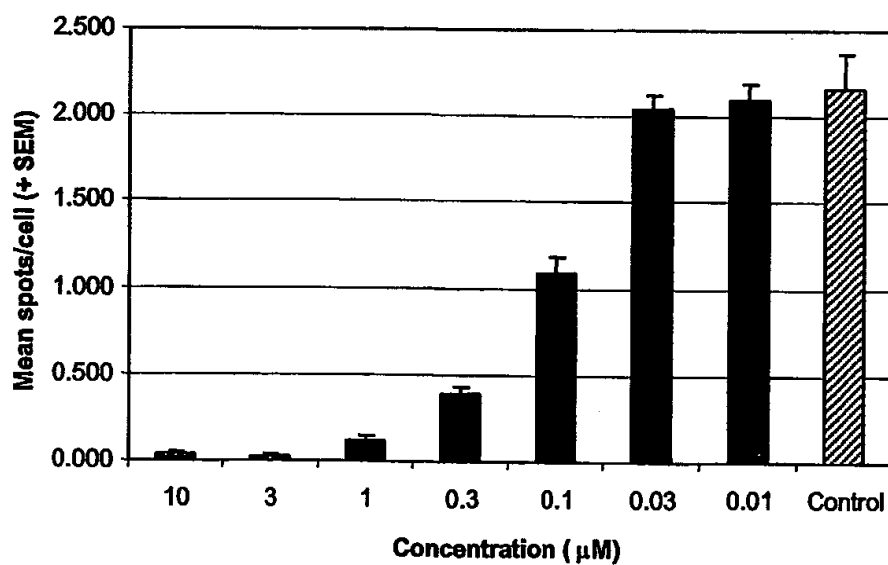


Fig. 43

PDE4A1-E222G vs. Rolipram



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His Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile	
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Gln Pro Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile	
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ttc aag gag gac ggc aac atc ctg ggg cac aag ctg gag tac aac tac Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr 785 790 795 800			2400
aac agc cac aac gtc tat atc atg gcc gac aag cag aag aac ggc atc Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile 805 810 815			2448
aag gtg aac ttc aag atc cgc cac aac atc gag gac ggc agc gtg cag Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln 820 825 830			2496
ctc gcc gac cac tac cag cag aac acc ccc atc ggc gac ggc ccc gtg			2544

Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val
 835 840 845

ctg ctg ccc gac aac cac tac ctg agc acc cag tcc gcc ctg agc aaa 2592
 Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys
 850 855 860

gac ccc aac gag aag cgc gat cac atg gtc ctg ctg gag ttc gtg acc 2640
 Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr
 865 870 875 880

gcc gcc ggg atc act ctc ggc atg gac gag ctg tac aag taa 2682
 Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys * * *
 885 890

<210> 2

<211> 893

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion between Aequorea victoria and human

<400> 2

Met Pro Leu Val Asp Phe Phe Cys Glu Thr Cys Ser Lys Pro Trp Leu
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 Val Gly Trp Trp Asp Gln Phe Lys Arg Met Leu Asn Arg Glu Leu Thr
 20 25 30
 His Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile
 35 40 45
 Ser Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro
 50 55 60
 Thr Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser
 65 70 75 80
 Gln Pro Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile
 85 90 95
 Thr Gly Leu Lys Lys Leu Met His Ser Asn Ser Leu Asn Asn Ser Asn
 100 105 110
 Ile Pro Arg Phe Gly Val Lys Thr Asp Gln Glu Glu Leu Leu Ala Gln
 115 120 125
 Glu Leu Glu Asn Leu Asn Lys Trp Gly Leu Asn Ile Phe Cys Val Ser
 130 135 140
 Asp Tyr Ala Gly Gly Arg Ser Leu Thr Cys Ile Met Tyr Met Ile Phe
 145 150 155 160
 Gln Glu Arg Asp Leu Leu Lys Lys Phe Arg Ile Pro Val Asp Thr Met
 165 170 175
 Val Thr Tyr Met Leu Thr Leu Glu Asp His Tyr His Ala Asp Val Ala
 180 185 190
 Tyr His Asn Ser Leu His Ala Ala Asp Val Leu Gln Ser Thr His Val
 195 200 205
 Leu Leu Ala Thr Pro Ala Leu Asp Ala Val Phe Thr Asp Leu Glu Ile
 210 215 220
 Leu Ala Ala Leu Phe Ala Ala Ala Ile His Asp Val Asp His Pro Gly
 225 230 235 240
 Val Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met
 245 250 255
 Tyr Asn Asp Glu Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe
 260 265 270

Lys Leu Leu Gln Glu Asp Asn Cys Asp Ile Phe Gln Asn Leu Ser Lys
 275 280 285
 Arg Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp Met Val Leu Ala
 290 295 300
 Thr Asp Met Ser Lys His Met Thr Leu Leu Ala Asp Leu Lys Thr Met
 305 310 315 320
 Val Glu Thr Lys Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn
 325 330 335
 Tyr Ser Asp Arg Ile Gln Val Leu Arg Asn Met Val His Cys Ala Asp
 340 345 350
 Leu Ser Asn Pro Thr Lys Pro Leu Glu Leu Tyr Arg Gln Trp Thr Asp
 355 360 365
 Arg Ile Met Ala Glu Phe Phe Gln Gln Gly Asp Arg Glu Arg Glu Arg
 370 375 380
 Gly Met Glu Ile Ser Pro Met Cys Asp Lys His Thr Ala Ser Val Glu
 385 390 395 400
 Lys Ser Gln Val Gly Phe Ile Asp Tyr Ile Val His Pro Leu Trp Glu
 405 410 415
 Thr Trp Ala Asp Leu Val His Pro Asp Ala Gln Glu Ile Leu Asp Thr
 420 425 430
 Leu Glu Asp Asn Arg Asp Trp Tyr Tyr Ser Ala Ile Arg Gln Ser Pro
 435 440 445
 Ser Pro Pro Pro Glu Glu Glu Ser Arg Gly Pro Gly His Pro Pro Leu
 450 455 460
 Pro Asp Lys Phe Gln Phe Glu Leu Thr Leu Glu Glu Glu Glu Glu
 465 470 475 480
 Glu Ile Ser Met Ala Gln Ile Pro Cys Thr Ala Gln Glu Ala Leu Thr
 485 490 495
 Ala Gln Gly Leu Ser Gly Val Glu Glu Ala Leu Asp Ala Thr Ile Ala
 500 505 510
 Trp Glu Ala Ser Pro Ala Gln Glu Ser Leu Glu Val Met Ala Gln Glu
 515 520 525
 Ala Ser Leu Glu Ala Glu Leu Glu Ala Val Tyr Leu Thr Gln Gln Ala
 530 535 540
 Gln Ser Thr Gly Ser Ala Pro Val Ala Pro Asp Glu Phe Ser Ser Arg
 545 550 555 560
 Glu Glu Phe Val Val Ala Val Ser His Ser Ser Pro Ser Ala Leu Ala
 565 570 575
 Leu Gln Ser Pro Leu Leu Pro Ala Trp Arg Thr Leu Ser Val Ser Glu
 580 585 590
 His Ala Pro Gly Leu Pro Gly Leu Pro Ser Thr Ala Ala Glu Val Glu
 595 600 605
 Ala Gln Arg Glu His Gln Ala Ala Lys Arg Ala Cys Ser Ala Cys Ala
 610 615 620
 Gly Thr Phe Gly Glu Asp Thr Ser Ala Leu Pro Ala Pro Gly Gly Gly
 625 630 635 640
 Gly Ser Gly Gly Asp Pro Thr Trp Asp Pro Pro Val Ala Thr Met Val
 645 650 655
 Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu
 660 665 670
 Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly
 675 680 685
 Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr
 690 695 700
 Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr
 705 710 715 720
 Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His
 725 730 735
 Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr

			740					745					750			
Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	
		755					760					765				
Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	
		770					775					780				
Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Tyr	
785					790					795					800	
Asn	Ser	His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	
			805						810						815	
Lys	Val	Asn	Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Ser	Val	Gln	
			820					825					830			
Leu	Ala	Asp	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	
		835					840					845				
Leu	Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Ala	Leu	Ser	Lys	
		850				855					860					
Asp	Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Glu	Phe	Val	Thr	
865					870					875					880	
Ala	Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys				
			885						890							

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<210> 3
<211> 3399
<212> DNA
<213> Artificial Sequence
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<220>
<221> CDS
<222> (1)...(3399)
<223> Fusion between Aequorea victoria and human
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<400> 3																
atg	gaa	ccc	ccg	acc	gtc	ccc	tcg	gaa	agg	agc	ctg	tct	ctg	tca	ctg	48
Met	Glu	Pro	Pro	Thr	Val	Pro	Ser	Glu	Arg	Ser	Leu	Ser	Leu	Ser	Leu	
1		5				10				15						
ccc	ggg	ccc	cgg	gag	ggc	cag	gcc	acc	ctg	aag	cct	ccc	ccg	cag	cac	96
Pro	Gly	Pro	Arg	Glu	Gly	Gln	Ala	Thr	Leu	Lys	Pro	Pro	Pro	Gln	His	
			20		25				30							
ctg	tgg	cgg	cag	cct	cgg	acc	ccc	atc	cgt	atc	cag	cag	cgc	ggc	tac	144
Leu	Trp	Arg	Gln	Pro	Arg	Thr	Pro	Ile	Arg	Ile	Gln	Gln	Arg	Gly	Tyr	
		35		40				45								
tcc	gac	agc	gcg	gag	cgc	gcc	gag	cgg	gag	cgg	cag	ccg	cac	cgg	ccc	192
Ser	Asp	Ser	Ala	Glu	Arg	Ala	Glu	Arg	Glu	Arg	Gln	Pro	His	Arg	Pro	
50		55				60										
ata	gag	cgc	gcc	gat	gcc	atg	gac	acc	agc	gac	cgg	ccc	ggc	ctg	cgc	240
Ile	Glu	Arg	Ala	Asp	Ala	Met	Asp	Thr	Ser	Asp	Arg	Pro	Gly	Leu	Arg	
65		70				75				80						
acg	acc	cgc	atg	tcc	tgg	ccc	tcg	tcc	ttc	cat	ggc	act	ggc	acc	ggc	288
Thr	Thr	Arg	Met	Ser	Trp	Pro	Ser	Ser	Phe	His	Gly	Thr	Gly	Thr	Gly	
				85		90				95						
agc	ggc	ggc	gcg	ggc	gga	ggc	agc	agc	agg	cgc	ttc	gag	gca	gag	aat	336
Ser	Gly	Gly	Ala	Gly	Gly	Gly	Ser	Ser	Arg	Arg	Phe	Glu	Ala	Glu	Asn	
			100		105				110							

ggg ccg aca cca tct cct ggc cgc agc ccc ctg gac tcg cag gcg agc Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser 115 120 125	384
cca gga ctc gtg ctg cac gcc ggg gcg gcc acc agc cag cgc cgg gag Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu 130 135 140	432
tcc ttc ctg tac cgc tca gac agc gac tat gac atg tca ccc aag acc Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp Met Ser Pro Lys Thr 145 150 155 160	480
atg tcc cgg aac tca tcg gtc acc agc gag gcg cac gct gaa gac ctc Met Ser Arg Asn Ser Ser Val Thr Ser Glu Ala His Ala Glu Asp Leu 165 170 175	528
atc gta aca cca ttt gct cag gtg ctg gcc agc ctc cgg agc gtc cgt Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser Leu Arg Ser Val Arg 180 185 190	576
agc aac ttc tca ctc ctg acc aat gtg ccc gtt ccc agt aac aag cgg Ser Asn Phe Ser Leu Leu Thr Asn Val Pro Val Pro Ser Asn Lys Arg 195 200 205	624
tcc ccg ctg ggc ggc ccc acc cct gtc tgc aag gcc acg ctg tca gaa Ser Pro Leu Gly Gly Pro Thr Pro Val Cys Lys Ala Thr Leu Ser Glu 210 215 220	672
gaa acg tgt cag cag ttg gcc cgg gag act ctg gag gag ctg gac tgg Glu Thr Cys Gln Gln Leu Ala Arg Glu Thr Leu Glu Glu Leu Asp Trp 225 230 235 240	720
tgt ctg gag cag ctg gag acc atg cag acc tat cgc tct gtc agc gag Cys Leu Glu Gln Leu Glu Thr Met Gln Thr Tyr Arg Ser Val Ser Glu 245 250 255	768
atg gcc tcg cac aag ttt aaa agg atg ttg aac cgt gag ctc aca cac Met Ala Ser His Lys Phe Lys Arg Met Leu Asn Arg Glu Leu Thr His 260 265 270	816
ctg tca gaa atg agc agg tcc gga aac cag gtc tca gag tac att tcc Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile Ser 275 280 285	864
aca aca ttc ctg gac aaa cag aat gaa gtg gag atc cca tca ccc acg Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro Thr 290 295 300	912
atg aag gaa cga gaa aaa cag caa gcg ccg cga cca aga ccc tcc cag Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser Gln 305 310 315 320	960
ccg ccc ccg ccc cct gta cca cac tta cag ccc atg tcc caa atc aca Pro Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile Thr 325 330 335	1008
ggg ttg aaa aag ttg atg cat agt aac agc ctg aac aac tct aac att Gly Leu Lys Lys Leu Met His Ser Asn Ser Leu Asn Asn Ser Asn Ile 340 345 350	1056

ccc cga ttt ggg gtg aag acc gat caa gaa gag ctc ctg gcc caa gaa Pro Arg Phe Gly Val Lys Thr Asp Gln Glu Glu Leu Leu Ala Gln Glu 355 360 365	1104
ctg gag aac ctg aac aag tgg ggc ctg aac atc ttt tgc gtg tcg gat Leu Glu Asn Leu Asn Lys Trp Gly Leu Asn Ile Phe Cys Val Ser Asp 370 375 380	1152
tac gct gga ggc cgc tca ctc acc tgc atc atg tac atg ata ttc cag Tyr Ala Gly Gly Arg Ser Leu Thr Cys Ile Met Tyr Met Ile Phe Gln 385 390 395 400	1200
gag cgg gac ctg ctg aag aaa ttc cgc atc ccg gtg gac acg atg gtg Glu Arg Asp Leu Leu Lys Lys Phe Arg Ile Pro Val Asp Thr Met Val 405 410 415	1248
aca tac atg ctg acg ctg gag gat cac tac cac gct gac gtg gcc tac Thr Tyr Met Leu Thr Leu Glu Asp His Tyr His Ala Asp Val Ala Tyr 420 425 430	1296
cat aac agc ctg cac gca gct gac gtg ctg cag tcc acc cac gta ctg His Asn Ser Leu His Ala Ala Asp Val Leu Gln Ser Thr His Val Leu 435 440 445	1344
ctg gcc acg cct gca cta gat gca gtg ttc acg gac ctg gag att ctc Leu Ala Thr Pro Ala Leu Asp Ala Val Phe Thr Asp Leu Glu Ile Leu 450 455 460	1392
gcc gcc ctc ttc gcg gct gcc atc cac gat gtg gat cac cct ggg gtc Ala Ala Leu Phe Ala Ala Ala Ile His Asp Val Asp His Pro Gly Val 465 470 475 480	1440
tcc aac cag ttc ctc atc aac acc aat tcg gag ctg gcg ctc atg tac Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met Tyr 485 490 495	1488
aac gat gag tcg gtg ctc gag aat cac cac ctg gcc gtg ggc ttc aag Asn Asp Glu Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe Lys 500 505 510	1536
ctg ctg cag gag gac aac tgc gac atc ttc cag aac ctc agc aag cgc Leu Leu Gln Glu Asp Asn Cys Asp Ile Phe Gln Asn Leu Ser Lys Arg 515 520 525	1584
cag cgg cag agc cta cgc aag atg gtc atc gac atg gtg ctg gcc acg Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp Met Val Leu Ala Thr 530 535 540	1632
gac atg tcc aag cac atg acc ctc ctg gct gac ctg aag acc atg gtg Asp Met Ser Lys His Met Thr Leu Leu Ala Asp Leu Lys Thr Met Val 545 550 555 560	1680
gag acc aag aaa gtg acc agc tca ggg gtc ctc ctg cta gat aac tac Glu Thr Lys Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn Tyr 565 570 575	1728
tcc gac cgc atc cag gtc ctc cgg aac atg gtg cac tgt gcc gac ctc Ser Asp Arg Ile Gln Val Leu Arg Asn Met Val His Cys Ala Asp Leu	1776

580						585						590						
agc aac ccc acc aag ccg ctg gag ctg tac cgc cag tgg aca gac cgc	1824																	
Ser Asn Pro Thr Lys Pro Leu Glu Leu Tyr Arg Gln Trp Thr Asp Arg																		
595 600 605																		
atc atg gcc gag ttc ttc cag cag ggt gac cga gag cgc gag cgt ggc	1872																	
Ile Met Ala Glu Phe Phe Gln Gln Gly Asp Arg Glu Arg Glu Arg Gly																		
610 615 620																		
atg gaa atc agc ccc atg tgt gac aag cac act gcc tcc gtg gag aag	1920																	
Met Glu Ile Ser Pro Met Cys Asp Lys His Thr Ala Ser Val Glu Lys																		
625 630 635 640																		
tct cag gtg ggt ttt att gac tac att gtg cac cca ttg tgg gag acc	1968																	
Ser Gln Val Gly Phe Ile Asp Tyr Ile Val His Pro Leu Trp Glu Thr																		
645 650 655																		
tgg gcg gac ctt gtc cac cca gat gcc cag gag atc ttg gac act ttg	2016																	
Trp Ala Asp Leu Val His Pro Asp Ala Gln Glu Ile Leu Asp Thr Leu																		
660 665 670																		
gag gac aac cgg gac tgg tac tac agc gcc atc cgg cag agc cca tct	2064																	
Glu Asp Asn Arg Asp Trp Tyr Tyr Ser Ala Ile Arg Gln Ser Pro Ser																		
675 680 685																		
ccg cca ccc gag gag gag tca agg ggg cca ggc cac cca ccc ctg cct	2112																	
Pro Pro Pro Glu Glu Glu Ser Arg Gly Pro Gly His Pro Pro Leu Pro																		
690 695 700																		
gac aag ttc cag ttt gag ctg acg ctg gag gag gaa gag gag gaa gaa	2160																	
Asp Lys Phe Gln Phe Glu Leu Thr Leu Glu Glu Glu Glu Glu Glu																		
705 710 715 720																		
ata tca atg gcc cag ata ccg tgc aca gcc caa gag gca ttg act gcg	2208																	
Ile Ser Met Ala Gln Ile Pro Cys Thr Ala Gln Glu Ala Leu Thr Ala																		
725 730 735																		
cag gga ttg tca gga gtc gag gaa gct ctg gat gca acc ata gcc tgg	2256																	
Gln Gly Leu Ser Gly Val Glu Glu Ala Leu Asp Ala Thr Ile Ala Trp																		
740 745 750																		
gag gca tcc ccg gcc cag gag tcg ttg gaa gtt atg gca cag gaa gca	2304																	
Glu Ala Ser Pro Ala Gln Glu Ser Leu Glu Val Met Ala Gln Glu Ala																		
755 760 765																		
tcc ctg gag gcc gag ctg gag gca gtg tat ttg aca cag cag gca cag	2352																	
Ser Leu Glu Ala Glu Leu Glu Ala Val Tyr Leu Thr Gln Gln Ala Gln																		
770 775 780																		
tcc aca ggc agt gca cct gtg gct ccg gat gag ttc tcg tcc cgg gag	2400																	
Ser Thr Gly Ser Ala Pro Val Ala Pro Asp Glu Phe Ser Ser Arg Glu																		
785 790 795 800																		
gaa ttc gtg gtt gct gta agc cac agc agc ccc tct gcc ctg gct ctt	2448																	
Glu Phe Val Val Ala Val Ser His Ser Ser Pro Ser Ala Leu Ala Leu																		
805 810 815																		
caa agc ccc ctt ctc cct gct tgg agg acc ctg tct gtt tca gag cat	2496																	

Gln	Ser	Pro	Leu	Leu	Pro	Ala	Trp	Arg	Thr	Leu	Ser	Val	Ser	Glu	His	
			820					825					830			
gcc	ccg	ggc	ctc	ccg	ggc	ctc	ccc	tcc	acg	gcg	gcc	gag	gtg	gag	gcc	2544
Ala	Pro	Gly	Leu	Pro	Gly	Leu	Pro	Ser	Thr	Ala	Ala	Glu	Val	Glu	Ala	
		835					840					845				
caa	cga	gag	cac	cag	gct	gcc	aag	agg	gct	tgc	agt	gcc	tgc	gca	ggg	2592
Gln	Arg	Glu	His	Gln	Ala	Ala	Lys	Arg	Ala	Cys	Ser	Ala	Cys	Ala	Gly	
	850					855					860					
aca	ttt	ggg	gag	gac	aca	tcc	gca	ctc	cca	gct	cct	ggt	ggc	ggg	ggg	2640
Thr	Phe	Gly	Glu	Asp	Thr	Ser	Ala	Leu	Pro		Pro	Gly	Gly	Gly	Gly	
865					870				875						880	
tca	ggt	gga	gac	cct	acc	tgg	gat	cca	ccg	gtc	gcc	acc	atg	gtg	agc	2688
Ser	Gly	Gly	Asp	Pro	Thr	Trp	Asp	Pro	Pro	Val	Ala	Thr	Met	Val	Ser	
			885					890					895			
aag	ggc	gag	gag	ctg	ttc	acc	ggg	gtg	gtg	ccc	atc	ctg	gtc	gag	ctg	2736
Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	
		900					905					910				
gac	ggc	gac	gta	aac	ggc	cac	aag	ttc	agc	gtg	tcc	ggc	gag	ggc	gag	2784
Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	
	915						920					925				
ggc	gat	gcc	acc	tac	ggc	aag	ctg	acc	ctg	aag	ttc	atc	tgc	acc	acc	2832
Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	
	930					935					940					
ggc	aag	ctg	ccc	gtg	ccc	tgg	ccc	acc	ctc	gtg	acc	acc	ctg	acc	tac	2880
Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	
945					950				955						960	
ggc	gtg	cag	tgc	ttc	agc	cgc	tac	ccc	gac	cac	atg	aag	cag	cac	gac	2928
Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp	
			965					970					975			
ttc	ttc	aag	tcc	gcc	atg	ccc	gaa	ggc	tac	gtc	cag	gag	cgc	acc	atc	2976
Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln	Glu	Arg	Thr	Ile	
		980					985					990				
ttc	ttc	aag	gac	gac	ggc	aac	tac	aag	acc	cgc	gcc	gag	gtg	aag	ttc	3024
Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	
		995				1000					1005					
gag	ggc	gac	acc	ctg	gtg	aac	cgc	atc	gag	ctg	aag	ggc	atc	gac	ttc	3072
Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	Phe	
	1010					1015					1020					
aag	gag	gac	ggc	aac	atc	ctg	ggg	cac	aag	ctg	gag	tac	aac	tac	aac	3120
Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Tyr	Asn	
1025					1030				1035						1040	
agc	cac	aac	gtc	tat	atc	atg	gcc	gac	aag	cag	aag	aac	ggc	atc	aag	3168
Ser	His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	
			1045					1050					1055			

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gtg aac ttc aag atc cgc cac aac atc gag gac ggc agc gtg cag ctc      3216
Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu
      1060                      1065                      1070

gcc gac cac tac cag cag aac acc ccc atc ggc gac ggc ccc gtg ctg      3264
Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu
      1075                      1080                      1085

ctg ccc gac aac cac tac ctg agc acc cag tcc gcc ctg agc aaa gac      3312
Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp
      1090                      1095                      1100

ccc aac gag aag cgc gat cac atg gtc ctg ctg gag ttc gtg acc gcc      3360
Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala
      1105                      1110                      1115                      1120

gcc ggg atc act ctc ggc atg gac gag ctg tac aag taa      3399
Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys *
      1125                      1130

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<210> 4
 <211> 1132
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Fusion between Aequorea victoria and human

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<400> 4
Met Glu Pro Pro Thr Val Pro Ser Glu Arg Ser Leu Ser Leu Ser Leu
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Pro Gly Pro Arg Glu Gly Gln Ala Thr Leu Lys Pro Pro Pro Gln His
 20          25          30
Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr
 35          40          45
Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro
 50          55          60
Ile Glu Arg Ala Asp Ala Met Asp Thr Ser Asp Arg Pro Gly Leu Arg
 65          70          75          80
Thr Thr Arg Met Ser Trp Pro Ser Ser Phe His Gly Thr Gly Thr Gly
 85          90          95
Ser Gly Gly Ala Gly Gly Gly Ser Ser Arg Arg Phe Glu Ala Glu Asn
100          105          110
Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser
115          120          125
Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu
130          135          140
Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp Met Ser Pro Lys Thr
145          150          155          160
Met Ser Arg Asn Ser Ser Val Thr Ser Glu Ala His Ala Glu Asp Leu
165          170          175
Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser Leu Arg Ser Val Arg
180          185          190
Ser Asn Phe Ser Leu Leu Thr Asn Val Pro Val Pro Ser Asn Lys Arg
195          200          205
Ser Pro Leu Gly Gly Pro Thr Pro Val Cys Lys Ala Thr Leu Ser Glu
210          215          220
Glu Thr Cys Gln Gln Leu Ala Arg Glu Thr Leu Glu Glu Leu Asp Trp

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225					230					235				240	
Cys	Leu	Glu	Gln	Leu	Glu	Thr	Met	Gln	Thr	Tyr	Arg	Ser	Val	Ser	Glu
				245					250					255	
Met	Ala	Ser	His	Lys	Phe	Lys	Arg	Met	Leu	Asn	Arg	Glu	Leu	Thr	His
			260					265					270		
Leu	Ser	Glu	Met	Ser	Arg	Ser	Gly	Asn	Gln	Val	Ser	Glu	Tyr	Ile	Ser
		275					280					285			
Thr	Thr	Phe	Leu	Asp	Lys	Gln	Asn	Glu	Val	Glu	Ile	Pro	Ser	Pro	Thr
	290					295				300					
Met	Lys	Glu	Arg	Glu	Lys	Gln	Gln	Ala	Pro	Arg	Pro	Arg	Pro	Ser	Gln
305					310					315					320
Pro	Pro	Pro	Pro	Pro	Val	Pro	His	Leu	Gln	Pro	Met	Ser	Gln	Ile	Thr
				325				330						335	
Gly	Leu	Lys	Lys	Leu	Met	His	Ser	Asn	Ser	Leu	Asn	Asn	Ser	Asn	Ile
		340						345					350		
Pro	Arg	Phe	Gly	Val	Lys	Thr	Asp	Gln	Glu	Glu	Leu	Leu	Ala	Gln	Glu
		355					360					365			
Leu	Glu	Asn	Leu	Asn	Lys	Trp	Gly	Leu	Asn	Ile	Phe	Cys	Val	Ser	Asp
	370					375				380					
Tyr	Ala	Gly	Gly	Arg	Ser	Leu	Thr	Cys	Ile	Met	Tyr	Met	Ile	Phe	Gln
385					390					395					400
Glu	Arg	Asp	Leu	Leu	Lys	Lys	Phe	Arg	Ile	Pro	Val	Asp	Thr	Met	Val
			405					410						415	
Thr	Tyr	Met	Leu	Thr	Leu	Glu	Asp	His	Tyr	His	Ala	Asp	Val	Ala	Tyr
		420						425					430		
His	Asn	Ser	Leu	His	Ala	Ala	Asp	Val	Leu	Gln	Ser	Thr	His	Val	Leu
	435						440					445			
Leu	Ala	Thr	Pro	Ala	Leu	Asp	Ala	Val	Phe	Thr	Asp	Leu	Glu	Ile	Leu
	450					455				460					
Ala	Ala	Leu	Phe	Ala	Ala	Ala	Ile	His	Asp	Val	Asp	His	Pro	Gly	Val
465					470					475					480
Ser	Asn	Gln	Phe	Leu	Ile	Asn	Thr	Asn	Ser	Glu	Leu	Ala	Leu	Met	Tyr
			485					490					495		
Asn	Asp	Glu	Ser	Val	Leu	Glu	Asn	His	His	Leu	Ala	Val	Gly	Phe	Lys
		500						505					510		
Leu	Leu	Gln	Glu	Asp	Asn	Cys	Asp	Ile	Phe	Gln	Asn	Leu	Ser	Lys	Arg
		515					520					525			
Gln	Arg	Gln	Ser	Leu	Arg	Lys	Met	Val	Ile	Asp	Met	Val	Leu	Ala	Thr
	530					535					540				
Asp	Met	Ser	Lys	His	Met	Thr	Leu	Leu	Ala	Asp	Leu	Lys	Thr	Met	Val
545					550					555					560
Glu	Thr	Lys	Lys	Val	Thr	Ser	Ser	Gly	Val	Leu	Leu	Leu	Asp	Asn	Tyr
			565					570						575	
Ser	Asp	Arg	Ile	Gln	Val	Leu	Arg	Asn	Met	Val	His	Cys	Ala	Asp	Leu
		580						585					590		
Ser	Asn	Pro	Thr	Lys	Pro	Leu	Glu	Leu	Tyr	Arg	Gln	Trp	Thr	Asp	Arg
		595					600					605			
Ile	Met	Ala	Glu	Phe	Phe	Gln	Gln	Gly	Asp	Arg	Glu	Arg	Glu	Arg	Gly
	610					615				620					
Met	Glu	Ile	Ser	Pro	Met	Cys	Asp	Lys	His	Thr	Ala	Ser	Val	Glu	Lys
625					630					635					640
Ser	Gln	Val	Gly	Phe	Ile	Asp	Tyr	Ile	Val	His	Pro	Leu	Trp	Glu	Thr
			645					650						655	
Trp	Ala	Asp	Leu	Val	His	Pro	Asp	Ala	Gln	Glu	Ile	Leu	Asp	Thr	Leu
		660						665					670		
Glu	Asp	Asn	Arg	Asp	Trp	Tyr	Tyr	Ser	Ala	Ile	Arg	Gln	Ser	Pro	Ser
		675					680					685			
Pro	Pro	Pro	Glu	Glu	Glu	Ser	Arg	Gly	Pro	Gly	His	Pro	Pro	Leu	Pro
	690					695					700				

Asp Lys Phe Gln Phe Glu Leu Thr Leu Glu Glu Glu Glu Glu Glu
 705 710 715 720
 Ile Ser Met Ala Gln Ile Pro Cys Thr Ala Gln Glu Ala Leu Thr Ala
 725 730 735
 Gln Gly Leu Ser Gly Val Glu Glu Ala Leu Asp Ala Thr Ile Ala Trp
 740 745 750
 Glu Ala Ser Pro Ala Gln Glu Ser Leu Glu Val Met Ala Gln Glu Ala
 755 760 765
 Ser Leu Glu Ala Glu Leu Glu Ala Val Tyr Leu Thr Gln Gln Ala Gln
 770 775 780
 Ser Thr Gly Ser Ala Pro Val Ala Pro Asp Glu Phe Ser Ser Arg Glu
 785 790 795 800
 Glu Phe Val Val Ala Val Ser His Ser Ser Pro Ser Ala Leu Ala Leu
 805 810 815
 Gln Ser Pro Leu Leu Pro Ala Trp Arg Thr Leu Ser Val Ser Glu His
 820 825 830
 Ala Pro Gly Leu Pro Gly Leu Pro Ser Thr Ala Ala Glu Val Glu Ala
 835 840 845
 Gln Arg Glu His Gln Ala Ala Lys Arg Ala Cys Ser Ala Cys Ala Gly
 850 855 860
 Thr Phe Gly Glu Asp Thr Ser Ala Leu Pro Ala Pro Gly Gly Gly Gly
 865 870 875 880
 Ser Gly Gly Asp Pro Thr Trp Asp Pro Pro Val Ala Thr Met Val Ser
 885 890 895
 Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu
 900 905 910
 Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu
 915 920 925
 Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr
 930 935 940
 Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr
 945 950 955 960
 Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp
 965 970 975
 Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile
 980 985 990
 Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe
 995 1000 1005
 Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe
 1010 1015 1020
 Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn
 1025 1030 1035 1040
 Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys
 1045 1050 1055
 Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu
 1060 1065 1070
 Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu
 1075 1080 1085
 Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp
 1090 1095 1100
 Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala
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 Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 1125 1130

<210> 5

<211> 3375

<212> DNA

<213> Artificial Sequence

<220>

<221> CDS

<222> (1)...(3375)

<223> Fusion between Aequorea victoria and human

<400> 5

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Met Glu Pro Pro Thr Val Pro Ser Glu Arg Ser Leu Ser Leu Ser Leu	
1 5 10 15	
ccc ggg ccc cgg gag ggc cag gcc acc ctg aag cct ccc ccg cag cac	96
Pro Gly Pro Arg Glu Gly Gln Ala Thr Leu Lys Pro Pro Pro Gln His	
20 25 30	
ctg tgg cgg cag cct cgg acc ccc atc cgt atc cag cag cgc ggc tac	144
Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr	
35 40 45	
tcc gac agc gcg gag cgc gcc gag cgg gag cgg cag ccg cac cgg ccc	192
Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro	
50 55 60	
ata gag cgc gcc gat gcc atg gac acc agc gac cgg ccc ggc ctg cgc	240
Ile Glu Arg Ala Asp Ala Met Asp Thr Ser Asp Arg Pro Gly Leu Arg	
65 70 75 80	
acg acc cgc atg tcc tgg ccc tcg tcc ttc cat ggc act ggc acc ggc	288
Thr Thr Arg Met Ser Trp Pro Ser Ser Phe His Gly Thr Gly Thr Gly	
85 90 95	
agc ggc ggc gcg ggc gga ggc agc agc agg cgc ttc gag gca gag aat	336
Ser Gly Gly Ala Gly Gly Gly Ser Ser Arg Arg Phe Glu Ala Glu Asn	
100 105 110	
ggg ccg aca cca tct cct ggc cgc agc ccc ctg gac tcg cag gcg agc	384
Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser	
115 120 125	
cca gga ctc gtg ctg cac gcc ggg gcg gcc acc agc cag cgc cgg gag	432
Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu	
130 135 140	
tcc ttc ctg tac cgc tca gac agc gac tat gac atg tca ccc aag acc	480
Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp Met Ser Pro Lys Thr	
145 150 155 160	
atg tcc cgg aac tca tcg gtc acc agc gag gcg cac gct gaa gac ctc	528
Met Ser Arg Asn Ser Ser Val Thr Ser Glu Ala His Ala Glu Asp Leu	
165 170 175	
atc gta aca cca ttt gct cag gtg ctg gcc agc ctc cgg agc gtc cgt	576
Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser Leu Arg Ser Val Arg	
180 185 190	
agc aac ttc tca ctc ctg acc aat gtg ccc gtt ccc agt aac aag cgg	624
Ser Asn Phe Ser Leu Leu Thr Asn Val Pro Val Pro Ser Asn Lys Arg	
195 200 205	

tcc ccg ctg ggc ggc ccc acc cct gtc tgc aag gcc acg ctg tca gaa	672
Ser Pro Leu Gly Gly Pro Thr Pro Val Cys Lys Ala Thr Leu Ser Glu	
210 215 220	
gaa acg tgt cag cag ttg gcc cgg gag act ctg gag gag ctg gac tgg	720
Glu Thr Cys Gln Gln Leu Ala Arg Glu Thr Leu Glu Glu Leu Asp Trp	
225 230 235 240	
tgt ctg gag cag ctg gag acc atg cag acc tat cgc tct gtc agc gag	768
Cys Leu Glu Gln Leu Glu Thr Met Gln Thr Tyr Arg Ser Val Ser Glu	
245 250 255	
atg gcc tcg cac aag ttt aaa agg atg ttg aac cgt gag ctc aca cac	816
Met Ala Ser His Lys Phe Lys Arg Met Leu Asn Arg Glu Leu Thr His	
260 265 270	
ctg tca gaa atg agc agg tcc gga aac cag gtc tca gag tac att tcc	864
Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile Ser	
275 280 285	
aca aca ttc ctg gac aaa cag aat gaa gtg gag atc cca tca ccc acg	912
Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro Thr	
290 295 300	
atg aag gaa cga gaa aaa cag caa ccg ccc ccg ccc ccg gta cca cac	960
Met Lys Glu Arg Glu Lys Gln Gln Pro Pro Pro Pro Pro Val Pro His	
305 310 315 320	
tta cag ccc atg tcc caa atc aca ggg ttg aaa aag ttg atg cat agt	1008
Leu Gln Pro Met Ser Gln Ile Thr Gly Leu Lys Lys Leu Met His Ser	
325 330 335	
aac agc ctg aac aac tct aac att ccc cga ttt ggg gtg aag acc gat	1056
Asn Ser Leu Asn Asn Ser Asn Ile Pro Arg Phe Gly Val Lys Thr Asp	
340 345 350	
caa gaa gag ctc ctg gcc caa gaa ctg gag aac ctg aac aag tgg ggc	1104
Gln Glu Glu Leu Leu Ala Gln Glu Leu Glu Asn Leu Asn Lys Trp Gly	
355 360 365	
ctg aac atc ttt tgc gtg tcg gat tac gct gga ggc cgc tca ctc acc	1152
Leu Asn Ile Phe Cys Val Ser Asp Tyr Ala Gly Gly Arg Ser Leu Thr	
370 375 380	
tgc atc atg tac atg ata ttc cag gag cgg gac ctg ctg aag aaa ttc	1200
Cys Ile Met Tyr Met Ile Phe Gln Glu Arg Asp Leu Leu Lys Lys Phe	
385 390 395 400	
cgc atc ccg gtg gac acg atg gtg aca tac atg ctg acg ctg gag gat	1248
Arg Ile Pro Val Asp Thr Met Val Thr Tyr Met Leu Thr Leu Glu Asp	
405 410 415	
cac tac cac gct gac gtg gcc tac cat aac agc ctg cac gca gct gac	1296
His Tyr His Ala Asp Val Ala Tyr His Asn Ser Leu His Ala Ala Asp	
420 425 430	
gtg ctg cag tcc acc cac gta ctg ctg gcc acg cct gca cta gat gca	1344
Val Leu Gln Ser Thr His Val Leu Leu Ala Thr Pro Ala Leu Asp Ala	
435 440 445	

gtg ttc acg gac ctg gag att ctc gcc gcc ctc ttc gcg gct gcc atc Val Phe Thr Asp Leu Glu Ile Leu Ala Ala Leu Phe Ala Ala Ala Ile 450 455 460	1392
cac gat gtg gat cac cct ggg gtc tcc aac cag ttc ctc atc aac acc His Asp Val Asp His Pro Gly Val Ser Asn Gln Phe Leu Ile Asn Thr 465 470 475 480	1440
aat tcg gag ctg gcg ctc atg tac aac gat gag tcg gtg ctc gag aat Asn Ser Glu Leu Ala Leu Met Tyr Asn Asp Glu Ser Val Leu Glu Asn 485 490 495	1488
cac cac ctg gcc gtg ggc ttc aag ctg ctg cag gag gac aac tgc gac His His Leu Ala Val Gly Phe Lys Leu Leu Gln Glu Asp Asn Cys Asp 500 505 510	1536
atc ttc cag aac ctc agc aag cgc cag cgg cag agc cta cgc aag atg Ile Phe Gln Asn Leu Ser Lys Arg Gln Arg Gln Ser Leu Arg Lys Met 515 520 525	1584
gtc atc gac atg gtg ctg gcc acg gac atg tcc aag cac atg acc ctc Val Ile Asp Met Val Leu Ala Thr Asp Met Ser Lys His Met Thr Leu 530 535 540	1632
ctg gct gac ctg aag acc atg gtg gag acc aag aaa gtg acc agc tca Leu Ala Asp Leu Lys Thr Met Val Glu Thr Lys Lys Val Thr Ser Ser 545 550 555 560	1680
ggg gtc ctc ctg cta gat aac tac tcc gac cgc atc cag gtc ctc cgg Gly Val Leu Leu Leu Asp Asn Tyr Ser Asp Arg Ile Gln Val Leu Arg 565 570 575	1728
aac atg gtg cac tgt gcc gac ctc agc aac ccc acc aag ccg ctg gag Asn Met Val His Cys Ala Asp Leu Ser Asn Pro Thr Lys Pro Leu Glu 580 585 590	1776
ctg tac cgc cag tgg aca gac cgc atc atg gcc gag ttc ttc cag cag Leu Tyr Arg Gln Trp Thr Asp Arg Ile Met Ala Glu Phe Phe Gln Gln 595 600 605	1824
ggg gac cga gag cgc gag cgt ggc atg gaa atc agc ccc atg tgt gac Gly Asp Arg Glu Arg Glu Arg Gly Met Glu Ile Ser Pro Met Cys Asp 610 615 620	1872
aag cac act gcc tcc gtg gag aag tct cag gtg ggt ttt att gac tac Lys His Thr Ala Ser Val Glu Lys Ser Gln Val Gly Phe Ile Asp Tyr 625 630 635 640	1920
att gtg cac cca ttg tgg gag acc tgg gcg gac ctt gtc cac cca gat Ile Val His Pro Leu Trp Glu Thr Trp Ala Asp Leu Val His Pro Asp 645 650 655	1968
gcc cag gag atc ttg gac act ttg gag gac aac cgg gac tgg tac tac Ala Gln Glu Ile Leu Asp Thr Leu Glu Asp Asn Arg Asp Trp Tyr Tyr 660 665 670	2016
agc gcc atc cgg cag agc cca tct ccg cca ccc gag gag gag tca agg Ser Ala Ile Arg Gln Ser Pro Ser Pro Pro Pro Glu Glu Glu Ser Arg	2064

675	680	685	
ggg cca ggc cac cca ccc ctg cct gac aag ttc cag ttt gag ctg acg Gly Pro Gly His Pro Pro Leu Pro Asp Lys Phe Gln Phe Glu Leu Thr 690 695 700			2112
ctg gag gag gaa gag gag gaa gaa ata tca atg gcc cag ata ccg tgc Leu Glu Glu Glu Glu Glu Glu Glu Ile Ser Met Ala Gln Ile Pro Cys 705 710 715 720			2160
aca gcc caa gag gca ttg act gcg cag gga ttg tca gga gtc gag gaa Thr Ala Gln Glu Ala Leu Thr Ala Gln Gly Leu Ser Gly Val Glu Glu 725 730 735			2208
gct ctg gat gca acc ata gcc tgg gag gca tcc ccg gcc cag gag tcg Ala Leu Asp Ala Thr Ile Ala Trp Glu Ala Ser Pro Ala Gln Glu Ser 740 745 750			2256
ttg gaa gtt atg gca cag gaa gca tcc ctg gag gcc gag ctg gag gca Leu Glu Val Met Ala Gln Glu Ala Ser Leu Glu Ala Glu Leu Glu Ala 755 760 765			2304
gtg tat ttg aca cag cag gca cag tcc aca ggc agt gca cct gtg gct Val Tyr Leu Thr Gln Gln Ala Gln Ser Thr Gly Ser Ala Pro Val Ala 770 775 780			2352
ccg gat gag ttc tcg tcc cgg gag gaa ttc gtg gtt gct gta agc cac Pro Asp Glu Phe Ser Ser Arg Glu Glu Phe Val Val Ala Val Ser His 785 790 795 800			2400
agc agc ccc tct gcc ctg gct ctt caa agc ccc ctt ctc cct gct tgg Ser Ser Pro Ser Ala Leu Ala Leu Gln Ser Pro Leu Leu Pro Ala Trp 805 810 815			2448
agg acc ctg tct gtt tca gag cat gcc ccg ggc ctc ccg ggc ctc ccc Arg Thr Leu Ser Val Ser Glu His Ala Pro Gly Leu Pro Gly Leu Pro 820 825 830			2496
tcc acg gcg gcc gag gtg gag gcc caa cga gag cac cag gct gcc aag Ser Thr Ala Ala Glu Val Glu Ala Gln Arg Glu His Gln Ala Ala Lys 835 840 845			2544
agg gct tgc agt gcc tgc gca ggg aca ttt ggg gag gac aca tcc gca Arg Ala Cys Ser Ala Cys Ala Gly Thr Phe Gly Glu Asp Thr Ser Ala 850 855 860			2592
ctc cca gct cct ggt ggc ggg ggg tca ggt gga gac cct acc tgg gat Leu Pro Ala Pro Gly Gly Gly Gly Ser Gly Gly Asp Pro Thr Trp Asp 865 870 875 880			2640
cca ccg gtc gcc acc atg gtg agc aag ggc gag gag ctg ttc acc ggg Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly 885 890 895			2688
gtg gtg ccc atc ctg gtc gag ctg gac ggc gac gta aac ggc cac aag Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys 900 905 910			2736
ttc agc gtg tcc ggc gag ggc gag ggc gat gcc acc tac ggc aag ctg			2784

Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu	
915 920 925	
acc ctg aag ttc atc tgc acc acc ggc aag ctg ccc gtg ccc tgg ccc	2832
Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro	
930 935 940	
acc ctc gtg acc acc ctg acc tac ggc gtg cag tgc ttc agc cgc tac	2880
Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr	
945 950 955 960	
ccc gac cac atg aag cag cac gac ttc ttc aag tcc gcc atg ccc gaa	2928
Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu	
965 970 975	
ggc tac gtc cag gag cgc acc atc ttc ttc aag gac gac ggc aac tac	2976
Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr	
980 985 990	
aag acc cgc gcc gag gtg aag ttc gag ggc gac acc ctg gtg aac cgc	3024
Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg	
995 1000 1005	
atc gag ctg aag ggc atc gac ttc aag gag gac ggc aac atc ctg ggg	3072
Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly	
1010 1015 1020	
cac aag ctg gag tac aac tac aac agc cac aac gtc tat atc atg gcc	3120
His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala	
1025 1030 1035 1040	
gac aag cag aag aac ggc atc aag gtg aac ttc aag atc cgc cac aac	3168
Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn	
1045 1050 1055	
atc gag gac ggc agc gtg cag ctc gcc gac cac tac cag cag aac acc	3216
Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr	
1060 1065 1070	
ccc atc ggc gac ggc ccc gtg ctg ctg ccc gac aac cac tac ctg agc	3264
Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser	
1075 1080 1085	
acc cag tcc gcc ctg agc aaa gac ccc aac gag aag cgc gat cac atg	3312
Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met	
1090 1095 1100	
gtc ctg ctg gag ttc gtg acc gcc gcc ggg atc act ctc ggc atg gac	3360
Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp	
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Glu Leu Tyr Lys *	

<210> 6

<211> 1124

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion between Aequorea victoria and human

<400> 6

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      20          25          30
Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr
      35          40          45
Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro
 50          55          60
Ile Glu Arg Ala Asp Ala Met Asp Thr Ser Asp Arg Pro Gly Leu Arg
 65          70          75          80
Thr Thr Arg Met Ser Trp Pro Ser Ser Phe His Gly Thr Gly Thr Gly
      85          90          95
Ser Gly Gly Ala Gly Gly Gly Ser Ser Arg Arg Phe Glu Ala Glu Asn
      100          105          110
Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser
      115          120          125
Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu
      130          135          140
Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp Met Ser Pro Lys Thr
 145          150          155          160
Met Ser Arg Asn Ser Ser Val Thr Ser Glu Ala His Ala Glu Asp Leu
      165          170          175
Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser Leu Arg Ser Val Arg
      180          185          190
Ser Asn Phe Ser Leu Leu Thr Asn Val Pro Val Pro Ser Asn Lys Arg
      195          200          205
Ser Pro Leu Gly Gly Pro Thr Pro Val Cys Lys Ala Thr Leu Ser Glu
      210          215          220
Glu Thr Cys Gln Gln Leu Ala Arg Glu Thr Leu Glu Glu Leu Asp Trp
 225          230          235          240
Cys Leu Glu Gln Leu Glu Thr Met Gln Thr Tyr Arg Ser Val Ser Glu
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Met Ala Ser His Lys Phe Lys Arg Met Leu Asn Arg Glu Leu Thr His
      260          265          270
Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile Ser
      275          280          285
Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro Thr
      290          295          300
Met Lys Glu Arg Glu Lys Gln Gln Pro Pro Pro Pro Pro Val Pro His
 305          310          315          320
Leu Gln Pro Met Ser Gln Ile Thr Gly Leu Lys Lys Leu Met His Ser
      325          330          335
Asn Ser Leu Asn Asn Ser Asn Ile Pro Arg Phe Gly Val Lys Thr Asp
      340          345          350
Gln Glu Glu Leu Leu Ala Gln Glu Leu Glu Asn Leu Asn Lys Trp Gly
      355          360          365
Leu Asn Ile Phe Cys Val Ser Asp Tyr Ala Gly Gly Arg Ser Leu Thr
      370          375          380
Cys Ile Met Tyr Met Ile Phe Gln Glu Arg Asp Leu Leu Lys Lys Phe
 385          390          395          400
Arg Ile Pro Val Asp Thr Met Val Thr Tyr Met Leu Thr Leu Glu Asp
      405          410          415
His Tyr His Ala Asp Val Ala Tyr His Asn Ser Leu His Ala Ala Asp

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420					425					430					
Val	Leu	Gln	Ser	Thr	His	Val	Leu	Leu	Ala	Thr	Pro	Ala	Leu	Asp	Ala
435					440					445					
Val	Phe	Thr	Asp	Leu	Glu	Ile	Leu	Ala	Ala	Leu	Phe	Ala	Ala	Ala	Ile
450					455					460					
His	Asp	Val	Asp	His	Pro	Gly	Val	Ser	Asn	Gln	Phe	Leu	Ile	Asn	Thr
465					470					475					
Asn	Ser	Glu	Leu	Ala	Leu	Met	Tyr	Asn	Asp	Glu	Ser	Val	Leu	Glu	Asn
485					490					495					
His	His	Leu	Ala	Val	Gly	Phe	Lys	Leu	Leu	Gln	Glu	Asp	Asn	Cys	Asp
500					505					510					
Ile	Phe	Gln	Asn	Leu	Ser	Lys	Arg	Gln	Arg	Gln	Ser	Leu	Arg	Lys	Met
515					520					525					
Val	Ile	Asp	Met	Val	Leu	Ala	Thr	Asp	Met	Ser	Lys	His	Met	Thr	Leu
530					535					540					
Leu	Ala	Asp	Leu	Lys	Thr	Met	Val	Glu	Thr	Lys	Lys	Val	Thr	Ser	Ser
545					550					555					
Gly	Val	Leu	Leu	Leu	Asp	Asn	Tyr	Ser	Asp	Arg	Ile	Gln	Val	Leu	Arg
565					570					575					
Asn	Met	Val	His	Cys	Ala	Asp	Leu	Ser	Asn	Pro	Thr	Lys	Pro	Leu	Glu
580					585					590					
Leu	Tyr	Arg	Gln	Trp	Thr	Asp	Arg	Ile	Met	Ala	Glu	Phe	Phe	Gln	Gln
595					600					605					
Gly	Asp	Arg	Glu	Arg	Glu	Arg	Gly	Met	Glu	Ile	Ser	Pro	Met	Cys	Asp
610					615					620					
Lys	His	Thr	Ala	Ser	Val	Glu	Lys	Ser	Gln	Val	Gly	Phe	Ile	Asp	Tyr
625					630					635					
Ile	Val	His	Pro	Leu	Trp	Glu	Thr	Trp	Ala	Asp	Leu	Val	His	Pro	Asp
645					650					655					
Ala	Gln	Glu	Ile	Leu	Asp	Thr	Leu	Glu	Asp	Asn	Arg	Asp	Trp	Tyr	Tyr
660					665					670					
Ser	Ala	Ile	Arg	Gln	Ser	Pro	Ser	Pro	Pro	Pro	Glu	Glu	Glu	Ser	Arg
675					680					685					
Gly	Pro	Gly	His	Pro	Pro	Leu	Pro	Asp	Lys	Phe	Gln	Phe	Glu	Leu	Thr
690					695					700					
Leu	Glu	Glu	Glu	Glu	Glu	Glu	Glu	Ile	Ser	Met	Ala	Gln	Ile	Pro	Cys
705					710					715					
Thr	Ala	Gln	Glu	Ala	Leu	Thr	Ala	Gln	Gly	Leu	Ser	Gly	Val	Glu	Glu
725					730					735					
Ala	Leu	Asp	Ala	Thr	Ile	Ala	Trp	Glu	Ala	Ser	Pro	Ala	Gln	Glu	Ser
740					745					750					
Leu	Glu	Val	Met	Ala	Gln	Glu	Ala	Ser	Leu	Glu	Ala	Glu	Leu	Glu	Ala
755					760					765					
Val	Tyr	Leu	Thr	Gln	Gln	Ala	Gln	Ser	Thr	Gly	Ser	Ala	Pro	Val	Ala
770					775					780					
Pro	Asp	Glu	Phe	Ser	Ser	Arg	Glu	Glu	Phe	Val	Val	Ala	Val	Ser	His
785					790					795					
Ser	Ser	Pro	Ser	Ala	Leu	Ala	Leu	Gln	Ser	Pro	Leu	Leu	Pro	Ala	Trp
805					810					815					
Arg	Thr	Leu	Ser	Val	Ser	Glu	His	Ala	Pro	Gly	Leu	Pro	Gly	Leu	Pro
820					825					830					
Ser	Thr	Ala	Ala	Glu	Val	Glu	Ala	Gln	Arg	Glu	His	Gln	Ala	Ala	Lys
835					840					845					
Arg	Ala	Cys	Ser	Ala	Cys	Ala	Gly	Thr	Phe	Gly	Glu	Asp	Thr	Ser	Ala
850					855					860					
Leu	Pro	Ala	Pro	Gly	Gly	Gly	Gly	Ser	Gly	Gly	Asp	Pro	Thr	Trp	Asp
865					870					875					
Pro	Pro	Val	Ala	Thr	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly
885					890					895					

Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys
 900 905 910
 Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu
 915 920 925
 Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro
 930 935 940
 Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr
 945 950 955 960
 Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu
 965 970 975
 Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr
 980 985 990
 Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg
 995 1000 1005
 Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly
 1010 1015 1020
 His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala
 1025 1030 1035 1040
 Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn
 1045 1050 1055
 Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr
 1060 1065 1070
 Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser
 1075 1080 1085
 Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met
 1090 1095 1100
 Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp
 1105 1110 1115 1120
 Glu Leu Tyr Lys

<210> 7

<211> 3399

<212> DNA

<213> Artificial Sequence

<220>

<221> CDS

<222> (1)...(3399)

<223> Fusion between Aequorea victoria and human

<400> 7

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Met Glu Pro Pro Thr Val Pro Ser Glu Arg Ser Leu Ser Leu Ser Leu	
1 5 10 15	
ccc ggg ccc cgg gag ggc cag gcc acc ctg aag cct ccc ccg cag cac	96
Pro Gly Pro Arg Glu Gly Gln Ala Thr Leu Lys Pro Pro Pro Gln His	
20 25 30	
ctg tgg cgg cag cct cgg acc ccc atc cgt atc cag cag cgc ggc tac	144
Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr	
35 40 45	
tcc gac agc gcg gag cgc gcc gag cgg gag cgg cag ccg cac cgg ccc	192
Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro	
50 55 60	
ata gag cgc gcc gat gcc atg gac acc agc gac cgg ccc ggc ctg cgc	240

23

Ile	Glu	Arg	Ala	Asp	Ala	Met	Asp	Thr	Ser	Asp	Arg	Pro	Gly	Leu	Arg	
65					70					75					80	
acg	acc	cgc	atg	tcc	tgg	ccc	tcg	tcc	ttc	cat	ggc	act	ggc	acc	ggc	288
Thr	Thr	Arg	Met	Ser	Trp	Pro	Ser	Ser	Phe	His	Gly	Thr	Gly	Thr	Gly	
				85					90					95		
agc	ggc	ggc	gcg	ggc	gga	ggc	agc	agc	agg	cgc	ttc	gag	gca	gag	aat	336
Ser	Gly	Gly	Ala	Gly	Gly	Gly	Ser	Ser	Arg	Arg	Phe	Glu	Ala	Glu	Asn	
			100					105					110			
ggg	ccg	aca	cca	tct	cct	ggc	cgc	agc	ccc	ctg	gac	tcg	cag	gcg	agc	384
Gly	Pro	Thr	Pro	Ser	Pro	Gly	Arg	Ser	Pro	Leu	Asp	Ser	Gln	Ala	Ser	
		115					120					125				
cca	gga	ctc	gtg	ctg	cac	gcc	ggg	gcg	gcc	acc	agc	cag	cgc	cgg	gag	432
Pro	Gly	Leu	Val	Leu	His	Ala	Gly	Ala	Ala	Thr	Ser	Gln	Arg	Arg	Glu	
		130				135				140						
tcc	ttc	ctg	tac	cgc	tca	gac	agc	gac	tat	gac	atg	tca	ccc	aag	acc	480
Ser	Phe	Leu	Tyr	Arg	Ser	Asp	Ser	Asp	Tyr	Asp	Met	Ser	Pro	Lys	Thr	
145					150				155					160		
atg	tcc	cgg	aac	tca	tcg	gtc	acc	agc	gag	gcg	cac	gct	gaa	gac	ctc	528
Met	Ser	Arg	Asn	Ser	Ser	Val	Thr	Ser	Glu	Ala	His	Ala	Glu	Asp	Leu	
				165				170					175			
atc	gta	aca	cca	ttt	gct	cag	gtg	ctg	gcc	agc	ctc	cgg	agc	gtc	cgt	576
Ile	Val	Thr	Pro	Phe	Ala	Gln	Val	Leu	Ala	Ser	Leu	Arg	Ser	Val	Arg	
			180					185					190			
agc	aac	ttc	tca	ctc	ctg	acc	aat	gtg	ccc	gtt	ccc	agt	aac	aag	cgg	624
Ser	Asn	Phe	Ser	Leu	Leu	Thr	Asn	Val	Pro	Val	Pro	Ser	Asn	Lys	Arg	
		195					200				205					
tcc	ccg	ctg	ggc	ggc	ccc	acc	cct	gtc	tgc	aag	gcc	acg	ctg	tca	gaa	672
Ser	Pro	Leu	Gly	Gly	Pro	Thr	Pro	Val	Cys	Lys	Ala	Thr	Leu	Ser	Glu	
		210				215					220					
gaa	acg	tgt	cag	cag	ttg	gcc	cgg	gag	act	ctg	gag	gag	ctg	gac	tgg	720
Glu	Thr	Cys	Gln	Gln	Leu	Ala	Arg	Glu	Thr	Leu	Glu	Glu	Leu	Asp	Trp	
225					230				235					240		
tgt	ctg	gag	cag	ctg	gag	acc	atg	cag	acc	tat	cgc	tct	gtc	agc	gag	768
Cys	Leu	Glu	Gln	Leu	Glu	Thr	Met	Gln	Thr	Tyr	Arg	Ser	Val	Ser	Glu	
				245				250					255			
atg	gcc	tcg	cac	aag	ttt	aaa	agg	atg	ttg	aac	cgt	gag	ctc	aca	cac	816
Met	Ala	Ser	His	Lys	Phe	Lys	Arg	Met	Leu	Asn	Arg	Glu	Leu	Thr	His	
			260					265				270				
ctg	tca	gaa	atg	agc	agg	tcc	gga	aac	cag	gtc	tca	gag	tac	att	tcc	864
Leu	Ser	Glu	Met	Ser	Arg	Ser	Gly	Asn	Gln	Val	Ser	Glu	Tyr	Ile	Ser	
		275					280					285				
aca	aca	ttc	ctg	gac	aaa	cag	aat	gaa	gtg	gag	atc	cca	tca	ccc	acg	912
Thr	Thr	Phe	Leu	Asp	Lys	Gln	Asn	Glu	Val	Glu	Ile	Pro	Ser	Pro	Thr	
		290				295					300					

atg aag gaa cga gaa aaa cag caa gcg ccg cga cca aga ccc tcc cag Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser Gln 305 310 315 320	960
ccg ccc ccg ccc cct gta cca cac tta cag ccc atg tcc caa atc aca Pro Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile Thr 325 330 335	1008
ggg ttg aaa aag ttg atg cat agt aac agc ctg aac aac tct aac att Gly Leu Lys Lys Leu Met His Ser Asn Ser Leu Asn Asn Ser Asn Ile 340 345 350	1056
ccc cga ttt ggg gtg aag acc gat caa gaa gag ctc ctg gcc caa gaa Pro Arg Phe Gly Val Lys Thr Asp Gln Glu Glu Leu Leu Ala Gln Glu 355 360 365	1104
ctg gag aac ctg aac aag tgg ggc ctg aac atc ttt tgc gtg tcg gat Leu Glu Asn Leu Asn Lys Trp Gly Leu Asn Ile Phe Cys Val Ser Asp 370 375 380	1152
tac gct gga ggc cgc tca ctc acc tgc atc atg tac atg ata ttc cag Tyr Ala Gly Gly Arg Ser Leu Thr Cys Ile Met Tyr Met Ile Phe Gln 385 390 395 400	1200
gag cgg gac ctg ctg aag aaa ttc cgc atc ccg gtg gac acg atg gtg Glu Arg Asp Leu Leu Lys Lys Phe Arg Ile Pro Val Asp Thr Met Val 405 410 415	1248
aca tac atg ctg acg ctg gag gat cac tac cac gct gac gtg gcc tac Thr Tyr Met Leu Thr Leu Glu Asp His Tyr His Ala Asp Val Ala Tyr 420 425 430	1296
cat aac agc ctg cac gca gct gac gtg ctg cag tcc acc cac gta ctg His Asn Ser Leu His Ala Ala Asp Val Leu Gln Ser Thr His Val Leu 435 440 445	1344
ctg gcc acg cct gca cta gat gca gtg ttc acg gac ctg gag att ctc Leu Ala Thr Pro Ala Leu Asp Ala Val Phe Thr Asp Leu Glu Ile Leu 450 455 460	1392
gcc gcc ctc ttc gcg gct gcc atc cac gat gtg gat cac cct ggg gtc Ala Ala Leu Phe Ala Ala Ala Ile His Asp Val Asp His Pro Gly Val 465 470 475 480	1440
tcc aac cag ttc ctc atc aac acc aat tcg gag ctg gcg ctc atg tac Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met Tyr 485 490 495	1488
aac gat gag tcg gtg ctc gaa aat cac aac ctg gcc gtg ggc ttc aag Asn Asp Glu Ser Val Leu Glu Asn His Asn Leu Ala Val Gly Phe Lys 500 505 510	1536
ctg ctg cag gag gac aac tgc gac atc ttc cag aac ctc agc aag cgc Leu Leu Gln Glu Asp Asn Cys Asp Ile Phe Gln Asn Leu Ser Lys Arg 515 520 525	1584
cag cgg cag agc cta cgc aag atg gtc atc gac atg gtg ctg gcc acg Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp Met Val Leu Ala Thr 530 535 540	1632

gac atg tcc aag cac atg acc ctc ctg gct gac ctg aag acc atg gtg Asp Met Ser Lys His Met Thr Leu Leu Ala Asp Leu Lys Thr Met Val 545 550 555 560	1680
gag acc aag aaa gtg acc agc tca ggg gtc ctc ctg cta gat aac tac Glu Thr Lys Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn Tyr 565 570 575	1728
tcc gac cgc atc cag gtc ctc cgg aac atg gtg cac tgt gcc gac ctc Ser Asp Arg Ile Gln Val Leu Arg Asn Met Val His Cys Ala Asp Leu 580 585 590	1776
agc aac ccc acc aag ccg ctg gag ctg tac cgc cag tgg aca gac cgc Ser Asn Pro Thr Lys Pro Leu Glu Leu Tyr Arg Gln Trp Thr Asp Arg 595 600 605	1824
atc atg gcc gag ttc ttc cag cag ggt gac cga gag cgc gag cgt ggc Ile Met Ala Glu Phe Phe Gln Gln Gly Asp Arg Glu Arg Glu Arg Gly 610 615 620	1872
atg gaa atc agc ccc atg tgt gac aag cac act gcc tcc gtg gag aag Met Glu Ile Ser Pro Met Cys Asp Lys His Thr Ala Ser Val Glu Lys 625 630 635 640	1920
tct cag gtg ggt ttt att gac tac att gtg cac cca ttg tgg gag acc Ser Gln Val Gly Phe Ile Asp Tyr Ile Val His Pro Leu Trp Glu Thr 645 650 655	1968
tgg gcg gac ctt gtc cac cca gat gcc cag gag atc ttg gac act ttg Trp Ala Asp Leu Val His Pro Asp Ala Gln Glu Ile Leu Asp Thr Leu 660 665 670	2016
gag gac aac cgg gac tgg tac tac agc gcc atc cgg cag agc cca tct Glu Asp Asn Arg Asp Trp Tyr Tyr Ser Ala Ile Arg Gln Ser Pro Ser 675 680 685	2064
ccg cca ccc gag gag gag tca agg ggg cca ggc cac cca ccc ctg cct Pro Pro Pro Glu Glu Glu Ser Arg Gly Pro Gly His Pro Pro Leu Pro 690 695 700	2112
gac aag ttc cag ttt gag ctg acg ctg gag gag gaa gag gag gaa gaa Asp Lys Phe Gln Phe Glu Leu Thr Leu Glu Glu Glu Glu Glu Glu 705 710 715 720	2160
ata tca atg gcc cag ata ccg tgc aca gcc caa gag gca ttg act gcg Ile Ser Met Ala Gln Ile Pro Cys Thr Ala Gln Glu Ala Leu Thr Ala 725 730 735	2208
cag gga ttg tca gga gtc gag gaa gct ctg gat gca acc ata gcc tgg Gln Gly Leu Ser Gly Val Glu Glu Ala Leu Asp Ala Thr Ile Ala Trp 740 745 750	2256
gag gca tcc ccg gcc cag gag tcg ttg gaa gtt atg gca cag gaa gca Glu Ala Ser Pro Ala Gln Glu Ser Leu Glu Val Met Ala Gln Glu Ala 755 760 765	2304
tcc ctg gag gcc gag ctg gag gca gtg tat ttg aca cag cag gca cag Ser Leu Glu Ala Glu Leu Glu Ala Val Tyr Leu Thr Gln Gln Ala Gln	2352

770	775	780	
tcc aca ggc agt gca cct gtg gct ccg gat gag ttc tcg tcc cgg gag Ser Thr Gly Ser Ala Pro Val Ala Pro Asp Glu Phe Ser Ser Arg Glu 785 790 795 800			2400
gaa ttc gtg gtt gct gta agc cac agc agc ccc tct gcc ctg gct ctt Glu Phe Val Val Ala Val Ser His Ser Ser Pro Ser Ala Leu Ala Leu 805 810 815			2448
caa agc ccc ctt ctc cct gct tgg agg acc ctg tct gtt tca gag cat Gln Ser Pro Leu Leu Pro Ala Trp Arg Thr Leu Ser Val Ser Glu His 820 825 830			2496
gcc ccg ggc ctc ccg ggc ctc ccc tcc acg gcg gcc gag gtg gag gcc Ala Pro Gly Leu Pro Gly Leu Pro Ser Thr Ala Ala Glu Val Glu Ala 835 840 845			2544
caa cga gag cac cag gct gcc aag agg gct tgc agt gcc tgc gca ggg Gln Arg Glu His Gln Ala Ala Lys Arg Ala Cys Ser Ala Cys Ala Gly 850 855 860			2592
aca ttt ggg gag gac aca tcc gca ctc cca gct cct ggt ggc ggg ggg Thr Phe Gly Glu Asp Thr Ser Ala Leu Pro Ala Pro Gly Gly Gly Gly 865 870 875 880			2640
tca ggt gga gac cct acc tgg gat cca ccg gtc gcc acc atg gtg agc Ser Gly Gly Asp Pro Thr Trp Asp Pro Pro Val Ala Thr Met Val Ser 885 890 895			2688
aag ggc gag gag ctg ttc acc ggg gtg gtg ccc atc ctg gtc gag ctg Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu 900 905 910			2736
gac ggc gac gta aac ggc cac aag ttc agc gtg tcc ggc gag ggc gag Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu 915 920 925			2784
ggc gat gcc acc tac ggc aag ctg acc ctg aag ttc atc tgc acc acc Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr 930 935 940			2832
ggc aag ctg ccc gtg ccc tgg ccc acc ctc gtg acc acc ctg acc tac Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Thr Tyr 945 950 955 960			2880
ggc gtg cag tgc ttc agc cgc tac ccc gac cac atg aag cag cac gac Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp 965 970 975			2928
ttc ttc aag tcc gcc atg ccc gaa ggc tac gtc cag gag cgc acc atc Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile 980 985 990			2976
ttc ttc aag gac gac ggc aac tac aag acc cgc gcc gag gtg aag ttc Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe 995 1000 1005			3024
gag ggc gac acc ctg gtg aac cgc atc gag ctg aag ggc atc gac ttc			3072

Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe
 1010 1015 1020
 aag gag gac ggc aac atc ctg ggg cac aag ctg gag tac aac tac aac 3120
 Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn
 1025 1030 1035 1040
 agc cac aac gtc tat atc atg gcc gac aag cag aag aac ggc atc aag 3168
 Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys
 1045 1050 1055
 gtg aac ttc aag atc cgc cac aac atc gag gac ggc agc gtg cag ctc 3216
 Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu
 1060 1065 1070
 gcc gac cac tac cag cag aac acc ccc atc ggc gac ggc ccc gtg ctg 3264
 Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu
 1075 1080 1085
 ctg ccc gac aac cac tac ctg agc acc cag tcc gcc ctg agc aaa gac 3312
 Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp
 1090 1095 1100
 ccc aac gag aag cgc gat cac atg gtc ctg ctg gag ttc gtg acc gcc 3360
 Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala
 1105 1110 1115 1120
 gcc ggc atc act ctc ggc atg gac gag ctg tac aag taa 3399
 Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys *
 1125 1130

<210> 8

<211> 1132

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion between Aequorea victoria and human

<400> 8

Met Glu Pro Pro Thr Val Pro Ser Glu Arg Ser Leu Ser Leu Ser Leu
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 Pro Gly Pro Arg Glu Gly Gln Ala Thr Leu Lys Pro Pro Pro Gln His
 20 25 30
 Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr
 35 40 45
 Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro
 50 55 60
 Ile Glu Arg Ala Asp Ala Met Asp Thr Ser Asp Arg Pro Gly Leu Arg
 65 70 75 80
 Thr Thr Arg Met Ser Trp Pro Ser Ser Phe His Gly Thr Gly Thr Gly
 85 90 95
 Ser Gly Gly Ala Gly Gly Gly Ser Ser Arg Arg Phe Glu Ala Glu Asn
 100 105 110
 Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser
 115 120 125
 Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu
 130 135 140

Ser	Phe	Leu	Tyr	Arg	Ser	Asp	Ser	Asp	Tyr	Asp	Met	Ser	Pro	Lys	Thr
145					150					155					160
Met	Ser	Arg	Asn	Ser	Ser	Val	Thr	Ser	Glu	Ala	His	Ala	Glu	Asp	Leu
			165						170						175
Ile	Val	Thr	Pro	Phe	Ala	Gln	Val	Leu	Ala	Ser	Leu	Arg	Ser	Val	Arg
			180					185						190	
Ser	Asn	Phe	Ser	Leu	Leu	Thr	Asn	Val	Pro	Val	Pro	Ser	Asn	Lys	Arg
		195					200					205			
Ser	Pro	Leu	Gly	Gly	Pro	Thr	Pro	Val	Cys	Lys	Ala	Thr	Leu	Ser	Glu
	210					215					220				
Glu	Thr	Cys	Gln	Gln	Leu	Ala	Arg	Glu	Thr	Leu	Glu	Glu	Leu	Asp	Trp
225					230					235					240
Cys	Leu	Glu	Gln	Leu	Glu	Thr	Met	Gln	Thr	Tyr	Arg	Ser	Val	Ser	Glu
			245						250						255
Met	Ala	Ser	His	Lys	Phe	Lys	Arg	Met	Leu	Asn	Arg	Glu	Leu	Thr	His
			260					265					270		
Leu	Ser	Glu	Met	Ser	Arg	Ser	Gly	Asn	Gln	Val	Ser	Glu	Tyr	Ile	Ser
		275					280					285			
Thr	Thr	Phe	Leu	Asp	Lys	Gln	Asn	Glu	Val	Glu	Ile	Pro	Ser	Pro	Thr
	290					295					300				
Met	Lys	Glu	Arg	Glu	Lys	Gln	Gln	Ala	Pro	Arg	Pro	Arg	Pro	Ser	Gln
305					310					315					320
Pro	Pro	Pro	Pro	Pro	Val	Pro	His	Leu	Gln	Pro	Met	Ser	Gln	Ile	Thr
				325					330						335
Gly	Leu	Lys	Lys	Leu	Met	His	Ser	Asn	Ser	Leu	Asn	Asn	Ser	Asn	Ile
		340						345					350		
Pro	Arg	Phe	Gly	Val	Lys	Thr	Asp	Gln	Glu	Glu	Leu	Leu	Ala	Gln	Glu
		355					360					365			
Leu	Glu	Asn	Leu	Asn	Lys	Trp	Gly	Leu	Asn	Ile	Phe	Cys	Val	Ser	Asp
	370				375					380					
Tyr	Ala	Gly	Gly	Arg	Ser	Leu	Thr	Cys	Ile	Met	Tyr	Met	Ile	Phe	Gln
385					390					395					400
Glu	Arg	Asp	Leu	Leu	Lys	Lys	Phe	Arg	Ile	Pro	Val	Asp	Thr	Met	Val
			405						410						415
Thr	Tyr	Met	Leu	Thr	Leu	Glu	Asp	His	Tyr	His	Ala	Asp	Val	Ala	Tyr
		420						425					430		
His	Asn	Ser	Leu	His	Ala	Ala	Asp	Val	Leu	Gln	Ser	Thr	His	Val	Leu
		435					440						445		
Leu	Ala	Thr	Pro	Ala	Leu	Asp	Ala	Val	Phe	Thr	Asp	Leu	Glu	Ile	Leu
	450					455					460				
Ala	Ala	Leu	Phe	Ala	Ala	Ala	Ile	His	Asp	Val	Asp	His	Pro	Gly	Val
465					470					475					480
Ser	Asn	Gln	Phe	Leu	Ile	Asn	Thr	Asn	Ser	Glu	Leu	Ala	Leu	Met	Tyr
			485						490					495	
Asn	Asp	Glu	Ser	Val	Leu	Glu	Asn	His	Asn	Leu	Ala	Val	Gly	Phe	Lys
		500						505					510		
Leu	Leu	Gln	Glu	Asp	Asn	Cys	Asp	Ile	Phe	Gln	Asn	Leu	Ser	Lys	Arg
		515					520					525			
Gln	Arg	Gln	Ser	Leu	Arg	Lys	Met	Val	Ile	Asp	Met	Val	Leu	Ala	Thr
	530					535					540				
Asp	Met	Ser	Lys	His	Met	Thr	Leu	Leu	Ala	Asp	Leu	Lys	Thr	Met	Val
545					550					555					560
Glu	Thr	Lys	Lys	Val	Thr	Ser	Ser	Gly	Val	Leu	Leu	Leu	Asp	Asn	Tyr
			565					570						575	
Ser	Asp	Arg	Ile	Gln	Val	Leu	Arg	Asn	Met	Val	His	Cys	Ala	Asp	Leu
			580					585					590		
Ser	Asn	Pro	Thr	Lys	Pro	Leu	Glu	Leu	Tyr	Arg	Gln	Trp	Thr	Asp	Arg
		595					600					605			
Ile	Met	Ala	Glu	Phe	Phe	Gln	Gln	Gly	Asp	Arg	Glu	Arg	Glu	Arg	Gly

610	615	620
Met Glu Ile Ser Pro	Met Cys Asp Lys His Thr Ala	Ser Val Glu Lys
625	630	635
Ser Gln Val Gly Phe	Ile Asp Tyr Ile Val His Pro	Leu Trp Glu Thr
645	650	655
Trp Ala Asp Leu Val	His Pro Asp Ala Gln Glu Ile	Leu Asp Thr Leu
660	665	670
Glu Asp Asn Arg Asp	Trp Tyr Tyr Ser Ala Ile Arg	Gln Ser Pro Ser
675	680	685
Pro Pro Pro Glu Glu	Glu Ser Arg Gly Pro Gly His	Pro Pro Leu Pro
690	695	700
Asp Lys Phe Gln Phe	Glu Leu Thr Leu Glu Glu Glu Glu	Glu Glu Glu
705	710	715
Ile Ser Met Ala Gln	Ile Pro Cys Thr Ala Gln Glu	Ala Leu Thr Ala
725	730	735
Gln Gly Leu Ser Gly	Val Glu Glu Ala Leu Asp	Ala Thr Ile Ala Trp
740	745	750
Glu Ala Ser Pro Ala	Gln Glu Ser Leu Glu Val Met	Ala Gln Glu Ala
755	760	765
Ser Leu Glu Ala Glu	Leu Glu Ala Val Tyr Leu	Thr Gln Gln Ala Gln
770	775	780
Ser Thr Gly Ser Ala	Pro Val Ala Pro Asp Glu Phe	Ser Ser Arg Glu
785	790	795
Glu Phe Val Val Ala	Val Ser His Ser Ser Pro	Ser Ala Leu Ala Leu
805	810	815
Gln Ser Pro Leu Leu	Pro Ala Trp Arg Thr Leu	Ser Val Ser Glu His
820	825	830
Ala Pro Gly Leu Pro	Gly Leu Pro Ser Thr Ala	Ala Glu Val Glu Ala
835	840	845
Gln Arg Glu His Gln	Ala Ala Lys Arg Ala Cys	Ser Ala Cys Ala Gly
850	855	860
Thr Phe Gly Glu Asp	Thr Ser Ala Leu Pro	Ala Pro Gly Gly Gly Gly
865	870	875
Ser Gly Gly Asp Pro	Thr Trp Asp Pro Pro	Val Ala Thr Met Val Ser
885	890	895
Lys Gly Glu Glu Leu	Phe Thr Gly Val Val	Pro Ile Leu Val Glu Leu
900	905	910
Asp Gly Asp Val Asn	Gly His Lys Phe Ser	Val Ser Gly Glu Gly Glu
915	920	925
Gly Asp Ala Thr Tyr	Gly Lys Leu Thr Leu	Lys Phe Ile Cys Thr Thr
930	935	940
Gly Lys Leu Pro Val	Pro Trp Pro Thr Leu	Val Thr Thr Leu Thr Tyr
945	950	955
Gly Val Gln Cys Phe	Ser Arg Tyr Pro Asp	His Met Lys Gln His Asp
965	970	975
Phe Phe Lys Ser Ala	Met Pro Glu Gly Tyr	Val Gln Glu Arg Thr Ile
980	985	990
Phe Phe Lys Asp Asp	Gly Asn Tyr Lys Thr	Arg Ala Glu Val Lys Phe
995	1000	1005
Glu Gly Asp Thr Leu	Val Asn Arg Ile Glu	Leu Lys Gly Ile Asp Phe
1010	1015	1020
Lys Glu Asp Gly Asn	Ile Leu Gly His Lys	Leu Glu Tyr Asn Tyr Asn
1025	1030	1035
Ser His Asn Val Tyr	Ile Met Ala Asp	Lys Gln Lys Asn Gly Ile Lys
1045	1050	1055
Val Asn Phe Lys Ile	Arg His Asn Ile	Glu Asp Gly Ser Val Gln Leu
1060	1065	1070
Ala Asp His Tyr Gln	Gln Asn Thr Pro	Ile Gly Asp Gly Pro Val Leu
1075	1080	1085

Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp
 1090 1095 1100
 Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Glu Phe Val Thr Ala
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 Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 1125 1130

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<210> 13
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<210> 17
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<212> DNA
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cttacagccc 70

<210> 18
<211> 70
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 <210> 20
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 <400> 20
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 1 5 10 15

ccc ggg ccc cgg gag ggc cag gcc acc ctg aag cct ccc ccg cag cac Pro Gly Pro Arg Glu Gly Gln Ala Thr Leu Lys Pro Pro Pro Gln His 20 25 30	96
ctg tgg cgg cag cct cgg acc ccc atc cgt atc cag cag cgc ggc tac Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr 35 40 45	144
tcc gac agc gcg gag cgc gcc gag cgg gag cgg cag ccg cac cgg ccc Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro 50 55 60	192
ata gag cgc gcc gat gcc atg gac acc agc gac cgg ccc ggc ctg cgc Ile Glu Arg Ala Asp Ala Met Asp Thr Ser Asp Arg Pro Gly Leu Arg 65 70 75 80	240
acg acc cgc atg tcc tgg ccc tcg tcc ttc cat ggc act ggc acc ggc Thr Thr Arg Met Ser Trp Pro Ser Ser Phe His Gly Thr Gly Thr Gly 85 90 95	288
agc ggc ggc gcg ggc gga ggc agc agc agg cgc ttc gag gca gag aat Ser Gly Gly Ala Gly Gly Ser Ser Arg Arg Phe Glu Ala Glu Asn 100 105 110	336
ggg ccg aca cca tct cct ggc cgc agc ccc ctg gac tcg cag gcg agc Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser 115 120 125	384
cca gga ctc gtg ctg cac gcc ggg gcg gcc acc agc cag cgc cgg gag Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu 130 135 140	432
tcc ttc ctg tac cgc tca gac agc gac tat gac atg tca ccc aag acc Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp Met Ser Pro Lys Thr 145 150 155 160	480
atg tcc cgg aac tca tcg gtc acc agc gag gcg cac gct gaa gac ctc Met Ser Arg Asn Ser Ser Val Thr Ser Glu Ala His Ala Glu Asp Leu 165 170 175	528
atc gta aca cca ttt gct cag gtg ctg gcc agc ctc cgg agc gtc cgt Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser Leu Arg Ser Val Arg 180 185 190	576
agc aac ttc tca ctc ctg acc aat gtg ccc gtt ccc agt aac aag cgg Ser Asn Phe Ser Leu Leu Thr Asn Val Pro Val Pro Ser Asn Lys Arg 195 200 205	624
tcc ccg ctg ggc ggc ccc acc cct gtc tgc aag gcc acg ctg tca gaa Ser Pro Leu Gly Gly Pro Thr Pro Val Cys Lys Ala Thr Leu Ser Glu 210 215 220	672
gaa acg tgt cag cag ttg gcc cgg gag act ctg gag gag ctg gac tgg Glu Thr Cys Gln Gln Leu Ala Arg Glu Thr Leu Glu Leu Asp Trp 225 230 235 240	720
tgt ctg gag cag ctg gag acc atg cag acc tat cgc tct gtc agc gag Cys Leu Glu Gln Leu Glu Thr Met Gln Thr Tyr Arg Ser Val Ser Glu 245 250 255	768

atg gcc tcg cac aag ttc aaa agg atg ttg aac cgt gag ctc aca cac	816
Met Ala Ser His Lys Phe Lys Arg Met Leu Asn Arg Glu Leu Thr His	
260 265 270	
ctg tca gaa atg agc agg tcc gga aac cag gtc tca gag tac att tcc	864
Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile Ser	
275 280 285	
aca aca ttc ctg gac aaa cag aat gaa gtg gag atc cca tca ccc acg	912
Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro Thr	
290 295 300	
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Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser Gln	
305 310 315 320	
ccg ccc ccg ccc cct gta cca cac tta cag ccc atg tcc caa atc aca	1008
Pro Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile Thr	
325 330 335	
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Gly Leu Lys Lys Leu Met His Ser Ser Ser Leu Thr Asn Ser Ser Ile	
340 345 350	
cca agg ttt gga gtt aaa act gaa caa gaa gat gtc ctt gcc aag gaa	1104
Pro Arg Phe Gly Val Lys Thr Glu Gln Glu Asp Val Leu Ala Lys Glu	
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Leu Glu Asp Val Asn Lys Trp Gly Leu His Val Phe Arg Ile Ala Glu	
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Leu Ser Gly Asn Arg Pro Leu Thr Val Ile Met His Thr Ile Phe Gln	
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Glu Arg Asp Leu Leu Lys Thr Phe Lys Ile Pro Val Asp Thr Leu Ile	
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aca tat ctt atg act ctc gaa gac cat tac cat gct gat gtg gcc tat	1296
Thr Tyr Leu Met Thr Leu Glu Asp His Tyr His Ala Asp Val Ala Tyr	
420 425 430	
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His Asn Asn Ile His Ala Ala Asp Val Val Gln Ser Thr His Val Leu	
435 440 445	
tta tct aca cct gct ttg gag gct gtg ttt aca gat ttg gag att ctt	1392
Leu Ser Thr Pro Ala Leu Glu Ala Val Phe Thr Asp Leu Glu Ile Leu	
450 455 460	
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Ala Ala Ile Phe Ala Ser Ala Ile His Asp Val Asp His Pro Gly Val	
465 470 475 480	
tcc aat caa ttt ctg atc aat aca aac tct gaa ctt gcc ttg atg tac	1488
Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met Tyr	

485								490					495					
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Asn	Asp	Ser	Ser	Val	Leu	Glu	Asn	His	His	Leu	Ala	Val	Gly	Phe	Lys			
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ttg	ctt	cag	gaa	gaa	aac	tgt	gac	att	ttc	cag	aat	ttg	acc	aaa	aaa	1584		
Leu	Leu	Gln	Glu	Glu	Asn	Cys	Asp	Ile	Phe	Gln	Asn	Leu	Thr	Lys	Lys			
515								520					525					
caa	aga	caa	tct	tta	agg	aaa	atg	gtc	att	gac	atc	gta	ctt	gca	aca	1632		
Gln	Arg	Gln	Ser	Leu	Arg	Lys	Met	Val	Ile	Asp	Ile	Val	Leu	Ala	Thr			
530								535					540					
gat	atg	tca	aaa	cac	atg	aat	cta	ctg	gct	gat	ttg	aag	act	atg	gtt	1680		
Asp	Met	Ser	Lys	His	Met	Asn	Leu	Leu	Ala	Asp	Leu	Lys	Thr	Met	Val			
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Glu	Thr	Lys	Lys	Val	Thr	Ser	Ser	Gly	Val	Leu	Leu	Leu	Asp	Asn	Tyr			
565								570					575					
tcc	gat	agg	att	cag	gtt	ctt	cag	aat	atg	gtg	cac	tgt	gca	gat	ctg	1776		
Ser	Asp	Arg	Ile	Gln	Val	Leu	Gln	Asn	Met	Val	His	Cys	Ala	Asp	Leu			
580								585					590					
agc	aac	cca	aca	aag	cct	ctc	cag	ctg	tac	cgc	cag	tgg	acg	gac	cgg	1824		
Ser	Asn	Pro	Thr	Lys	Pro	Leu	Gln	Leu	Tyr	Arg	Gln	Trp	Thr	Asp	Arg			
595								600					605					
ata	atg	gag	gag	ttc	ttc	cgc	caa	gga	gac	cga	gag	agg	gaa	cgt	ggc	1872		
Ile	Met	Glu	Glu	Phe	Phe	Arg	Gln	Gly	Asp	Arg	Glu	Arg	Glu	Arg	Gly			
610								615					620					
atg	gag	ata	agc	ccc	atg	tgt	gac	aag	cac	aat	gct	tcc	gtg	gaa	aaa	1920		
Met	Glu	Ile	Ser	Pro	Met	Cys	Asp	Lys	His	Asn	Ala	Ser	Val	Glu	Lys			
625								630					635					640
tca	cag	gtg	ggc	ttc	ata	gac	tat	att	gtt	cat	ccc	ctc	tgg	gag	aca	1968		
Ser	Gln	Val	Gly	Phe	Ile	Asp	Tyr	Ile	Val	His	Pro	Leu	Trp	Glu	Thr			
645								650					655					
tgg	gca	gac	ctc	gtc	cac	cct	gac	gcc	cag	gat	att	ttg	gac	act	ttg	2016		
Trp	Ala	Asp	Leu	Val	His	Pro	Asp	Ala	Gln	Asp	Ile	Leu	Asp	Thr	Leu			
660								665					670					
gag	gac	aat	cgt	gaa	tgg	tac	cag	agc	aca	atc	cct	cag	agc	ccc	tct	2064		
Glu	Asp	Asn	Arg	Glu	Trp	Tyr	Gln	Ser	Thr	Ile	Pro	Gln	Ser	Pro	Ser			
675								680					685					
cct	gca	cct	gat	gac	cca	gag	gag	ggc	cgg	cag	ggt	caa	act	gag	aaa	2112		
Pro	Ala	Pro	Asp	Asp	Pro	Glu	Glu	Gly	Arg	Gln	Gly	Gln	Thr	Glu	Lys			
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ttc	cag	ttt	gaa	cta	act	tta	gag	gaa	gat	ggt	gag	tca	gac	acg	gaa	2160		
Phe	Gln	Phe	Glu	Leu	Thr	Leu	Glu	Glu	Asp	Gly	Glu	Ser	Asp	Thr	Glu			
705								710					715					720
aag	gac	agt	ggc	agt	caa	gtg	gaa	gaa	gac	act	agc	tgc	agt	gac	tcc	2208		

Lys	Asp	Ser	Gly	Ser	Gln	Val	Glu	Glu	Asp	Thr	Ser	Cys	Ser	Asp	Ser		
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Lys	Thr	Leu	Cys	Thr	Gln	Asp	Ser	Glu	Ser	Thr	Glu	Ile	Pro	Leu	Asp		
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Glu	Gln	Val	Glu	Glu	Glu	Ala	Val	Gly	Glu	Glu	Glu	Glu	Ser	Gln	Pro		
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gaa	gcc	tgt	gtc	ata	gat	gat	cgt	tct	cct	gac	acg	ccg	cgg	gcc	cgg	2352	
Glu	Ala	Cys	Val	Ile	Asp	Asp	Arg	Ser	Pro	Asp	Thr	Pro	Arg	Ala	Arg		
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Asp	Pro	Pro	Val	Ala	Thr	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr		
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ggg	gtg	gtg	ccc	atc	ctg	gtc	gag	ctg	gac	ggc	gac	gta	aac	ggc	cac	2448	
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Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys		
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ctg	acc	ctg	aag	ttc	atc	tgc	acc	acc	ggc	aag	ctg	ccc	gtg	ccc	tgg	2544	
Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp		
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Pro	Thr	Leu	Val	Thr	Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg		
	850					855				860							
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Tyr	Pro	Asp	His	Met	Lys	Gln	His	Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro		
	865				870				875					880			
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Glu	Gly	Tyr	Val	Gln	Glu	Arg	Thr	Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn		
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Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn		
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ggg	cac	aag	ctg	gag	tac	aac	tac	aac	agc	cac	aac	gtc	tat	atc	atg	2832	
Gly	His	Lys	Leu	Glu	Tyr	Asn	Tyr	Asn	Ser	His	Asn	Val	Tyr	Ile	Met		
	930					935					940						
gcc	gac	aag	cag	aag	aac	ggc	atc	aag	gtg	aac	ttc	aag	atc	cgc	cac	2880	
Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	Lys	Val	Asn	Phe	Lys	Ile	Arg	His		
	945				950					955				960			

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acc ccc atc ggc gac ggc ccc gtg ctg ctg ccc gac aac cac tac ctg      2976
Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu
          980                      985                      990

agc acc cag tcc gcc ctg agc aaa gac ccc aac gag aag cgc gat cac      3024
Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His
          995                      1000                      1005

atg gtc ctg ctg gag ttc gtg acc gcc gcc ggg atc act ctc ggc atg      3072
Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met
          1010                      1015                      1020

gac gag ctg tac aag taa      3090
Asp Glu Leu Tyr Lys *
1025

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<210> 24

<211> 1029

<212> PRT

<213> Fusion between Aequorea victoria and humanArtifici

<220>

<223> Fusion between Aequorea victoria and human

<400> 24

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Pro Gly Pro Arg Glu Gly Gln Ala Thr Leu Lys Pro Pro Pro Gln His
          20          25          30
Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr
          35          40          45
Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro
          50          55          60
Ile Glu Arg Ala Asp Ala Met Asp Thr Ser Asp Arg Pro Gly Leu Arg
          65          70          75          80
Thr Thr Arg Met Ser Trp Pro Ser Ser Phe His Gly Thr Gly Thr Gly
          85          90          95
Ser Gly Gly Ala Gly Gly Gly Ser Ser Arg Arg Phe Glu Ala Glu Asn
          100          105          110
Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser
          115          120          125
Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu
          130          135          140
Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp Met Ser Pro Lys Thr
          145          150          155          160
Met Ser Arg Asn Ser Ser Val Thr Ser Glu Ala His Ala Glu Asp Leu
          165          170          175
Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser Leu Arg Ser Val Arg
          180          185          190
Ser Asn Phe Ser Leu Leu Thr Asn Val Pro Val Pro Ser Asn Lys Arg
          195          200          205
Ser Pro Leu Gly Gly Pro Thr Pro Val Cys Lys Ala Thr Leu Ser Glu
          210          215          220
Glu Thr Cys Gln Gln Leu Ala Arg Glu Thr Leu Glu Glu Leu Asp Trp

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225		230		235		240									
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Met	Ala	Ser	His	Lys	Phe	Lys	Arg	Met	Leu	Asn	Arg	Glu	Leu	Thr	His
			260					265					270		
Leu	Ser	Glu	Met	Ser	Arg	Ser	Gly	Asn	Gln	Val	Ser	Glu	Tyr	Ile	Ser
		275					280					285			
Thr	Thr	Phe	Leu	Asp	Lys	Gln	Asn	Glu	Val	Glu	Ile	Pro	Ser	Pro	Thr
		290				295					300				
Met	Lys	Glu	Arg	Glu	Lys	Gln	Gln	Ala	Pro	Arg	Pro	Arg	Pro	Ser	Gln
305					310					315					320
Pro	Pro	Pro	Pro	Pro	Val	Pro	His	Leu	Gln	Pro	Met	Ser	Gln	Ile	Thr
				325					330					335	
Gly	Leu	Lys	Lys	Leu	Met	His	Ser	Ser	Ser	Leu	Thr	Asn	Ser	Ser	Ile
			340					345					350		
Pro	Arg	Phe	Gly	Val	Lys	Thr	Glu	Gln	Glu	Asp	Val	Leu	Ala	Lys	Glu
		355					360					365			
Leu	Glu	Asp	Val	Asn	Lys	Trp	Gly	Leu	His	Val	Phe	Arg	Ile	Ala	Glu
370						375					380				
Leu	Ser	Gly	Asn	Arg	Pro	Leu	Thr	Val	Ile	Met	His	Thr	Ile	Phe	Gln
385					390					395					400
Glu	Arg	Asp	Leu	Leu	Lys	Thr	Phe	Lys	Ile	Pro	Val	Asp	Thr	Leu	Ile
			405						410					415	
Thr	Tyr	Leu	Met	Thr	Leu	Glu	Asp	His	Tyr	His	Ala	Asp	Val	Ala	Tyr
		420						425					430		
His	Asn	Asn	Ile	His	Ala	Ala	Asp	Val	Val	Gln	Ser	Thr	His	Val	Leu
	435						440					445			
Leu	Ser	Thr	Pro	Ala	Leu	Glu	Ala	Val	Phe	Thr	Asp	Leu	Glu	Ile	Leu
450						455					460				
Ala	Ala	Ile	Phe	Ala	Ser	Ala	Ile	His	Asp	Val	Asp	His	Pro	Gly	Val
465					470					475					480
Ser	Asn	Gln	Phe	Leu	Ile	Asn	Thr	Asn	Ser	Glu	Leu	Ala	Leu	Met	Tyr
			485					490					495		
Asn	Asp	Ser	Ser	Val	Leu	Glu	Asn	His	His	Leu	Ala	Val	Gly	Phe	Lys
		500						505					510		
Leu	Leu	Gln	Glu	Glu	Asn	Cys	Asp	Ile	Phe	Gln	Asn	Leu	Thr	Lys	Lys
		515					520					525			
Gln	Arg	Gln	Ser	Leu	Arg	Lys	Met	Val	Ile	Asp	Ile	Val	Leu	Ala	Thr
530						535					540				
Asp	Met	Ser	Lys	His	Met	Asn	Leu	Leu	Ala	Asp	Leu	Lys	Thr	Met	Val
545					550					555					560
Glu	Thr	Lys	Lys	Val	Thr	Ser	Ser	Gly	Val	Leu	Leu	Leu	Asp	Asn	Tyr
			565					570					575		
Ser	Asp	Arg	Ile	Gln	Val	Leu	Gln	Asn	Met	Val	His	Cys	Ala	Asp	Leu
			580					585					590		
Ser	Asn	Pro	Thr	Lys	Pro	Leu	Gln	Leu	Tyr	Arg	Gln	Trp	Thr	Asp	Arg
		595					600					605			
Ile	Met	Glu	Glu	Phe	Phe	Arg	Gln	Gly	Asp	Arg	Glu	Arg	Glu	Arg	Gly
610						615					620				
Met	Glu	Ile	Ser	Pro	Met	Cys	Asp	Lys	His	Asn	Ala	Ser	Val	Glu	Lys
625					630					635					640
Ser	Gln	Val	Gly	Phe	Ile	Asp	Tyr	Ile	Val	His	Pro	Leu	Trp	Glu	Thr
			645						650				655		
Trp	Ala	Asp	Leu	Val	His	Pro	Asp	Ala	Gln	Asp	Ile	Leu	Asp	Thr	Leu
		660						665				670			
Glu	Asp	Asn	Arg	Glu	Trp	Tyr	Gln	Ser	Thr	Ile	Pro	Gln	Ser	Pro	Ser
		675					680					685			
Pro	Ala	Pro	Asp	Asp	Pro	Glu	Glu	Gly	Arg	Gln	Gly	Gln	Thr	Glu	Lys
690						695					700				

Phe Gln Phe Glu Leu Thr Leu Glu Glu Asp Gly Glu Ser Asp Thr Glu
 705 710 715 720
 Lys Asp Ser Gly Ser Gln Val Glu Glu Asp Thr Ser Cys Ser Asp Ser
 725 730 735
 Lys Thr Leu Cys Thr Gln Asp Ser Glu Ser Thr Glu Ile Pro Leu Asp
 740 745 750
 Glu Gln Val Glu Glu Glu Ala Val Gly Glu Glu Glu Glu Ser Gln Pro
 755 760 765
 Glu Ala Cys Val Ile Asp Asp Arg Ser Pro Asp Thr Pro Arg Ala Arg
 770 775 780
 Asp Pro Pro Val Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr
 785 790 795 800
 Gly Val Val Pro Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His
 805 810 815
 Lys Phe Ser Val Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys
 820 825 830
 Leu Thr Leu Lys Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp
 835 840 845
 Pro Thr Leu Val Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg
 850 855 860
 Tyr Pro Asp His Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro
 865 870 875 880
 Glu Gly Tyr Val Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn
 885 890 895
 Tyr Lys Thr Arg Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn
 900 905 910
 Arg Ile Glu Leu Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu
 915 920 925
 Gly His Lys Leu Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met
 930 935 940
 Ala Asp Lys Gln Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His
 945 950 955 960
 Asn Ile Glu Asp Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn
 965 970 975
 Thr Pro Ile Gly Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu
 980 985 990
 Ser Thr Gln Ser Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His
 995 1000 1005
 Met Val Leu Leu Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met
 1010 1015 1020
 Asp Glu Leu Tyr Lys
 1025

<210> 25

<211> 2790

<212> DNA

<213> Fusion between Aequorea victoria and humanArtifici

<220>

<223> Fusion between Aequorea victoria and human

<221> CDS

<222> (1)... (2790)

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 Met Met His Val Asn Asn Phe Pro Phe Arg Arg His Ser Trp Ile Cys
 1 5 10 15

40

ttt gat gtg gac aat ggc aca tct gcg gga cgg agt ccc ttg gat ccc Phe Asp Val Asp Asn Gly Thr Ser Ala Gly Arg Ser Pro Leu Asp Pro 20 25 30	96
atg acc agc cca gga tcc ggg cta att ctc caa gca aat ttt gtc cac Met Thr Ser Pro Gly Ser Gly Leu Ile Leu Gln Ala Asn Phe Val His 35 40 45	144
agt caa cga cgg gag tcc ttc ctg tat cga tcc gac agc gat tat gac Ser Gln Arg Arg Glu Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp 50 55 60	192
ctc tct cca aag tct atg tcc cgg aac tcc tcc att gcc agt gat ata Leu Ser Pro Lys Ser Met Ser Arg Asn Ser Ser Ile Ala Ser Asp Ile 65 70 75 80	240
cac gga gat gac ttg att gtg act cca ttt gct cag gtc ttg gcc agt His Gly Asp Asp Leu Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser 85 90 95	288
ctg cga act gta cga aac aac ttt gct gca tta act aat ttg caa gat Leu Arg Thr Val Arg Asn Asn Phe Ala Leu Thr Asn Leu Gln Asp 100 105 110	336
cga gca cct agc aaa aga tca ccc atg tgc aac caa cca tcc atc aac Arg Ala Pro Ser Lys Arg Ser Pro Met Cys Asn Gln Pro Ser Ile Asn 115 120 125	384
aaa gcc acc ata aca gag gag gcc tac cag aaa ctg gcc agc gag acc Lys Ala Thr Ile Thr Glu Glu Ala Tyr Gln Lys Leu Ala Ser Glu Thr 130 135 140	432
ctg gag gag ctg gac tgg tgt ctg gac cag cta gag acc cta cag acc Leu Glu Glu Leu Asp Trp Cys Leu Asp Gln Leu Glu Thr Leu Gln Thr 145 150 155 160	480
agg cac tcc gtc agt gag atg gcc tcc aac aag ttt aaa agg atg ctt Arg His Ser Val Ser Glu Met Ala Ser Asn Lys Phe Lys Arg Met Leu 165 170 175	528
aat cgg gag ctc acc cat ctc tct gaa atg agt cgg tct gga aat caa Asn Arg Glu Leu Thr His Leu Ser Glu Met Ser Arg Ser Gly Asn Gln 180 185 190	576
gtg tca gag ttt ata tca aac aca ttc tta gat aag caa cat gaa gtg Val Ser Glu Phe Ile Ser Asn Thr Phe Leu Asp Lys Gln His Glu Val 195 200 205	624
gaa att cct tct cca act cag aag gaa aag gag aaa aag aaa aga cca Glu Ile Pro Ser Pro Thr Gln Lys Glu Lys Glu Lys Lys Lys Arg Pro 210 215 220	672
atg tct cag atc agt gga gtc aag aaa ttg atg cac agc tct agt ctg Met Ser Gln Ile Ser Gly Val Lys Lys Leu Met His Ser Ser Ser Leu 225 230 235 240	720
act aat tca agt atc cca agg ttt gga gtt aaa act gaa caa gaa gat Thr Asn Ser Ser Ile Pro Arg Phe Gly Val Lys Thr Glu Gln Glu Asp 245 250 255	768

gtc ctt gcc aag gaa cta gaa gat gtg aac aaa tgg ggt ctt cat gtt	816
Val Leu Ala Lys Glu Leu Glu Asp Val Asn Lys Trp Gly Leu His Val	
260 265 270	
ttc aga ata gca gag ttg tct ggt aac cgg ccc ttg act gtt atc atg	864
Phe Arg Ile Ala Glu Leu Ser Gly Asn Arg Pro Leu Thr Val Ile Met	
275 280 285	
cac acc att ttt cag gaa cgg gat tta tta aaa aca ttt aaa att cca	912
His Thr Ile Phe Gln Glu Arg Asp Leu Leu Lys Thr Phe Lys Ile Pro	
290 295 300	
gta gat act tta att aca tat ctt atg act ctc gaa gac cat tac cat	960
Val Asp Thr Leu Ile Thr Tyr Leu Met Thr Leu Glu Asp His Tyr His	
305 310 315 320	
gct gat gtg gcc tat cac aac aat atc cat gct gca gat gtt gtc cag	1008
Ala Asp Val Ala Tyr His Asn Asn Ile His Ala Ala Asp Val Val Gln	
325 330 335	
tct act cat gtg cta tta tct aca cct gct ttg gag gct gtg ttt aca	1056
Ser Thr His Val Leu Leu Ser Thr Pro Ala Leu Glu Ala Val Phe Thr	
340 345 350	
gat ttg gag att ctt gca gca att ttt gcc agt gca ata cat gat gta	1104
Asp Leu Glu Ile Leu Ala Ala Ile Phe Ala Ser Ala Ile His Asp Val	
355 360 365	
gat cat cct ggt gtg tcc aat caa ttt ctg atc aat aca aac tct gaa	1152
Asp His Pro Gly Val Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu	
370 375 380	
ctt gcc ttg atg tac aat gat tcc tca gtc tta gag aac cat cat ttg	1200
Leu Ala Leu Met Tyr Asn Asp Ser Ser Val Leu Glu Asn His His Leu	
385 390 395 400	
gct gtg ggc ttt aaa ttg ctt cag gaa gaa aac tgt gac att ttc cag	1248
Ala Val Gly Phe Lys Leu Leu Gln Glu Glu Asn Cys Asp Ile Phe Gln	
405 410 415	
aat ttg acc aaa aaa caa aga caa tct tta agg aaa atg gtc att gac	1296
Asn Leu Thr Lys Lys Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp	
420 425 430	
atc gta ctt gca aca gat atg tca aaa cac atg aat cta ctg gct gat	1344
Ile Val Leu Ala Thr Asp Met Ser Lys His Met Asn Leu Leu Ala Asp	
435 440 445	
ttg aag act atg gtt gaa act aag aaa gtg aca agc tct gga gtt ctt	1392
Leu Lys Thr Met Val Glu Thr Lys Lys Val Thr Ser Ser Gly Val Leu	
450 455 460	
ctt ctt gat aat tat tcc gat agg att cag gtt ctt cag aat atg gtg	1440
Leu Leu Asp Asn Tyr Ser Asp Arg Ile Gln Val Leu Gln Asn Met Val	
465 470 475 480	
cac tgt gca gat ctg agc aac cca aca aag cct ctc cag ctg tac cgc	1488
His Cys Ala Asp Leu Ser Asn Pro Thr Lys Pro Leu Gln Leu Tyr Arg	

485	490	495	
cag tgg acg gac cgg ata atg gag gag ttc ttc cgc caa gga gac cga Gln Trp Thr Asp Arg Ile Met Glu Glu Phe Phe Arg Gln Gly Asp Arg 500 505 510			1536
gag agg gaa cgt ggc atg gag ata agc ccc atg tgt gac aag cac aat Glu Arg Glu Arg Gly Met Glu Ile Ser Pro Met Cys Asp Lys His Asn 515 520 525			1584
gct tcc gtg gaa aaa tca cag gtg ggc ttc ata gac tat att gtt cat Ala Ser Val Glu Lys Ser Gln Val Gly Phe Ile Asp Tyr Ile Val His 530 535 540			1632
ccc ctc tgg gag aca tgg gca gac ctc gtc cac cct gac gcc cag gat Pro Leu Trp Glu Thr Trp Ala Asp Leu Val His Pro Asp Ala Gln Asp 545 550 555 560			1680
att ttg gac act ttg gag gac aat cgt gaa tgg tac cag agc aca atc Ile Leu Asp Thr Leu Glu Asp Asn Arg Glu Trp Tyr Gln Ser Thr Ile 565 570 575			1728
cct cag agc ccc tct cct gca cct gat gac cca gag gag ggc cgg cag Pro Gln Ser Pro Ser Pro Ala Pro Asp Asp Pro Glu Glu Gly Arg Gln 580 585 590			1776
ggt caa act gag aaa ttc cag ttt gaa cta act tta gag gaa gat ggt Gly Gln Thr Glu Lys Phe Gln Phe Glu Leu Thr Leu Glu Glu Asp Gly 595 600 605			1824
gag tca gac acg gaa aag gac agt ggc agt caa gtg gaa gaa gac act Glu Ser Asp Thr Glu Lys Asp Ser Gly Ser Gln Val Glu Glu Asp Thr 610 615 620			1872
agc tgc agt gac tcc aag act ctt tgt act caa gac tca gag tct act Ser Cys Ser Asp Ser Lys Thr Leu Cys Thr Gln Asp Ser Glu Ser Thr 625 630 635 640			1920
gaa att ccc ctt gat gaa cag gtt gaa gag gag gca gta ggg gaa gaa Glu Ile Pro Leu Asp Glu Gln Val Glu Glu Glu Ala Val Gly Glu Glu 645 650 655			1968
gag gaa agc cag cct gaa gcc tgt gtc ata gat gat cgt tct cct gac Glu Glu Ser Gln Pro Glu Ala Cys Val Ile Asp Asp Arg Ser Pro Asp 660 665 670			2016
acg cga att ctg cag tcg acg gta ccg cgg gcc cgg gat cca ccg gtc Thr Arg Ile Leu Gln Ser Thr Val Pro Arg Ala Arg Asp Pro Pro Val 675 680 685			2064
gcc acc atg gtg agc aag ggc gag gag ctg ttc acc ggg gtg gtg ccc Ala Thr Met Val Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro 690 695 700			2112
atc ctg gtc gag ctg gac ggc gac gta aac ggc cac aag ttc agc gtg Ile Leu Val Glu Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val 705 710 715 720			2160
tcc ggc gag ggc gag ggc gat gcc acc tac ggc aag ctg acc ctg aag			2208

Ser Gly Glu Gly Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys	
725 730 735	
ttc atc tgc acc acc ggc aag ctg ccc gtg ccc tgg ccc acc ctc gtg	2256
Phe Ile Cys Thr Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val	
740 745 750	
acc acc ctg acc tac ggc gtg cag tgc ttc agc cgc tac ccc gac cac	2304
Thr Thr Leu Thr Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His	
755 760 765	
atg aag cag cac gac ttc ttc aag tcc gcc atg ccc gaa ggc tac gtc	2352
Met Lys Gln His Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val	
770 775 780	
cag gag cgc acc atc ttc ttc aag gac gac ggc aac tac aag acc cgc	2400
Gln Glu Arg Thr Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg	
785 790 795 800	
gcc gag gtg aag ttc gag ggc gac acc ctg gtg aac cgc atc gag ctg	2448
Ala Glu Val Lys Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu	
805 810 815	
aag ggc atc gac ttc aag gag gac ggc aac atc ctg ggg cac aag ctg	2496
Lys Gly Ile Asp Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu	
820 825 830	
gag tac aac tac aac agc cac aac gtc tat atc atg gcc gac aag cag	2544
Glu Tyr Asn Tyr Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln	
835 840 845	
aag aac ggc atc aag gtg aac ttc aag atc cgc cac aac atc gag gac	2592
Lys Asn Gly Ile Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp	
850 855 860	
ggc agc gtg cag ctc gcc gac cac tac cag cag aac acc ccc atc ggc	2640
Gly Ser Val Gln Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly	
865 870 875 880	
gac ggc ccc gtg ctg ctg ccc gac aac cac tac ctg agc acc cag tcc	2688
Asp Gly Pro Val Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser	
885 890 895	
gcc ctg agc aaa gac ccc aac gag aag cgc gat cac atg gtc ctg ctg	2736
Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu	
900 905 910	
gag ttc gtg acc gcc gcc ggg atc act ctc ggc atg gac gag ctg tac	2784
Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr	
915 920 925	
aag taa	2790
Lys *	

<210> 26
 <211> 929
 <212> PRT

<213> Fusion between Aequorea victoria and humanArtifici

<220>

<223> Fusion between Aequorea victoria and human

<400> 26

Met	Met	His	Val	Asn	Asn	Phe	Pro	Phe	Arg	Arg	His	Ser	Trp	Ile	Cys
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Phe	Asp	Val	Asp	Asn	Gly	Thr	Ser	Ala	Gly	Arg	Ser	Pro	Leu	Asp	Pro
			20					25					30		
Met	Thr	Ser	Pro	Gly	Ser	Gly	Leu	Ile	Leu	Gln	Ala	Asn	Phe	Val	His
			35				40					45			
Ser	Gln	Arg	Arg	Glu	Ser	Phe	Leu	Tyr	Arg	Ser	Asp	Ser	Asp	Tyr	Asp
	50					55				60					
Leu	Ser	Pro	Lys	Ser	Met	Ser	Arg	Asn	Ser	Ser	Ile	Ala	Ser	Asp	Ile
65					70					75					80
His	Gly	Asp	Asp	Leu	Ile	Val	Thr	Pro	Phe	Ala	Gln	Val	Leu	Ala	Ser
				85					90					95	
Leu	Arg	Thr	Val	Arg	Asn	Asn	Phe	Ala	Ala	Leu	Thr	Asn	Leu	Gln	Asp
			100				105						110		
Arg	Ala	Pro	Ser	Lys	Arg	Ser	Pro	Met	Cys	Asn	Gln	Pro	Ser	Ile	Asn
			115				120					125			
Lys	Ala	Thr	Ile	Thr	Glu	Glu	Ala	Tyr	Gln	Lys	Leu	Ala	Ser	Glu	Thr
	130					135					140				
Leu	Glu	Glu	Leu	Asp	Trp	Cys	Leu	Asp	Gln	Leu	Glu	Thr	Leu	Gln	Thr
145					150					155					160
Arg	His	Ser	Val	Ser	Glu	Met	Ala	Ser	Asn	Lys	Phe	Lys	Arg	Met	Leu
				165					170					175	
Asn	Arg	Glu	Leu	Thr	His	Leu	Ser	Glu	Met	Ser	Arg	Ser	Gly	Asn	Gln
			180					185					190		
Val	Ser	Glu	Phe	Ile	Ser	Asn	Thr	Phe	Leu	Asp	Lys	Gln	His	Glu	Val
			195				200					205			
Glu	Ile	Pro	Ser	Pro	Thr	Gln	Lys	Glu	Lys	Glu	Lys	Lys	Lys	Arg	Pro
	210					215					220				
Met	Ser	Gln	Ile	Ser	Gly	Val	Lys	Lys	Leu	Met	His	Ser	Ser	Ser	Leu
225					230					235					240
Thr	Asn	Ser	Ser	Ile	Pro	Arg	Phe	Gly	Val	Lys	Thr	Glu	Gln	Glu	Asp
				245					250					255	
Val	Leu	Ala	Lys	Glu	Leu	Glu	Asp	Val	Asn	Lys	Trp	Gly	Leu	His	Val
			260					265					270		
Phe	Arg	Ile	Ala	Glu	Leu	Ser	Gly	Asn	Arg	Pro	Leu	Thr	Val	Ile	Met
			275				280					285			
His	Thr	Ile	Phe	Gln	Glu	Arg	Asp	Leu	Leu	Lys	Thr	Phe	Lys	Ile	Pro
	290					295					300				
Val	Asp	Thr	Leu	Ile	Thr	Tyr	Leu	Met	Thr	Leu	Glu	Asp	His	Tyr	His
				310						315					320
Ala	Asp	Val	Ala	Tyr	His	Asn	Asn	Ile	His	Ala	Ala	Asp	Val	Val	Gln
				325					330					335	
Ser	Thr	His	Val	Leu	Leu	Ser	Thr	Pro	Ala	Leu	Glu	Ala	Val	Phe	Thr
			340					345					350		
Asp	Leu	Glu	Ile	Leu	Ala	Ala	Ile	Phe	Ala	Ser	Ala	Ile	His	Asp	Val
			355				360					365			
Asp	His	Pro	Gly	Val	Ser	Asn	Gln	Phe	Leu	Ile	Asn	Thr	Asn	Ser	Glu
	370					375					380				
Leu	Ala	Leu	Met	Tyr	Asn	Asp	Ser	Ser	Val	Leu	Glu	Asn	His	His	Leu
385					390						395				400
Ala	Val	Gly	Phe	Lys	Leu	Leu	Gln	Glu	Glu	Asn	Cys	Asp	Ile	Phe	Gln
				405					410					415	
Asn	Leu	Thr	Lys	Lys	Gln	Arg	Gln	Ser	Leu	Arg	Lys	Met	Val	Ile	Asp

			420						425						430					
Ile	Val	Leu	Ala	Thr	Asp	Met	Ser	Lys	His	Met	Asn	Leu	Leu	Ala	Asp					
		435					440					445								
Leu	Lys	Thr	Met	Val	Glu	Thr	Lys	Lys	Val	Thr	Ser	Ser	Gly	Val	Leu					
		450				455					460									
Leu	Leu	Asp	Asn	Tyr	Ser	Asp	Arg	Ile	Gln	Val	Leu	Gln	Asn	Met	Val					
465					470					475					480					
His	Cys	Ala	Asp	Leu	Ser	Asn	Pro	Thr	Lys	Pro	Leu	Gln	Leu	Tyr	Arg					
				485					490					495						
Gln	Trp	Thr	Asp	Arg	Ile	Met	Glu	Glu	Phe	Phe	Arg	Gln	Gly	Asp	Arg					
			500				505						510							
Glu	Arg	Glu	Arg	Gly	Met	Glu	Ile	Ser	Pro	Met	Cys	Asp	Lys	His	Asn					
		515				520					525									
Ala	Ser	Val	Glu	Lys	Ser	Gln	Val	Gly	Phe	Ile	Asp	Tyr	Ile	Val	His					
		530				535					540									
Pro	Leu	Trp	Glu	Thr	Trp	Ala	Asp	Leu	Val	His	Pro	Asp	Ala	Gln	Asp					
545					550					555					560					
Ile	Leu	Asp	Thr	Leu	Glu	Asp	Asn	Arg	Glu	Trp	Tyr	Gln	Ser	Thr	Ile					
				565					570					575						
Pro	Gln	Ser	Pro	Ser	Pro	Ala	Pro	Asp	Asp	Pro	Glu	Glu	Gly	Arg	Gln					
			580					585					590							
Gly	Gln	Thr	Glu	Lys	Phe	Gln	Phe	Glu	Leu	Thr	Leu	Glu	Glu	Asp	Gly					
		595				600					605									
Glu	Ser	Asp	Thr	Glu	Lys	Asp	Ser	Gly	Ser	Gln	Val	Glu	Glu	Asp	Thr					
		610				615				620										
Ser	Cys	Ser	Asp	Ser	Lys	Thr	Leu	Cys	Thr	Gln	Asp	Ser	Glu	Ser	Thr					
625					630					635					640					
Glu	Ile	Pro	Leu	Asp	Glu	Gln	Val	Glu	Glu	Ala	Val	Gly	Glu	Glu						
			645					650					655							
Glu	Glu	Ser	Gln	Pro	Glu	Ala	Cys	Val	Ile	Asp	Asp	Arg	Ser	Pro	Asp					
			660					665					670							
Thr	Arg	Ile	Leu	Gln	Ser	Thr	Val	Pro	Arg	Ala	Arg	Asp	Pro	Pro	Val					
		675					680					685								
Ala	Thr	Met	Val	Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro					
						695				700										
Ile	Leu	Val	Glu	Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val					
705					710					715					720					
Ser	Gly	Glu	Gly	Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys					
				725					730					735						
Phe	Ile	Cys	Thr	Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val					
			740					745					750							
Thr	Thr	Leu	Thr	Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His					
		755					760					765								

Ala Leu Ser Lys Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu
 900 905 910
 Glu Phe Val Thr Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr
 915 920 925
 Lys

<210> 27
 <211> 29
 <212> DNA
 <213> PArtificial Sequence

<220>
 <223> Primer sequence

<400> 27
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<210> 28
 <211> 29
 <212> DNA
 <213> PArtificial Sequence

<220>
 <223> Primer sequence

<400> 28
 gtgaattccc gtgtcaggag aacgatcat 29

<210> 29
 <211> 2682
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Fusion between Aequorea victoria and human

<221> CDS
 <222> (1)...(2682)

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 Met Pro Leu Val Asp Phe Phe Cys Glu Thr Cys Ser Lys Pro Trp Leu
 1 5 10 15

gtg ggc tgg tgg gac cag ttt aaa agg atg ttg aac cgt gag ctc aca 96
 Val Gly Trp Trp Asp Gln Phe Lys Arg Met Leu Asn Arg Glu Leu Thr
 20 25 30

cac ctg tca gaa atg agc agg tcc gga aac cag gtc tca gag tac att 144
 His Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile
 35 40 45

tcc aca aca ttc ctg gac aaa cag aat gaa gtg gag atc cca tca ccc 192
 Ser Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro
 50 55 60

acg atg aag gaa cga gaa aaa cag caa gcg ccg cga cca aga ccc tcc 240
 Thr Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser

65	70	75	80	
cag ccg ccc ccg ccc cct gta cca cac tta cag ccc atg tcc caa atc				288
Gln Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile				
	85	90	95	
aca ggg ttg aaa aag ttg atg cat agt aac agc ctg aac aac tct aac				336
Thr Gly Leu Lys Lys Leu Met His Ser Asn Ser Leu Asn Asn Ser Asn				
	100	105	110	
att ccc cga ttt ggg gtg aag acc gat caa gaa gag ctc ctg gcc caa				384
Ile Pro Arg Phe Gly Val Lys Thr Asp Gln Glu Glu Leu Leu Ala Gln				
	115	120	125	
gaa ctg gag aac ctg aac aag tgg ggc ctg aac atc ttt tgc gtg tcg				432
Glu Leu Glu Asn Leu Asn Lys Trp Gly Leu Asn Ile Phe Cys Val Ser				
	130	135	140	
gat tac gct gga ggc cgc tca ctc acc tgc atc atg tac atg ata ttc				480
Asp Tyr Ala Gly Gly Arg Ser Leu Thr Cys Ile Met Tyr Met Ile Phe				
	145	150	155	160
cag gag cgg gac ctg ctg aag aaa ttc cgc atc ccg gtg gac acg atg				528
Gln Glu Arg Asp Leu Leu Lys Lys Phe Arg Ile Pro Val Asp Thr Met				
	165	170	175	
gtg aca tac atg ctg acg ctg gag gat cac tac cac gct gac gtg gcc				576
Val Thr Tyr Met Leu Thr Leu Glu Asp His Tyr His Ala Asp Val Ala				
	180	185	190	
tac cat aac agc ctg cac gca gct gac gtg ctg cag tcc acc cac gta				624
Tyr His Asn Ser Leu His Ala Ala Asp Val Leu Gln Ser Thr His Val				
	195	200	205	
ctg ctg gcc acg cct gca cta gat gca gtg ttc acg gac ctg gag att				672
Leu Leu Ala Thr Pro Ala Leu Asp Ala Val Phe Thr Asp Leu Glu Ile				
	210	215	220	
ctc gcc gcc ctc ttc gcg gct gcc atc cac gat gtg gat cac cct ggg				720
Leu Ala Ala Leu Phe Ala Ala Ala Ile His Asp Val Asp His Pro Gly				
	225	230	235	240
gtc tcc aac cag ttc ctc atc aac acc aat tcg gag ctg gcg ctc atg				768
Val Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met				
	245	250	255	
tac aac gat gag tcg gtg ctc gag aat cac cac ctg gcc gtg ggc ttc				816
Tyr Asn Asp Glu Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe				
	260	265	270	
aag ctg ctg cag gag gac aac tgc gac atc ttc cag aac ctc agc aag				864
Lys Leu Leu Gln Glu Asp Asn Cys Asp Ile Phe Gln Asn Leu Ser Lys				
	275	280	285	
cgc cag cgg cag agc cta cgc aag atg gtc atc gac atg gtg ctg gcc				912
Arg Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp Met Val Leu Ala				
	290	295	300	
acg gac atg tcc aag cac atg acc ctc ctg gct gac ctg aag acc atg				960

Thr	Asp	Met	Ser	Lys	His	Met	Thr	Leu	Leu	Ala	Asp	Leu	Lys	Thr	Met	
305					310					315					320	
gtg	gag	acc	aag	aaa	gtg	acc	agc	tca	ggg	gtc	ctc	ctg	cta	gat	aac	1008
Val	Glu	Thr	Lys	Lys	Val	Thr	Ser	Ser	Gly	Val	Leu	Leu	Leu	Asp	Asn	
			325						330					335		
tac	tcc	gac	cgc	atc	cag	gtc	ctc	cgg	aac	atg	gtg	cac	tgt	gcc	gac	1056
Tyr	Ser	Asp	Arg	Ile	Gln	Val	Leu	Arg	Asn	Met	Val	His	Cys	Ala	Asp	
			340					345					350			
ctc	agc	aac	ccc	acc	aag	cgg	ctg	gag	ctg	tac	cgc	cag	tgg	aca	gac	1104
Leu	Ser	Asn	Pro	Thr	Lys	Pro	Leu	Glu	Leu	Tyr	Arg	Gln	Trp	Thr	Asp	
		355					360					365				
cgc	atc	atg	gcc	gag	ttc	ttc	cag	cag	ggg	gac	cga	gag	cgc	gag	cgt	1152
Arg	Ile	Met	Ala	Glu	Phe	Phe	Gln	Gln	Gly	Asp	Arg	Glu	Arg	Glu	Arg	
	370					375				380						
ggc	atg	gaa	atc	agc	ccc	atg	tgt	gac	aag	cac	act	gcc	tcc	gtg	gag	1200
Gly	Met	Glu	Ile	Ser		Met	Cys	Asp	Lys	His	Thr	Ala	Ser	Val	Glu	
385					390					395					400	
aag	tct	cag	gtg	ggg	ttt	att	gac	tac	att	gtg	cac	cca	ttg	tgg	gag	1248
Lys	Ser	Gln	Val	Gly	Phe	Ile	Asp	Tyr	Ile	Val	His	Pro	Leu	Trp	Glu	
			405						410					415		
acc	tgg	gcg	gac	ctt	gtc	cac	cca	gat	gcc	cag	gag	atc	ttg	gac	act	1296
Thr	Trp	Ala	Asp	Leu	Val	His	Pro	Asp	Ala	Gln	Glu	Ile	Leu	Asp	Thr	
			420					425					430			
ttg	gag	gac	aac	cgg	gac	tgg	tac	tac	agc	gcc	atc	cgg	cag	agc	cca	1344
Leu	Glu	Asp	Asn	Arg	Asp	Trp	Tyr	Tyr	Ser	Ala	Ile	Arg	Gln	Ser	Pro	
		435				440						445				
tct	ccg	cca	ccc	gag	gag	gag	tca	agg	ggg	cca	ggc	cac	cca	ccc	ctg	1392
Ser	Pro	Pro	Pro	Glu	Glu	Glu	Ser	Arg	Gly	Pro	Gly	His	Pro	Pro	Leu	
	450					455					460					
cct	gac	aag	ttc	cag	ttt	gag	ctg	acg	ctg	gag	gag	gaa	gag	gag	gaa	1440
Pro	Asp	Lys	Phe	Gln	Phe	Glu	Leu	Thr	Leu	Glu	Glu	Glu	Glu	Glu	Glu	
465				470					475					480		
gaa	ata	tca	atg	gcc	cag	ata	ccg	tgc	aca	gcc	caa	gag	gca	ttg	act	1488
Glu	Ile	Ser	Met	Ala	Gln	Ile	Pro	Cys	Thr	Ala	Gln	Glu	Ala	Leu	Thr	
			485					490						495		
gcg	cag	gga	ttg	tca	gga	gtc	gag	gaa	gct	ctg	gat	gca	acc	ata	gcc	1536
Ala	Gln	Gly	Leu	Ser	Gly	Val	Glu	Glu	Ala	Leu	Asp	Ala	Thr	Ile	Ala	
		500						505				510				
tgg	gag	gca	tcc	ccg	gcc	cag	gag	tcg	ttg	gaa	gtt	atg	gca	cag	gaa	1584
Trp	Glu	Ala	Ser	Pro	Ala	Gln	Glu	Ser	Leu	Glu	Val	Met	Ala	Gln	Glu	
		515					520					525				
gca	tcc	ctg	gag	gcc	gag	ctg	gag	gca	gtg	tat	ttg	aca	cag	cag	gca	1632
Ala	Ser	Leu	Glu	Ala	Glu	Leu	Glu	Ala	Val	Tyr	Leu	Thr	Gln	Gln	Ala	
	530					535					540					

cag tcc aca ggc agt gca cct gtg gct ccg gat gag ttc tcg tcc cgg Gln Ser Thr Gly Ser Ala Pro Val Ala Pro Asp Glu Phe Ser Ser Arg 545 550 555 560	1680
gag gaa ttc gtg gtt gct gta agc cac agc agc ccc tct gcc ctg gct Glu Glu Phe Val Val Ala Val Ser His Ser Ser Pro Ser Ala Leu Ala 565 570 575	1728
ctt caa agc ccc ctt ctc cct gct tgg agg acc ctg tct gtt tca gag Leu Gln Ser Pro Leu Leu Pro Ala Trp Arg Thr Leu Ser Val Ser Glu 580 585 590	1776
cat gcc ccg ggc ctc ccg ggc ctc ccc tcc acg gcg gcc gag gtg gag His Ala Pro Gly Leu Pro Gly Leu Pro Ser Thr Ala Ala Glu Val Glu 595 600 605	1824
gcc caa cga gag cac cag gct gcc aag agg gct tgc agt gcc tgc gca Ala Gln Arg Glu His Gln Ala Ala Lys Arg Ala Cys Ser Ala Cys Ala 610 615 620	1872
ggg aca ttt ggg gag gac aca tcc gca ctc cca gct cct ggt ggc ggg Gly Thr Phe Gly Glu Asp Thr Ser Ala Leu Pro Ala Pro Gly Gly Gly 625 630 635 640	1920
ggg tca ggt gga gac cct acc tgg gat cca ccg gtc gcc acc atg gtg Gly Ser Gly Gly Asp Pro Thr Trp Asp Pro Pro Val Ala Thr Met Val 645 650 655	1968
agc aag ggc gag gag ctg ttc acc ggg gtg gtg ccc atc ctg gtc gag Ser Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu 660 665 670	2016
ctg gac ggc gac gta aac ggc cac aag ttc agc gtg tcc ggc gag ggc Leu Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly 675 680 685	2064
gag ggc gat gcc acc tac ggc aag ctg acc ctg aag ttc atc tgc acc Glu Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr 690 695 700	2112
acc ggc aag ctg ccc gtg ccc tgg ccc aca cta gtg acc acc ctg tct Thr Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Ser 705 710 715 720	2160
tac ggc gtg cag tgc ttc agc cgc tac ccc gac cac atg aag cag cac Tyr Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His 725 730 735	2208
gac ttc ttc aag tcc gcc atg ccc gaa ggc tac gtc cag gag cgc acc Asp Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr 740 745 750	2256
atc ttc ttc aag gac gac ggc aac tac aag acc cgc gcc gag gtg aag Ile Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys 755 760 765	2304
ttc gag ggc gac acc ctg gtg aac cgc atc gag ctg aag ggc atc gac Phe Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp 770 775 780	2352

ttc aag gag gac ggc aac atc ctg ggg cac aag ctg gag tac aac tac 2400
 Phe Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr
 785 790 795 800

 aac agc cac aac gtc tat atc atg gcc gac aag cag aag aac ggc atc 2448
 Asn Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile
 805 810 815

 aag gtg aac ttc aag atc cgc cac aac atc gag gac ggc agc gtg cag 2496
 Lys Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln
 820 825 830

 ctc gcc gac cac tac cag cag aac acc ccc atc ggc gac ggc ccc gtg 2544
 Leu Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val
 835 840 845

 ctg ctg ccc gac aac cac tac ctg agc acc cag tcc gcc ctg agc aaa 2592
 Leu Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys
 850 855 860

 gac ccc aac gag aag cgc gat cac atg gtc ctc cta ggg ttc gtg acc 2640
 Asp Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Gly Phe Val Thr
 865 870 875 880

 gcc gcc ggg atc act ctc ggc atg gac gag ctg tac aag taa 2682
 Ala Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys *
 885 890

<210> 30

<211> 893

<212> PRT

<213> Artificial Sequence

<220>

<223> Fusion between Aequorea victoria and human

<400> 30

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 Val Gly Trp Trp Asp Gln Phe Lys Arg Met Leu Asn Arg Glu Leu Thr
 20 25 30
 His Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile
 35 40 45
 Ser Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro
 50 55 60
 Thr Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser
 65 70 75 80
 Gln Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile
 85 90 95
 Thr Gly Leu Lys Lys Leu Met His Ser Asn Ser Leu Asn Asn Ser Asn
 100 105 110
 Ile Pro Arg Phe Gly Val Lys Thr Asp Gln Glu Glu Leu Leu Ala Gln
 115 120 125
 Glu Leu Glu Asn Leu Asn Lys Trp Gly Leu Asn Ile Phe Cys Val Ser
 130 135 140
 Asp Tyr Ala Gly Gly Arg Ser Leu Thr Cys Ile Met Tyr Met Ile Phe
 145 150 155 160

Gln Glu Arg Asp Leu Leu Lys Lys Phe Arg Ile Pro Val Asp Thr Met
 165 170 175
 Val Thr Tyr Met Leu Thr Leu Glu Asp His Tyr His Ala Asp Val Ala
 180 185 190
 Tyr His Asn Ser Leu His Ala Ala Asp Val Leu Gln Ser Thr His Val
 195 200 205
 Leu Leu Ala Thr Pro Ala Leu Asp Ala Val Phe Thr Asp Leu Glu Ile
 210 215 220
 Leu Ala Ala Leu Phe Ala Ala Ala Ile His Asp Val Asp His Pro Gly
 225 230 235 240
 Val Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met
 245 250 255
 Tyr Asn Asp Glu Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe
 260 265 270
 Lys Leu Leu Gln Glu Asp Asn Cys Asp Ile Phe Gln Asn Leu Ser Lys
 275 280 285
 Arg Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp Met Val Leu Ala
 290 295 300
 Thr Asp Met Ser Lys His Met Thr Leu Leu Ala Asp Leu Lys Thr Met
 305 310 315 320
 Val Glu Thr Lys Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn
 325 330 335
 Tyr Ser Asp Arg Ile Gln Val Leu Arg Asn Met Val His Cys Ala Asp
 340 345 350
 Leu Ser Asn Pro Thr Lys Pro Leu Glu Leu Tyr Arg Gln Trp Thr Asp
 355 360 365
 Arg Ile Met Ala Glu Phe Phe Gln Gln Gly Asp Arg Glu Arg Glu Arg
 370 375 380
 Gly Met Glu Ile Ser Pro Met Cys Asp Lys His Thr Ala Ser Val Glu
 385 390 395 400
 Lys Ser Gln Val Gly Phe Ile Asp Tyr Ile Val His Pro Leu Trp Glu
 405 410 415
 Thr Trp Ala Asp Leu Val His Pro Asp Ala Gln Glu Ile Leu Asp Thr
 420 425 430
 Leu Glu Asp Asn Arg Asp Trp Tyr Ser Ala Ile Arg Gln Ser Pro
 435 440 445
 Ser Pro Pro Pro Glu Glu Glu Ser Arg Gly Pro Gly His Pro Pro Leu
 450 455 460
 Pro Asp Lys Phe Gln Phe Glu Leu Thr Leu Glu Glu Glu Glu Glu
 465 470 475 480
 Glu Ile Ser Met Ala Gln Ile Pro Cys Thr Ala Gln Glu Ala Leu Thr
 485 490 495
 Ala Gln Gly Leu Ser Gly Val Glu Glu Ala Leu Asp Ala Thr Ile Ala
 500 505 510
 Trp Glu Ala Ser Pro Ala Gln Glu Ser Leu Glu Val Met Ala Gln Glu
 515 520 525
 Ala Ser Leu Glu Ala Glu Leu Glu Ala Val Tyr Leu Thr Gln Gln Ala
 530 535 540
 Gln Ser Thr Gly Ser Ala Pro Val Ala Pro Asp Glu Phe Ser Ser Arg
 545 550 555 560
 Glu Glu Phe Val Val Ala Val Ser His Ser Ser Pro Ser Ala Leu Ala
 565 570 575
 Leu Gln Ser Pro Leu Leu Pro Ala Trp Arg Thr Leu Ser Val Ser Glu
 580 585 590
 His Ala Pro Gly Leu Pro Gly Leu Pro Ser Thr Ala Ala Glu Val Glu
 595 600 605
 Ala Gln Arg Glu His Gln Ala Ala Lys Arg Ala Cys Ser Ala Cys Ala
 610 615 620
 Gly Thr Phe Gly Glu Asp Thr Ser Ala Leu Pro Ala Pro Gly Gly Gly

625					630					635					640	
Gly	Ser	Gly	Gly	Asp	Pro	Thr	Trp	Asp	Pro	Pro	Val	Ala	Thr	Met	Val	
				645					650					655		
Ser	Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	
				660					665					670		
Leu	Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	
				675					680					685		
Glu	Gly	Asp	Ala	Thr	Tyr	Gly	Lys	Leu	Thr	Leu	Lys	Phe	Ile	Cys	Thr	
				690					695					700		
Thr	Gly	Lys	Leu	Pro	Val	Pro	Trp	Pro	Thr	Leu	Val	Thr	Thr	Leu	Ser	
				705					710					715		
Tyr	Gly	Val	Gln	Cys	Phe	Ser	Arg	Tyr	Pro	Asp	His	Met	Lys	Gln	His	
				725					730					735		
Asp	Phe	Phe	Lys	Ser	Ala	Met	Pro	Glu	Gly	Tyr	Val	Gln	Glu	Arg	Thr	
				740					745					750		
Ile	Phe	Phe	Lys	Asp	Asp	Gly	Asn	Tyr	Lys	Thr	Arg	Ala	Glu	Val	Lys	
				755					760					765		
Phe	Glu	Gly	Asp	Thr	Leu	Val	Asn	Arg	Ile	Glu	Leu	Lys	Gly	Ile	Asp	
				770					775					780		
Phe	Lys	Glu	Asp	Gly	Asn	Ile	Leu	Gly	His	Lys	Leu	Glu	Tyr	Asn	Tyr	
				785					790					795		
Asn	Ser	His	Asn	Val	Tyr	Ile	Met	Ala	Asp	Lys	Gln	Lys	Asn	Gly	Ile	
				805					810					815		
Lys	Val	Asn	Phe	Lys	Ile	Arg	His	Asn	Ile	Glu	Asp	Gly	Ser	Val	Gln	
				820					825					830		
Leu	Ala	Asp	His	Tyr	Gln	Gln	Asn	Thr	Pro	Ile	Gly	Asp	Gly	Pro	Val	
				835					840					845		
Leu	Leu	Pro	Asp	Asn	His	Tyr	Leu	Ser	Thr	Gln	Ser	Ala	Leu	Ser	Lys	
				850					855					860		
Asp	Pro	Asn	Glu	Lys	Arg	Asp	His	Met	Val	Leu	Leu	Gly	Phe	Val	Thr	
				865					870					875		
Ala	Ala	Gly	Ile	Thr	Leu	Gly	Met	Asp	Glu	Leu	Tyr	Lys				
				885					890							

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<210> 31
<211> 3399
<212> DNA
<213> Artificial Sequence
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<220>
<223> Fusion between *Aequorea victoria* and human

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<221> CDS
<222> (1)...(3396)
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<400> 31																	
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Met	Glu	Pro	Pro	Thr	Val	Pro	Ser	Glu	Arg	Ser	Leu	Ser	Leu	Ser	Leu		
1				5					10					15			
ccc	ggg	ccc	cgg	gag	ggc	cag	gcc	acc	ctg	aag	cct	ccc	ccg	cag	cac	96	
Pro	Gly	Pro	Arg	Glu	Gly	Gln	Ala	Thr	Leu	Lys	Pro	Pro	Pro	Gln	His		
			20					25					30				
ctg	tgg	cgg	cag	cct	cgg	acc	ccc	atc	cgt	atc	cag	cag	cgc	ggc	tac	144	
Leu	Trp	Arg	Gln	Pro	Arg	Thr	Pro	Ile	Arg	Ile	Gln	Gln	Arg	Gly	Tyr		
		35					40					45					
tcc	gac	agc	gcg	gag	cgc	gcc	gag	cgg	gag	cgg	cag	ccg	cac	cgg	ccc	192	

Ser	Asp	Ser	Ala	Glu	Arg	Ala	Glu	Arg	Glu	Arg	Gln	Pro	His	Arg	Pro	
50						55					60					
ata	gag	cgc	gcc	gat	gcc	atg	gac	acc	agc	gac	cgg	ccc	ggc	ctg	cgc	240
Ile	Glu	Arg	Ala	Asp	Ala	Met	Asp	Thr	Ser	Asp	Arg	Pro	Gly	Leu	Arg	
65					70					75					80	
acg	acc	cgc	atg	tcc	tgg	ccc	tgc	tcc	ttc	cat	ggc	act	ggc	acc	ggc	288
Thr	Thr	Arg	Met	Ser	Trp	Pro	Ser	Ser	Phe	His	Gly	Thr	Gly	Thr	Gly	
				85					90					95		
agc	ggc	ggc	gcg	ggc	gga	ggc	agc	agc	agg	cgc	ttc	gag	gca	gag	aat	336
Ser	Gly	Gly	Ala	Gly	Gly	Gly	Ser	Ser	Arg	Arg	Phe	Glu	Ala	Glu	Asn	
			100					105					110			
ggg	ccg	aca	cca	tct	cct	ggc	cgc	agc	ccc	ctg	gac	tgc	cag	gcg	agc	384
Gly	Pro	Thr	Pro	Ser	Pro	Gly	Arg	Ser	Pro	Leu	Asp	Ser	Gln	Ala	Ser	
			115				120					125				
cca	gga	ctc	gtg	ctg	cac	gcc	ggg	gcg	gcc	acc	agc	cag	cgc	cgg	gag	432
Pro	Gly	Leu	Val	Leu	His	Ala	Gly	Ala	Ala	Thr	Ser	Gln	Arg	Arg	Glu	
			130			135					140					
tcc	ttc	ctg	tac	cgc	tca	gac	agc	gac	tat	gac	atg	tca	ccc	aag	acc	480
Ser	Phe	Leu	Tyr	Arg	Ser	Asp	Ser	Asp	Tyr	Asp	Met	Ser	Pro	Lys	Thr	
145					150					155					160	
atg	tcc	cgg	aac	tca	tgc	gtc	acc	agc	gag	gcg	cac	gct	gaa	gac	ctc	528
Met	Ser	Arg	Asn	Ser	Ser	Val	Thr	Ser	Glu	Ala	His	Ala	Glu	Asp	Leu	
				165					170					175		
atc	gta	aca	cca	ttt	gct	cag	gtg	ctg	gcc	agc	ctc	cgg	agc	gtc	cgt	576
Ile	Val	Thr	Pro	Phe	Ala	Gln	Val	Leu	Ala	Ser	Leu	Arg	Ser	Val	Arg	
			180					185					190			
agc	aac	ttc	tca	ctc	ctg	acc	aat	gtg	ccc	gtt	ccc	agt	aac	aag	cgg	624
Ser	Asn	Phe	Ser	Leu	Leu	Thr	Asn	Val	Pro	Val	Pro	Ser	Asn	Lys	Arg	
			195				200					205				
tcc	ccg	ctg	ggc	ggc	ccc	acc	cct	gtc	tgc	aag	gcc	acg	ctg	tca	gaa	672
Ser	Pro	Leu	Gly	Gly	Pro	Thr	Pro	Val	Cys	Lys	Ala	Thr	Leu	Ser	Glu	
			210			215					220					
gaa	acg	tgt	cag	cag	ttg	gcc	cgg	gag	act	ctg	gag	gag	ctg	gac	tgg	720
Glu	Thr	Cys	Gln	Gln	Leu	Ala	Arg	Glu	Thr	Leu	Glu	Glu	Leu	Asp	Trp	
225					230					235					240	
tgt	ctg	gag	cag	ctg	gag	acc	atg	cag	acc	tat	cgc	tct	gtc	agc	gag	768
Cys	Leu	Glu	Gln	Leu	Glu	Thr	Met	Gln	Thr	Tyr	Arg	Ser	Val	Ser	Glu	
				245					250					255		
atg	gcc	tgc	cac	aag	ttt	aaa	agg	atg	ttg	aac	cgt	gag	ctc	aca	cac	816
Met	Ala	Ser	His	Lys	Phe	Lys	Arg	Met	Leu	Asn	Arg	Glu	Leu	Thr	His	
			260					265					270			
ctg	tca	gaa	atg	agc	agg	tcc	gga	aac	cag	gtc	tca	gag	tac	att	tcc	864
Leu	Ser	Glu	Met	Ser	Arg	Ser	Gly	Asn	Gln	Val	Ser	Glu	Tyr	Ile	Ser	
			275				280						285			

aca aca ttc ctg gac aaa cag aat gaa gtg gag atc cca tca ccc acg Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro Thr 290 295 300	912
atg aag gaa cga gaa aaa cag caa gcg ccg cga cca aga ccc tcc cag Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser Gln 305 310 315 320	960
ccg ccc ccg ccc cct gta cca cac tta cag ccc atg tcc caa atc aca Pro Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile Thr 325 330 335	1008
ggg ttg aaa aag ttg atg cat agt aac agc ctg aac aac tct aac att Gly Leu Lys Lys Leu Met His Ser Asn Ser Leu Asn Asn Ser Asn Ile 340 345 350	1056
ccc cga ttt ggg gtg aag acc gat caa gaa gag ctc ctg gcc caa gaa Pro Arg Phe Gly Val Lys Thr Asp Gln Glu Glu Leu Leu Ala Gln Glu 355 360 365	1104
ctg gag aac ctg aac aag tgg ggc ctg aac atc ttt tgc gtg tcg gat Leu Glu Asn Leu Asn Lys Trp Gly Leu Asn Ile Phe Cys Val Ser Asp 370 375 380	1152
tac gct gga ggc cgc tca ctc acc tgc atc atg tac atg ata ttc cag Tyr Ala Gly Gly Arg Ser Leu Thr Cys Ile Met Tyr Met Ile Phe Gln 385 390 395 400	1200
gag cgg gac ctg ctg aag aaa ttc cgc atc ccg gtg gac acg atg gtg Glu Arg Asp Leu Leu Lys Lys Phe Arg Ile Pro Val Asp Thr Met Val 405 410 415	1248
aca tac atg ctg acg ctg gag gat cac tac cac gct gac gtg gcc tac Thr Tyr Met Leu Thr Leu Glu Asp His Tyr His Ala Asp Val Ala Tyr 420 425 430	1296
cat aac agc ctg cac gca gct gac gtg ctg cag tcc acc cac gta ctg His Asn Ser Leu His Ala Ala Asp Val Leu Gln Ser Thr His Val Leu 435 440 445	1344
ctg gcc acg cct gca cta gat gca gtg ttc acg gac ctg gag att ctc Leu Ala Thr Pro Ala Leu Asp Ala Val Phe Thr Asp Leu Glu Ile Leu 450 455 460	1392
gcc gcc ctc ttc gcg gct gcc atc cac gat gtg gat cac cct ggg gtc Ala Ala Leu Phe Ala Ala Ala Ile His Asp Val Asp His Pro Gly Val 465 470 475 480	1440
tcc aac cag ttc ctc atc aac acc aat tcg gag ctg gcg ctc atg tac Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met Tyr 485 490 495	1488
aac gat gag tcg gtg ctc gag aat cac cac ctg gcc gtg ggc ttc aag Asn Asp Glu Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe Lys 500 505 510	1536
ctg ctg cag gag gac aac tgc gac atc ttc cag aac ctc agc aag cgc Leu Leu Gln Glu Asp Asn Cys Asp Ile Phe Gln Asn Leu Ser Lys Arg 515 520 525	1584

cag cgg cag agc cta cgc aag atg gtc atc gac atg gtg ctg gcc acg Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp Met Val Leu Ala Thr 530 535 540	1632
gac atg tcc aag cac atg acc ctc ctg gct gac ctg aag acc atg gtg Asp Met Ser Lys His Met Thr Leu Leu Ala Asp Leu Lys Thr Met Val 545 550 555 560	1680
gag acc aag aaa gtg acc agc tca ggg gtc ctc ctg cta gat aac tac Glu Thr Lys Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn Tyr 565 570 575	1728
tcc gac cgc atc cag gtc ctc cgg aac atg gtg cac tgt gcc gac ctc Ser Asp Arg Ile Gln Val Leu Arg Asn Met Val His Cys Ala Asp Leu 580 585 590	1776
agc aac ccc acc aag ccg ctg gag ctg tac cgc cag tgg aca gac cgc Ser Asn Pro Thr Lys Pro Leu Glu Leu Tyr Arg Gln Trp Thr Asp Arg 595 600 605	1824
atc atg gcc gag ttc ttc cag cag ggt gac cga gag cgc gag cgt ggc Ile Met Ala Glu Phe Phe Gln Gln Gly Asp Arg Glu Arg Glu Arg Gly 610 615 620	1872
atg gaa atc agc ccc atg tgt gac aag cac act gcc tcc gtg gag aag Met Glu Ile Ser Pro Met Cys Asp Lys His Thr Ala Ser Val Glu Lys 625 630 635 640	1920
tct cag gtg ggt ttt att gac tac att gtg cac cca ttg tgg gag acc Ser Gln Val Gly Phe Ile Asp Tyr Ile Val His Pro Leu Trp Glu Thr 645 650 655	1968
tgg gcg gac ctt gtc cac cca gat gcc cag gag atc ttg gac act ttg Trp Ala Asp Leu Val His Pro Asp Ala Gln Glu Ile Leu Asp Thr Leu 660 665 670	2016
gag gac aac cgg gac tgg tac tac agc gcc atc cgg cag agc cca tct Glu Asp Asn Arg Asp Trp Tyr Tyr Ser Ala Ile Arg Gln Ser Pro Ser 675 680 685	2064
ccg cca ccc gag gag gag tca agg ggg cca ggc cac cca ccc ctg cct Pro Pro Pro Glu Glu Glu Ser Arg Gly Pro Gly His Pro Pro Leu Pro 690 695 700	2112
gac aag ttc cag ttt gag ctg acg ctg gag gag gaa gag gag gaa gaa Asp Lys Phe Gln Phe Glu Leu Thr Leu Glu Glu Glu Glu Glu 705 710 715 720	2160
ata tca atg gcc cag ata ccg tgc aca gcc caa gag gca ttg act gcg Ile Ser Met Ala Gln Ile Pro Cys Thr Ala Gln Glu Ala Leu Thr Ala 725 730 735	2208
cag gga ttg tca gga gtc gag gaa gct ctg gat gca acc ata gcc tgg Gln Gly Leu Ser Gly Val Glu Glu Ala Leu Asp Ala Thr Ile Ala Trp 740 745 750	2256
gag gca tcc ccg gcc cag gag tcg ttg gaa gtt atg gca cag gaa gca Glu Ala Ser Pro Ala Gln Glu Ser Leu Glu Val Met Ala Gln Glu Ala	2304

755	760	765	
tcc ctg gag gcc gag ctg gag gca gtg tat ttg aca cag cag gca cag			2352
Ser Leu Glu Ala Glu Leu Glu Ala Val Tyr Leu Thr Gln Gln Ala Gln			
770	775	780	
tcc aca ggc agt gca cct gtg gct ccg gat gag ttc tcg tcc cgg gag			2400
Ser Thr Gly Ser Ala Pro Val Ala Pro Asp Glu Phe Ser Ser Arg Glu			
785	790	795	800
gaa ttc gtg gtt gct gta agc cac agc agc ccc tct gcc ctg gct ctt			2448
Glu Phe Val Val Ala Val Ser His Ser Ser Pro Ser Ala Leu Ala Leu			
	805	810	815
caa agc ccc ctt ctc cct gct tgg agg acc ctg tct gtt tca gag cat			2496
Gln Ser Pro Leu Leu Pro Ala Trp Arg Thr Leu Ser Val Ser Glu His			
	820	825	830
gcc ccg ggc ctc ccg ggc ctc ccc tcc acg gcg gcc gag gtg gag gcc			2544
Ala Pro Gly Leu Pro Gly Leu Pro Ser Thr Ala Ala Glu Val Glu Ala			
	835	840	845
caa cga gag cac cag gct gcc aag agg gct tgc agt gcc tgc gca ggg			2592
Gln Arg Glu His Gln Ala Ala Lys Arg Ala Cys Ser Ala Cys Ala Gly			
	850	855	860
aca ttt ggg gag gac aca tcc gca ctc cca gct cct ggt ggc ggg ggg			2640
Thr Phe Gly Glu Asp Thr Ser Ala Leu Pro Ala Pro Gly Gly Gly Gly			
865	870	875	880
tca ggt gga gac cct acc tgg gat cca ccg gtc gcc acc atg gtg agc			2688
Ser Gly Gly Asp Pro Thr Trp Asp Pro Pro Val Ala Thr Met Val Ser			
	885	890	895
aag ggc gag gag ctg ttc acc ggg gtg gtg ccc atc ctg gtc gag ctg			2736
Lys Gly Glu Glu Leu Phe Thr Gly Val Val Pro Ile Leu Val Glu Leu			
	900	905	910
gac ggc gac gta aac ggc cac aag ttc agc gtg tcc ggc gag ggc gag			2784
Asp Gly Asp Val Asn Gly His Lys Phe Ser Val Ser Gly Glu Gly Glu			
	915	920	925
ggc gat gcc acc tac ggc aag ctg acc ctg aag ttc atc tgc acc acc			2832
Gly Asp Ala Thr Tyr Gly Lys Leu Thr Leu Lys Phe Ile Cys Thr Thr			
	930	935	940
ggc aag ctg ccc gtg ccc tgg ccc aca cta gtg acc acc ctg tct tac			2880
Gly Lys Leu Pro Val Pro Trp Pro Thr Leu Val Thr Thr Leu Ser Tyr			
945	950	955	960
ggc gtg cag tgc ttc agc cgc tac ccc gac cac atg aag cag cac gac			2928
Gly Val Gln Cys Phe Ser Arg Tyr Pro Asp His Met Lys Gln His Asp			
	965	970	975
ttc ttc aag tcc gcc atg ccc gaa ggc tac gtc cag gag cgc acc atc			2976
Phe Phe Lys Ser Ala Met Pro Glu Gly Tyr Val Gln Glu Arg Thr Ile			
	980	985	990
ttc ttc aag gac gac ggc aac tac aag acc cgc gcc gag gtg aag ttc			3024

Phe Phe Lys Asp Asp Gly Asn Tyr Lys Thr Arg Ala Glu Val Lys Phe
 995 1000 1005
 gag ggc gac acc ctg gtg aac cgc atc gag ctg aag ggc atc gac ttc 3072
 Glu Gly Asp Thr Leu Val Asn Arg Ile Glu Leu Lys Gly Ile Asp Phe
 1010 1015 1020
 aag gag gac ggc aac atc ctg ggg cac aag ctg gag tac aac tac aac 3120
 Lys Glu Asp Gly Asn Ile Leu Gly His Lys Leu Glu Tyr Asn Tyr Asn
 1025 1030 1035 1040
 agc cac aac gtc tat atc atg gcc gac aag cag aag aac ggc atc aag 3168
 Ser His Asn Val Tyr Ile Met Ala Asp Lys Gln Lys Asn Gly Ile Lys
 1045 1050 1055
 gtg aac ttc aag atc cgc cac aac atc gag gac ggc agc gtg cag ctg 3216
 Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu
 1060 1065 1070
 gcc gac cac tac cag cag aac acc ccc atc ggc gac ggc ccc gtg ctg 3264
 Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu
 1075 1080 1085
 ctg ccc gac aac cac tac ctg agc acc cag tcc gcc ctg agc aaa gac 3312
 Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp
 1090 1095 1100
 ccc aac gag aag cgc gat cac atg gtc ctc cta ggg ttc gtg acc gcc 3360
 Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Gly Phe Val Thr Ala
 1105 1110 1115 1120
 gcc ggg atc act ctc ggc atg gac gag ctg tac aag taa 3399
 Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 1125 1130

<210> 32
 <211> 1132
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Fusion between Aequorea victoria and human

<400> 32
 Met Glu Pro Pro Thr Val Pro Ser Glu Arg Ser Leu Ser Leu Ser Leu
 1 5 10 15
 Pro Gly Pro Arg Glu Gly Gln Ala Thr Leu Lys Pro Pro Pro Gln His
 20 25 30
 Leu Trp Arg Gln Pro Arg Thr Pro Ile Arg Ile Gln Gln Arg Gly Tyr
 35 40 45
 Ser Asp Ser Ala Glu Arg Ala Glu Arg Glu Arg Gln Pro His Arg Pro
 50 55 60
 Ile Glu Arg Ala Asp Ala Met Asp Thr Ser Asp Arg Pro Gly Leu Arg
 65 70 75 80
 Thr Thr Arg Met Ser Trp Pro Ser Ser Phe His Gly Thr Gly Thr Gly
 85 90 95
 Ser Gly Gly Ala Gly Gly Gly Ser Ser Arg Arg Phe Glu Ala Glu Asn
 100 105 110

Gly Pro Thr Pro Ser Pro Gly Arg Ser Pro Leu Asp Ser Gln Ala Ser
 115 120 125
 Pro Gly Leu Val Leu His Ala Gly Ala Ala Thr Ser Gln Arg Arg Glu
 130 135 140
 Ser Phe Leu Tyr Arg Ser Asp Ser Asp Tyr Asp Met Ser Pro Lys Thr
 145 150 155 160
 Met Ser Arg Asn Ser Ser Val Thr Ser Glu Ala His Ala Glu Asp Leu
 165 170 175
 Ile Val Thr Pro Phe Ala Gln Val Leu Ala Ser Leu Arg Ser Val Arg
 180 185 190
 Ser Asn Phe Ser Leu Leu Thr Asn Val Pro Val Pro Ser Asn Lys Arg
 195 200 205
 Ser Pro Leu Gly Gly Pro Thr Pro Val Cys Lys Ala Thr Leu Ser Glu
 210 215 220
 Glu Thr Cys Gln Gln Leu Ala Arg Glu Thr Leu Glu Glu Leu Asp Trp
 225 230 235 240
 Cys Leu Glu Gln Leu Glu Thr Met Gln Thr Tyr Arg Ser Val Ser Glu
 245 250 255
 Met Ala Ser His Lys Phe Lys Arg Met Leu Asn Arg Glu Leu Thr His
 260 265 270
 Leu Ser Glu Met Ser Arg Ser Gly Asn Gln Val Ser Glu Tyr Ile Ser
 275 280 285
 Thr Thr Phe Leu Asp Lys Gln Asn Glu Val Glu Ile Pro Ser Pro Thr
 290 295 300
 Met Lys Glu Arg Glu Lys Gln Gln Ala Pro Arg Pro Arg Pro Ser Gln
 305 310 315 320
 Pro Pro Pro Pro Pro Val Pro His Leu Gln Pro Met Ser Gln Ile Thr
 325 330 335
 Gly Leu Lys Lys Leu Met His Ser Asn Ser Leu Asn Asn Ser Asn Ile
 340 345 350
 Pro Arg Phe Gly Val Lys Thr Asp Gln Glu Glu Leu Leu Ala Gln Glu
 355 360 365
 Leu Glu Asn Leu Asn Lys Trp Gly Leu Asn Ile Phe Cys Val Ser Asp
 370 375 380
 Tyr Ala Gly Gly Arg Ser Leu Thr Cys Ile Met Tyr Met Ile Phe Gln
 385 390 395 400
 Glu Arg Asp Leu Leu Lys Lys Phe Arg Ile Pro Val Asp Thr Met Val
 405 410 415
 Thr Tyr Met Leu Thr Leu Glu Asp His Tyr His Ala Asp Val Ala Tyr
 420 425 430
 His Asn Ser Leu His Ala Ala Asp Val Leu Gln Ser Thr His Val Leu
 435 440 445
 Leu Ala Thr Pro Ala Leu Asp Ala Val Phe Thr Asp Leu Glu Ile Leu
 450 455 460
 Ala Ala Leu Phe Ala Ala Ala Ile His Asp Val Asp His Pro Gly Val
 465 470 475 480
 Ser Asn Gln Phe Leu Ile Asn Thr Asn Ser Glu Leu Ala Leu Met Tyr
 485 490 495
 Asn Asp Glu Ser Val Leu Glu Asn His His Leu Ala Val Gly Phe Lys
 500 505 510
 Leu Leu Gln Glu Asp Asn Cys Asp Ile Phe Gln Asn Leu Ser Lys Arg
 515 520 525
 Gln Arg Gln Ser Leu Arg Lys Met Val Ile Asp Met Val Leu Ala Thr
 530 535 540
 Asp Met Ser Lys His Met Thr Leu Leu Ala Asp Leu Lys Thr Met Val
 545 550 555 560
 Glu Thr Lys Lys Val Thr Ser Ser Gly Val Leu Leu Leu Asp Asn Tyr
 565 570 575
 Ser Asp Arg Ile Gln Val Leu Arg Asn Met Val His Cys Ala Asp Leu

				580					585					590		
Ser	Asn	Pro	Thr	Lys	Pro	Leu	Glu	Leu	Tyr	Arg	Gln	Trp	Thr	Asp	Arg	
		595					600					605				
Ile	Met	Ala	Glu	Phe	Phe	Gln	Gln	Gly	Asp	Arg	Glu	Arg	Glu	Arg	Gly	
	610					615					620					
Met	Glu	Ile	Ser	Pro	Met	Cys	Asp	Lys	His	Thr	Ala	Ser	Val	Glu	Lys	
625					630					635					640	
Ser	Gln	Val	Gly	Phe	Ile	Asp	Tyr	Ile	Val	His	Pro	Leu	Trp	Glu	Thr	
				645					650					655		
Trp	Ala	Asp	Leu	Val	His	Pro	Asp	Ala	Gln	Glu	Ile	Leu	Asp	Thr	Leu	
		660					665					670				
Glu	Asp	Asn	Arg	Asp	Trp	Tyr	Tyr	Ser	Ala	Ile	Arg	Gln	Ser	Pro	Ser	
		675					680					685				
Pro	Pro	Pro	Glu	Glu	Glu	Ser	Arg	Gly	Pro	Gly	His	Pro	Pro	Leu	Pro	
		690				695					700					
Asp	Lys	Phe	Gln	Phe	Glu	Leu	Thr	Leu	Glu	Glu	Glu	Glu	Glu	Glu	Glu	
705					710					715					720	
Ile	Ser	Met	Ala	Gln	Ile	Pro	Cys	Thr	Ala	Gln	Glu	Ala	Leu	Thr	Ala	
				725					730					735		
Gln	Gly	Leu	Ser	Gly	Val	Glu	Glu	Ala	Leu	Asp	Ala	Thr	Ile	Ala	Trp	
				740				745					750			
Glu	Ala	Ser	Pro	Ala	Gln	Glu	Ser	Leu	Glu	Val	Met	Ala	Gln	Glu	Ala	
		755					760					765				
Ser	Leu	Glu	Ala	Glu	Leu	Glu	Ala	Val	Tyr	Leu	Thr	Gln	Gln	Ala	Gln	
		770				775					780					
Ser	Thr	Gly	Ser	Ala	Pro	Val	Ala	Pro	Asp	Glu	Phe	Ser	Ser	Arg	Glu	
785					790					795					800	
Glu	Phe	Val	Val	Ala	Val	Ser	His	Ser	Ser	Pro	Ser	Ala	Leu	Ala	Leu	
				805					810					815		
Gln	Ser	Pro	Leu	Leu	Pro	Ala	Trp	Arg	Thr	Leu	Ser	Val	Ser	Glu	His	
			820					825					830			
Ala	Pro	Gly	Leu	Pro	Gly	Leu	Pro	Ser	Thr	Ala	Ala	Glu	Val	Glu	Ala	
		835					840					845				
Gln	Arg	Glu	His	Gln	Ala	Ala	Lys	Arg	Ala	Cys	Ser	Ala	Cys	Ala	Gly	
		850				855					860					
Thr	Phe	Gly	Glu	Asp	Thr	Ser	Ala	Leu	Pro	Ala	Pro	Gly	Gly	Gly	Gly	
865					870					875					880	
Ser	Gly	Gly	Asp	Pro	Thr	Trp	Asp	Pro	Pro	Val	Ala	Thr	Met	Val	Ser	
			885					890					895			
Lys	Gly	Glu	Glu	Leu	Phe	Thr	Gly	Val	Val	Pro	Ile	Leu	Val	Glu	Leu	
			900					905				910				
Asp	Gly	Asp	Val	Asn	Gly	His	Lys	Phe	Ser	Val	Ser	Gly	Glu	Gly	Glu	
		915					920									

Val Asn Phe Lys Ile Arg His Asn Ile Glu Asp Gly Ser Val Gln Leu
 1060 1065 1070
 Ala Asp His Tyr Gln Gln Asn Thr Pro Ile Gly Asp Gly Pro Val Leu
 1075 1080 1085
 Leu Pro Asp Asn His Tyr Leu Ser Thr Gln Ser Ala Leu Ser Lys Asp
 1090 1095 1100
 Pro Asn Glu Lys Arg Asp His Met Val Leu Leu Gly Phe Val Thr Ala
 1105 1110 1115 1120
 Ala Gly Ile Thr Leu Gly Met Asp Glu Leu Tyr Lys
 1125 1130

<210> 33
 <211> 33
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Primer sequence

<400> 33
 tgtactagt accaccctgt cttacggcgt gca 33

<210> 34
 <211> 30
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Primer sequence

<400> 34
 ctgactagt tgggccagg cagggcagc 30

<210> 35
 <211> 42
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Primer sequence

<400> 35
 cccggcggcg gtcacgaacc ctaggaggac catgtgatcg cg 42

<210> 36
 <211> 42
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Primer sequence

<400> 36
 cgcgatcaca tggctcctct agggttcgtg accgccgccg gg 42